

1 **Rhizosphere-Plant-Microbial system under Polycyclic Aromatic Hydrocarbon**
2 **stress**

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5 **Abstract**

6 Rhizosphere-plant-microbial association is a complex and intricate system susceptible to various
7 organic pollutants including Polycyclic Aromatic Hydrocarbons (PAH). Since soil acts as a sink of
8 PAH, their accumulation not only shifts the delicate rhizosphere-plant-microbe equilibrium, but also
9 enters the food chain through plants. The manner in which the presence of PAH in rhizosphere
10 affects rhizosphere-plant-microbial system is still unclear. This review aims to understand the effects
11 of PAH on rhizosphere-plant-microbial interactions. It also explores the potential use of microbes
12 to alleviate PAH stress in soil for effective and sustainable management.

13 **Keywords:** Bioaccumulation, Microbe-mediated remediation, Persistent Organic Pollutants, Rhizo-
14 microbiota

15 **1. Introduction**

16 PAH are among the most notorious, cosmopolitan, toxic and persistent organic pollutants. They are
17 ubiquitous in nature and in the last five decades, their global burden has increased by around 45%¹.
18 The average global PAH emission has been estimated to be 504 gigagrams/year in 2007 and is
19 majorly contributed by developing Asian countries². Africa, Europe, North America, Oceania and
20 South America contribute 18.8%, 9.5%, 8.0%, 1.5% and 6.0%, respectively to worldwide PAH
21 emissions³. The distribution and accumulation of PAH are not uniform and show significant regional
22 variation. India (90 gigagram) and China (114 gigagram) are the major contributors of PAH
23 annually⁴. This sharp rise is due to rapid industrialization, fossil-fuel-dependent transportation and
24 other anthropogenic activities. Sources of PAH can be broadly classified as petrogenic (petroleum
25 as the source) and pyrogenic (incomplete combustion of fossil fuels and biomass). Most of the PAH
26 are potential carcinogens and their exposure or indirect consumption leads to serious health effects.
27 They are considered as priority pollutants due to their 'three disease-causing' effects (carcinogenic,

28 teratogenic and mutagenic) on humans⁵. In spite of being volatile and present in air, their presence
29 in soil is of concern as they tend to accumulate in it.

30 Due to their hydrophobicity, around 90% of the global PAH is adsorbed in soil which not only
31 makes it a major sink but also drastically changes the physicochemical properties of soil, thereby
32 altering its structure and function⁶. Rhizosphere, the zone of interaction between soil and roots, is a
33 diverse and intricate habitat hosting a wide range of algae, protozoa, arthropods, nematodes, bacteria
34 and fungi. Among these, microorganisms play a significant role in regulating essential
35 biogeochemical processes and also facilitate xenobiotic degradation. Biodegradation of soil
36 pollutants such as pesticides, insecticides and PAH is a critical ecosystem service provided by
37 rhizospheric microbes⁷. The presence of PAH in rhizosphere enhances their risk of entering the food
38 chain as well as influences soil microbial diversity and dynamics⁸.

39 Plants are key players that influence the entry, accumulation and movement of PAH in the food
40 chain. They take up PAH either directly through roots or indirectly through the cuticle⁹. Higher the
41 PAH concentration in soil, greater is the risk of their bioaccumulation and subsequent magnification
42 in the plant. Excess PAH accumulation in plants is responsible for acute, chronic as well as latent
43 injuries, that eventually disrupt its primary metabolic functions such as photosynthesis¹⁰. As plants
44 regulate the growth and diversity of microbial communities through root exudates, when grown in
45 high PAH the diversity of rhizospheric microbes as well as their endophytes is observed to be
46 modified¹¹. Therefore, the interaction between soil microbes, plants and PAH is a tripartite process
47 and any change in one significantly affects not only the other two but also the food chain.

