

**Trade-off Between Salt Secretion and Gas Exchange by Stomata in
the Leaves of *Glycyrrhiza uralensis***

Peng you Chen, Miao Ma* , Ling yu Shi

Peng you Chen. Ministry of Education Key Laboratory of Xinjiang Phytomedicine Resource Utilization, College of Life Sciences, Shihezi University, Xinjiang 832003, the People's Republic of China. E-mail: 1305352283@qq.com. Tel: 13279933547.

Ling yu Shi. Ministry of Education Key Laboratory of Xinjiang Phytomedicine Resource Utilization, College of Life Sciences, Shihezi University, Xinjiang 832003, the People's Republic of China. E-mail: 1160879462@qq.com. Tel: 15886936395.

Corresponding author: Miao Ma. Ministry of Education Key Laboratory of Xinjiang Phytomedicine Resource Utilization, College of Life Sciences, Shihezi University, Xinjiang 832003, the People's Republic of China. E-mail: mamiaogg@126.com. Tel: 13779213556. Fax: 086-0993-2058500.

Abstract

Our previous study found that stomata in the leaves of *Glycyrrhiza uralensis* (licorice) could secrete salt crystals. In theory, secretion of salt should affect the normal functioning of stomata, thereby affecting the growth and development of *G. uralensis*; however, its populations grow well. We suspect that there may be a trade-off between stomatal salt secretion and gas exchange from the leaves at different positions; therefore, we compared stomatal salt secretion capacity, chlorophyll content, anatomical structure, net photosynthetic rate, and stomatal conductance from the leaves at different positions of licorice. The stomata of the lower leaves exhibited the strongest salt secretion capacity, whereas the stomata of the upper leaves do not secrete any salt. Additionally, the upper and middle leaves had significantly higher chlorophyll content than the lower leaves. The arrangement of mesophyll cells in the upper leaves was the densest, and that in the lower leaves was the least dense. The net photosynthetic rate and stomatal conductance in the upper leaves were the highest, and those in the lower leaves were the lowest. We conclude that the stomata of the upper leaves are mainly used for gas exchange. In contrast, stomata of the lower leaves, showing weak photosynthesis, are responsible for secreting excessive salt to maintain the inner ion balance and ensure normal metabolism in *G. uralensis*.

Introduction

Roots and rhizomes of licorice (*Glycyrrhiza uralensis* Fisch) are Chinese traditional herbs with various pharmacological effects¹, such as alleviating bacteriostasis², cough³ and spasmolysis⁴; combatting viruses⁵; alleviating inflammation and removing toxins⁶; blocking radiation effects⁷ and cancer⁸, making these herbs popular in the pharmaceutical world. Because *G. uralensis* generally grows in calcareous grasslands⁹ and salt-containing lowland meadows, it has been traditionally considered a halophyte. Salinity is an essential environmental factor that affects plant growth and development, and salt stress can directly affect plant growth and development, reduce photosynthetic rates, and inhibit photosynthesis, thereby altering the pigment content of plant leaves and indirectly affecting plant growth¹⁰. To date, although some experimental studies on the salt tolerance of *G. uralensis* seeds have been performed¹¹, the mechanism of adaptation of licorice to salted soil has not yet been reported.

Stomata are the channels through which plants exchange gases with the outside world. The outside CO₂ enters the plant leaves through the stomata, which are also essential for internal water vapor and O₂ elimination. The process of leaf photosynthesis is automatically regulated by stomata, which plays an important role in respiration, water metabolism¹², the immune system¹³, and maintenance of the leaf-surface temperature of plants¹⁴. Therefore, maintaining the normal function of the stomata is vital to the healthy growth and development of plants.

Salt secretion is a critical salt tolerance mechanism in plants. Studies have shown

that salts in plants are generally secreted by salt glands, glandular trichomes, vesicles, and other salt-secreting structures¹⁵. Previous studies have found that salt glands and stomata on the leaves of *G. uralensis* living in high-salt habitats have strong salt secretion capacity, resulting in secretion of large amounts of salts to the leaf surface. Secretion of salt crystals can block the stomata, thereby affecting the normal regulation of stomatal opening, gas exchange between the plant and its external environment, and CO₂ assimilation and water transpiration of the leaves. These effects can increase leaf temperature and decrease photosynthesis-related enzyme activity¹⁶, which may alter the growth and development of *G. uralensis* and the accumulation of secondary metabolites. However, *G. uralensis* populations grow well and do not show any symptoms of injury. Therefore, we hypothesized that there may be a trade-off or balance between gas exchange and salt secretion in the stomata of licorice leaves.

Accordingly, in this study, we aimed to determine this functional trade-off mechanism of the stomata by comparing the abilities of salt secretion and photosynthetic gas exchange by the stomata of licorice leaves placed at different positions on the plant body, to provide evidence for the adaptation mechanisms of the plant to salt-containing soil.

Materials and Methods

Sample area and materials

G. uralensis was sampled from the salinized lowland meadow (47°51' N, 86°51' E) in Altay, Xinjiang, China, which exhibits a temperate continental climate with a long, cold

winter and short, mild summer. Although the *G. uralensis* population in the sample area lives under soil conditions of extreme salinity, the plants grow normally and fulfill their lifecycle successfully. The field experiments started in July 2016.

Electronic conductivity and pH of the soil

The five-point method was used for stratified random sampling of the soil, and each point was divided into three layers: 0–10, 10–20, and 20–30 cm. Soil samples from the same layer were sealed into a bag after being mixed and then brought back quickly to the laboratory for air-drying. The soil samples were then ground and sieved through 1-mm mesh for determination of soil conductivity. Soil conductivity and pH was determined using a DDSJ-308A (Shanghai Tuxin Electronic Technology Co., Ltd.) conductivity meter ($k = 0.964$) in a 1:5 soil/water suspension. The methods for *soil agrochemical analysis* described by Bao, S.D¹⁷ were used to evaluate the grade of soil salinization, and the same soil sample was evaluated three times. The results are presented as averages.

Stomatal salt secretion from the leaves at different positions

Six individuals of *G. uralensis* in an open area were randomly selected and divided into three parts equally, i.e., upper, middle, and lower, according to height (plant height 75 cm). Then, one healthy and mature leaf from each of these three sections was selected randomly, cut into pieces (0.5 cm × 0.5 cm), and placed in FAA solution (70% alcohol:glacial acetic acid:formaldehyde = 18:1:1) for 48 h. Subsequently, the leaf pieces were fixed on the sample table and subjected to ion sputtering gold plating. Samples were then observed with a scanning electron microscope (SU8010, Hitachi High-tech

Company) and photographed. Chemical composition detection of secretions from the stomata by energy dispersive spectrometer (7353, Oxford) to determine whether they are salts. Five fields (600X magnification) were selected randomly for enumeration of the total number of stomata and the number of salt-secreting stomata, and the percentages of salt-secreting stomata were calculated to determine the average.

Chlorophyll content of the leaves

The upper, middle, and lower parts of the same fully-developed, healthy leaves used for microscopic analysis were selected randomly. The main veins were then removed, and the remnants were cut into small pieces and mixed well. Next, 0.25 g of the chopped leaves was accurately weighed and ground into homogenate with 95% ethanol (10 mL) in the dark until the tissue color turned white. The final chlorophyll concentrations were measured according to Callejas, R¹⁸.

Anatomical structure of the leaves

Healthy and fully expanded leaves growing at different positions were selected randomly, cut into small pieces (0.5 cm × 0.5 cm), and placed in FAA solution for 48 h. Transverse sections of the leaves (8-µm thick) were prepared using conventional paraffin sectioning. The sections were stained with safranin and fast green, sealed with optical resin, observed with an Olympus BX51 light microscope, and photographed with an Olympus DP70 system.

Photosynthetic capacity of the leaves

In the open field, six individual *G. uralensis* plants with similar heights and crown widths

were selected, and one healthy and completely expanded leaf was selected. The net photosynthetic rate (P_n) and stomatal conductance (G_s) of the leaves were determined using an LI-6400 portable photosynthesis analyzer (Beijing Ecotek Technology Co., Ltd.) at 2-h intervals. The measurements were done between 08:00 and 20:00 on a sunny, cloudless day, and each leaf was evaluated three times to obtain the average value.

Data analysis

The data were analyzed using SPSS 19.0. The conductivity among different soil layers and differences in chlorophyll, net photosynthetic rate, and stomatal conductance among the upper, middle, and lower leaves were analyzed by one-way analysis of variance. Duncan's multiple range tests were used to detect the significance levels of differences among treatments ($p < 0.05$).

Results

Electronic conductivity and pH value of the soil

The results showed that the conductivity of different soil layers decreased significantly as the soil depth increased. According to the standard in-soil agrochemical analysis method¹⁷, the 0–10 cm soil layer exhibited heavy salinization, and the 10–20 and 20–30 cm layers could also be classified as salinized soil. And the pH value of each soil layer is above 10, so the soil is alkaline (Table 1).

Comparison of stomatal salt secretion among the leaves at different positions

Field observations revealed that salts were secreted by the salt glands and stomata onto the leaf surface of *G. uralensis* (Fig. 1A–F), with significant differences in salt secretion

among the upper, middle, and lower leaves (Fig. 1G–I, Table 2). Among the leaf regions, the upper leaf stomata were permeable, and no salt accumulation was observed (Fig. 1G). In contrast, the stomata of the lower leaves had strong salt secretion capacity, resulting in accumulation of large amounts of salts, with some stomata even being completely blocked by salts (Fig. 1I). Additionally, 78% of the stomata in the middle leaves could secrete salt; however, salt secretion was weak, with only a small amount of salt accumulating in these leaves (Fig. 1H). These results indicate that the salt secretion in *G. uralensis* is mainly achieved by the lower leaves. The results of the energy spectrum analysis show that the secretions discharged from the stomata contain various components (Fig. 2), the metal elements are mainly K and Ca, and the non-metal elements are mainly S and O.

Comparison of chlorophyll content among the leaves at different positions

Chlorophyll content is closely related to plant photosynthesis, with higher chlorophyll content reflecting stronger leaf photosynthetic capacity. Our study showed that the chlorophyll content in the upper and middle leaves of *G. uralensis* was significantly higher than that in the lower leaves, and that there were no significant differences in the chlorophyll content between the upper and middle leaves (Table 3). Thus, the upper and middle leaves of *G. uralensis* in the salinized lowland meadow may have stronger photosynthetic capacity than the lower leaves because of their higher chlorophyll content.

Comparison of anatomical structures of the leaves at different positions

The thicknesses of the leaves at different positions were similar (Fig. 3); however, the differentiation and density of the mesophyll cells differed. The arrangement of mesophyll cells in the upper leaves was the densest; mesophyll cells in the middle leaves are moderate, whereas that in the lower leaves was sparse.

Comparison of photosynthesis among the leaves at different positions

The patterns of diurnal variation in the net photosynthetic rate (P_n) of leaves at different positions were similar, exhibiting a typical bimodal curve and an apparent “photosynthesis lunch break” phenomenon; however, there were significant differences in photosynthetic capacity among the leaves at different positions (Fig. 4). Notably, the P_n of the upper leaves was always the largest, followed by that of the middle leaves, while that of the lower leaves was the smallest. The diurnal variation pattern of stomatal conductance (G_s) in the upper leaves of *G. uralensis* also showed a bimodal curve; the stomatal conductance gradually increased from 08:00, with two peaks at 10:00 and 16:00, consistent with the diurnal variation pattern of the P_n . In contrast, the diurnal variation patterns of the G_s in the middle and lower leaves were typical unimodal curves. At the same time, the G_s of the upper leaves was always the highest, followed by that of the middle leaves, and the G_s of the lower leaves was always the lowest. The G_s values were significantly different among the leaves at different positions.

Discussion

Leaves are the main photosynthetic organs of the plants and the main exchange point of materials and energy between a plant and the outside world. Therefore, the morphology,

structure, and function of leaves are closely related to the surrounding environment and are most sensitive to the habitat. Accordingly, the structural characteristics of leaves can reflect the adaptation of the plant to the environment¹⁹. Salt secretion is one of the most important mechanisms of plant salt tolerance. The salt-secreting structures of plants include glandular hairs, salt glands, vesicles, and the waxy coat of the leaf surfaces¹⁵. However, *G. uralensis* is dependent on a unique structure, the stomata, to secrete salts out of its body. This study can not quantify the salt-secreting degree of stomata due to the existence of other salt-secreting structures such as salt glands (Fig. 1, D). There are many reports on the salt-tolerance mechanisms of *G. uralensis*, which include increasing the content of osmotic adjustment substances, improving the activity of antioxidant enzymes, and accelerating the development of casparian strips on the endodermis of the radicles²⁰, however, the adaptation strategy to high-salt soils induced by secretion of salts from stomata has not yet been found.

At present, it is not unusual for the use of energy dispersive spectrometry (EDS) to analyze the elements of secretions around plant stomata. Such as Pritchard, SG²¹, whose energy spectrum analysis shows that the salt crystals around the stomata of the longleaf pine (*Pinus palustris* Mill.) are mainly calcium sulfate (CaSO₄). Our EDS indicated a composition of Ca, S, K and O (Fig. 2), so the sulfate ion seems to be the major balancing anion in the secretions. Potassium is known to be associated with stomatal movement²² hence the presence of K is not surprising. Calcium is required for the growth of all plants, and is closely related to plant salt tolerance²³. We speculated that secretions

from the stomata of *G. uralensis* may be mainly sulfate, calcium and potassium salts.

Salt stress can lead to physiological drought of plants and affect their morphological structure and physiological functions. In this study, we found that the mesophyll cells in the upper leaves were closely arranged, with well-developed palisade tissues. Additionally, the upper leaves contained the greatest number of chlorophylls, which may explain why the photosynthetic capacity of these leaves was the highest. Mesophyll cells in the lower leaves were arranged loosely, and intercellular spaces were large, consistent with the results of a study by Xie, Z.C²⁴.

Photosynthetic pigments are the material basis for leaf photosynthesis, and the content of chlorophyll reflects, to some extent, the growth status of plants and the photosynthetic capacity of the leaves²⁵. Salt stress enhances the activity of chlorophyllase, resulting in degradation of chlorophyll b and leading to decreased chlorophyll content; these changes also reduce the photosynthetic capacity of the leaves²⁶. In this study, we found that the chlorophyll content in the lower leaves of *G. uralensis* was significantly lower than that in the upper or middle leaves, which may be due to damage to chloroplasts in the lower leaves, which are subjected to severe salt stress because of the large amounts of salt secreted from the stomata on the lower leaves.

As important structures for CO₂ absorption, stomata control the photosynthesis of plants; therefore, variations in stomatal conductance are essential for photosynthesis²⁷. To avoid the influence of these effects on licorice growth due to obstruction of photosynthetic gas exchange caused by salt-blocked stomata, functional differentiation of

the stomata occurs. In the present study, the salt-secreting capacity of the stomata on the leaves of different positions was distinct; the stomata on the lower leaves secreted high levels of salts, resulting in accumulation of larger quantities of salts and leading to stomatal blockage. Previous studies have confirmed that the leaf photosynthesis rate is closely related to stomatal conductance²⁸; thus, the maximum amount of salt secretion by stomata in the lower leaves resulted in lower stomatal conductance and net photosynthetic rate compared with those of the middle and upper leaves. In contrast, the stomatal conductance and net photosynthetic rate of the upper leaves were the highest because the stomata did not participate in salt secretion and remained unobstructed. This indicated a balance between salt secretion and photosynthetic gas exchange in the stomata of the leaves of *G. uralensis* living in a salt-stressed environment, and that the stomatal function of the leaves at different positions on the plant was distinct. Specifically, the stomata of the lower leaves functioned primarily in salt secretion and exhibited the lowest stomatal conductance, whereas the stomatal conductance of the upper leaves was the highest, ensuring regular photosynthetic gas exchange in the licorice plant; the upper leaves did not participate in salt secretion.

In summary, the lower leaves of *G. uralensis* growing in a salt-stressed habitat were relatively older than the upper and middle leaves, and their capacity for photosynthesis was weak. However, the salt secretion capacity of the stomata on the lower leaves was the highest, making these leaves responsible for excreting excess salt and maintaining the internal ion balance. In contrast, the stomata of the young leaves found in the middle and

upper positions appeared to be involved primarily in photosynthetic gas exchange. The major functions of the stomata of the licorice leaves at different positions on the plant body were significantly distinct, demonstrating a balance or trade-off between the two conflicting interests of the stomata as an adaptive strategy for *G. uralensis* living in a salt-stressed habitat. Although we found that *G. uralensis* could adapt to the surrounding high-salt environment through stomatal salt secretion, further studies are needed to explore the mechanisms involved in this process.

Acknowledgement

This study was supported by the National Natural Science Foundation of China (31360047). We would like to thank Editage [www.editage.cn] for English language editing.

References

1. Zhang, W., Lan, F. and Hong, X.X., *Chinese Pharmacopoeia*. China Medical Science and Technology press, Beijing, 2015.
2. Gafner, S. et al., Isoflavonoids and coumarins from *Glycyrrhiza uralensis*: antibacterial activity against oral pathogens and conversion of isoflavans into isoflavan-quinones during purification. *J Nat Prod.*, 2011, **74**, 2514-2519; doi:10.1021/np2004775.
3. Saha, S. et al., Structural features and in vivo antitussive activity of the water extracted polymer from *Glycyrrhiza glabra*. *Int J Biol Macromol.*, 2011, **48**, 634-638; doi:10.1016/j.ijbiomac.2011.02.003.
4. Yazdi, A., Sardari, S. and Sayyah, M., Evaluation of the anticonvulsant activity of the leaves of *Glycyrrhizaglabra* var. *glandulifera* grown in iran, as a possible renewable source for anticonvulsant compounds. *IJPR.*, 2011, **10**, 75.
5. Wang, J., Chen, X.Q. and Wang, W., *Glycyrrhizic* acid as the antiviral component of *Glycyrrhiza uralensis* Fisch. against coxsackievirus A16 and enterovirus 71 of hand foot and mouth disease. *J Ethnopharmacol.*, 2013, **147**, 114-121. doi:10.1016/j.jep.2013.02.017.
6. Kwon, H.J. et al., In vitro anti-rotavirus activity of polyphenol compounds isolated from the roots of *uralensis*. *Bioorgan Med Chem.*, 2010, **18**, 7668-7674.
7. Shetty, T.K., Satav, J.G. and Nair, C.K.K., Protection of DNA and microsomal membranes in vitro by *Glycyrrhiza glabra* L. against gamma irradiation. *Phytother*

- Res.*, 2002, **16**, 576-578, doi:10.1002/ptr.927.
8. Cai, H., Chen X. and Zhang, J.B., 18 β -glycyrrhetic acid inhibits migration and invasion of human gastric cancer cells via the ROS/PKC- α /ERK pathway. *J Nat Med.*, 2017, **72**, 252-259; doi:10.1007/s11418-017-1145-y.
 9. Zhao, K.F. and Feng, L.T., *Chinese Halophyte Resources*. Science Press, Beijing, 2001.
 10. Bhatt, A. and Santo, A., Effects of photoperiod, thermoperiod and salt stress on *Gymnocarpus decandrus* seeds: potential implications in restoration ecology activities. *Botany*, 2017, **95**, 1093-1098.
 11. Pan, Y., Wu, L.J. and Yu, Z.L., Effect of salt and drought stress on antioxidant enzymes activities and SOD isoenzymes of liquorice (*Glycyrrhiza uralensis Fisch.*). *Plant Growth Regul.*, 2006, **49**, 157-165.
 12. Belin, C., Thomine, S. and Schroeder, J., *Water balance and the regulation of stomatal movements*, Springer Netherlands, 2009.
 13. Melotto, M. et al., Plant stomata function in innate immunity against bacterial invasion. *Cell*, 2006, **126**, 969; doi:0.1016/j.cell.2006.06.054.
 14. Liu, M.H. et al., Influence of leaf size of plant on leaf transpiration and temperature in arid regions. *Chin J Plant Ecol.*, 2013, **37**, 436-442; doi:10.3724/SP.J.1258.2013.00045.
 15. Wei, C.X., Wang, J.B. and Chen, Y.F., Epicuticular wax of leaf epidermis: a functional structure for salt excretion in a halophyte *Puccinellia tenuiflora*. *Acta Ecol*

- Sin*, 2004, **24**, 2451-2456.
16. Khan, N.A., Nacl - inhibited chlorophyll synthesis and associated changes in ethylene evolution and antioxidative enzyme activities in wheat. *Biol Plantarum.*, 2003, **47**, 437-440; doi:10.1023/b:biop.0000023890.01126.43.
 17. BAO, S.D., *Soil Agricultural Chemistry Analysis*. China Agricultural Press, Beijing, 2000.
 18. Callejas, R. et al., Evaluation of a non-destructive method to estimate the concentration of chlorophyll in leaves of table grape cv. *Idesia (Arica)*, 2013, **31**, 19-26.
 19. He, N.P. et al., Variation in leaf anatomical traits from tropical to cold-temperate forests and linkage to ecosystem functions. *Funct Ecol.*, 2017, **32**, 10-19; doi:10.1111/1365-2435.12934.
 20. Koster, P. et al., The battle of two ions: Ca²⁺ signalling against Na⁺ stress. *Plant Biol*, 2018; doi:10.1111/plb.12704.
 21. Pritchard, SG. et al., Calcium sulfate deposits associated with needle substomatal cavities of container-grown longleaf pine (*Pinus palustris*) seedlings. *Int J Plant Sci*, 2000, **161**, 917-923.
 22. Michael F. R. et al., How can stomata contribute to salt tolerance? *Ann Bot-London*, 1997, **80**, 387-393.
 23. Hao, J.B., Zhang, F.S. and Tian C.Y., *Halophytes in Xinjiang*. Science Press, Beijing, 2006.
 24. Xie, Z.C., Luo, D. and Zhang, W.J., Effects of silicon on cell microscopic structure

under salt stress of *Glycyrrhiza uralensis*. *J. Chin. Med. rMed.*, 2016, **39**, 2698-2701;
doi:10.13863/j.issn1001-4454.2016.12.006.

25. Jothiramshekar, S. et al., Responses of selected C3 and C4 halophytes to elevated CO₂ concentration under salinity. *Curr. Sci.*, 2018, **115**, 129-135.
26. Mehta, P., Jajoo, A., and Mathur, S., Chlorophyll a fluorescence study revealing effects of high salt stress on Photosystem II in wheat leaves. *Plant Physiol Bioch*, 2010, **48**, 16-20.
27. Joshi, S. C. and Palni, L. M. S., Is dew useful for Himalayan plants? *Curr. Sci.*, 2010, **99**, 1434-1439.
28. Wong, S. C., Cowan, I. R. and Farquhar, G. D., Stomatal conductance correlates with photosynthetic capacity. *Nature*, 1979, **282**, 424-426.

Tables

Table 1. Conductivity and PH value of the soil

Soil layer/cm	Mean value EC ₂₅ /(dS/m)	PH value
0 – 10	8.49 ± 0.03a	10.33 ± 0.03a
10 – 20	3.51 ± 0.02b	10.29 ± 0.03a
20 – 30	3.42 ± 0.01c	10.20 ± 0.02b

Table 2. Comparison of the percentages of salt-secreting stomata among different leaves of *G. uralensis*

Location of leaf on the stem	Percentage of salt-secreting stomata/%	Salt-secreting degree of stomata
Upper leaf	0	Without secreted salt
Middle leaf	78	With a small amount of secreted salt
Lower leaf	100	With a high amount of secreted salt

Table 3. Comparison of chlorophyll content among the leaves at different positions

Leaf position	Chlorophyll a /(mg/g)	Chlorophyll b /(mg/g)	Chlorophyll a + b /(mg/g)
Upper leaf	9.13a	2.609a	11.739a
Middle leaf	9.222a	2.756a	11.978a
Lower leaf	7.45b	1.339b	8.789b

Figure legends

Figure 1. *Glycyrrhiza uralensis* in saline soil and its salt-secreting gland and stomata

GU: *G. uralensis*; L: leaf; S: salt; SA: saline-alkaline soil; SG: salt gland; St: stomata; A: licorice on saline-alkaline soil; B: salt on leaf surface of licorice; C: salt crystals on the leaf surface (5000×); D: salt-secreting glands (3000×); E, F: salt-secreting stomata (10000×); G: stomata on the upper leaf (15000×); H: stomata on the middle leaf (15000×); I: stomata on the lower leaf (15000×).

Figure 2. Energy spectrum analysis of secretions from stomata of *G. uralensis*

Figure 3. Anatomical characteristics of *G. uralensis* leaves at different positions.

A: upper leaf; B: middle leaf; C: lower leaf

a: palisade tissue; b: spongy mesophyll; c: upper epidermis; d: lower epidermis

Figure 4. Comparison of net photosynthetic rate and stomatal conductance of *G. uralensis* leaves at different positions

Figures

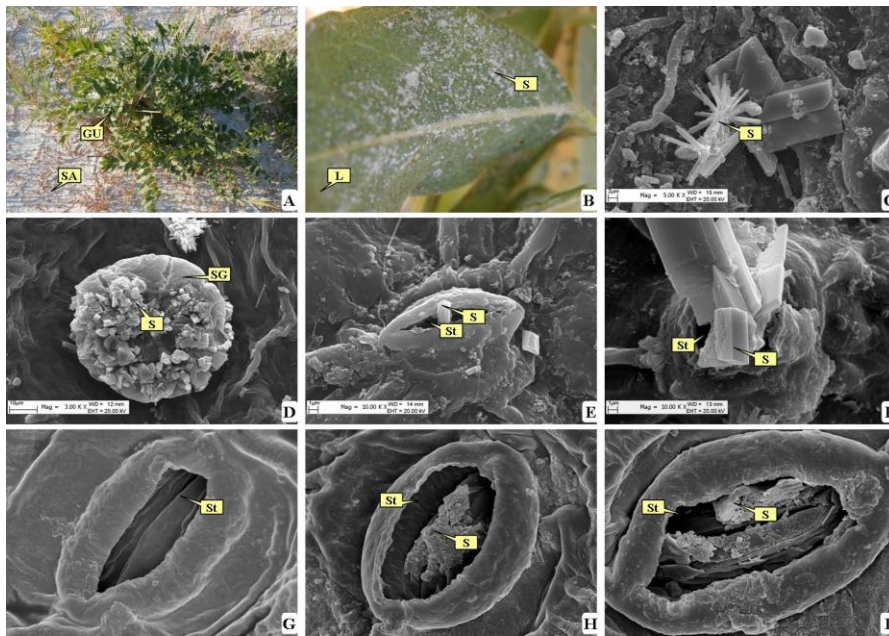


Figure 1

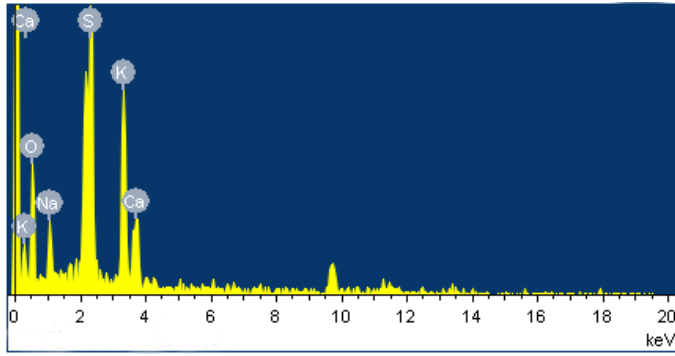


Figure 2

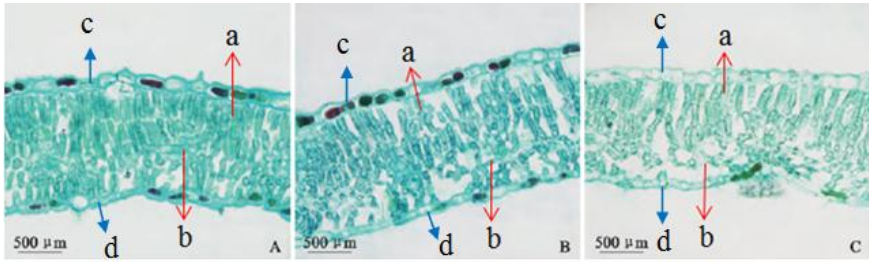


Figure 3

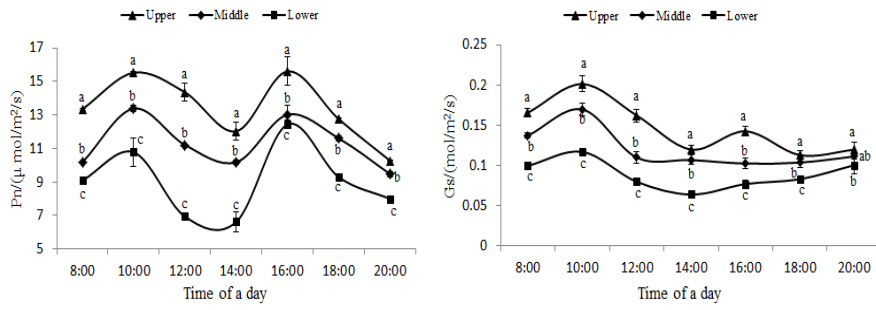


Figure 4