48 The interaction between rhizosphere-plant-microbe is delicate and sensitive to the presence of PAH.
49 However, no study has addressed the effect of PAH toxicity on this complex-yet important system.
50 This review summarizes the multifaceted interaction between rhizosphere-plant-microbes and
51 analyses, how this system responds to increased PAH levels. It also explores microbe-mediated,

52 sustainable mitigation measures for PAH polluted soil, to prevent their accumulation and
53 magnification in food chain.

54 **2. Soil: A sink of PAH**

55 PAH released from various anthropogenic activities accumulate in soil by wet or dry deposition.
56 Soil serves as their primary and largest steady repository. Their lipophilicity makes them less
57 available for biodegradation and hence, they accumulate in soil increasing their persistence in the
58 ecosystem¹². The distribution of PAH in the rhizosphere depends on their source and chemical
59 properties, soil characteristics and environmental conditions¹². The amount of PAH in soil is
60 inversely proportional to its proximity to the emission source. Industrial and urban soils have more
61 PAH contamination as compared to rural or remote areas¹³. Chemical properties of PAH also
62 influence their bioavailability. Low molecular weight (LMW) PAH with two or three rings such as
63 naphthalene, fluorene, phenanthrene and anthracene are present in negligible amounts as they
64 undergo degradation, leaching and volatilization in the soil¹⁴. In contrast, high molecular weight
65 (HMW) PAH such as pyrene, chrysene, benzo(*a*)pyrene and benzo(*a*)anthracene are abundant in
66 soil as they are not degraded or transformed completely into simpler products due to their highly
67 lipophilic, persistent and stable chemical composition¹⁵. It is known that 2-,3-,4-, 5- and 6-ring PAH
68 contribute to >3%, >16%, 45%, >27% and >6% of the total PAH respectively in the soil of urban
69 parks in China¹⁶. Thus, HMW PAH are the majorly responsible for soil contamination.

70 Based on the concentration of PAH, soil is classified as (i) unpolluted (<600 ngg⁻¹); (ii) slightly
71 polluted (600-1000 ngg⁻¹); (iii) polluted (1,000-5000 ngg⁻¹); (iv) heavily polluted (5000-10,000
72 ngg⁻¹) and (v) very high polluted (>10,000 ngg⁻¹)^{6, 17}. Figure 1 summarizes the country-wise
73 prevalence of PAH in soil based on the literature.

74 **3. Significance and role of rhizospheric microbes**

75 **3.1 Diversity of microbes**

76 Rhizosphere is a dynamic zone where numerous biochemical and biophysical processes take place
77 that shapes and organizes soil's physical and functional attributes. Soil characteristics such as
78 texture, pore volume and particle aggregation influence root-microbe associations. Interactions
79 between soil and its microbiome are influenced by plant species and soil type, and are important for
80 plant growth and matter turnover¹⁸.

81 Bacteria are among the most abundant microbes in the rhizosphere, covering nearly 15% of the total
82 root surface, dominated by phyla such as Proteobacteria, Actinobacteria, Firmicutes, Bacteroidetes
83 and Acidobacteria¹⁹. Proteobacteria are present in greater numbers due to their ability to act on labile
84 carbon sources and thus, grow at a fast rate making them capable of surviving in diverse
85 rhizospheres²⁰. Another dominant phylum is Acidobacteria that plays a key role in carbon cycling
86 as it is capable of degrading cellulose and lignin²¹. Anaerobic bacteria belonging to *Latescibacteria*
87 *and Planctomycetes*) dominate the rhizosphere whereas aerobic bacteria (*Parcubacteria, Firmicutes*
88 *and Saccharibacteria*) are comparatively lesser in number due to reduced oxygen levels in the
89 rhizosphere²².

90 Along with bacteria, rhizosphere is well represented by fungi primarily belonging to the
91 Ascomycota, Basidiomycota and Zygomycota²³. Arbuscular mycorrhizal (AM) fungi are among the
92 most ubiquitously present in the rhizosphere ecosystem globally and provide an array of services^{5,25}.
93 Moreover, several algae including species of *Arthrospira, Chlorella, Dunaliella, Nostoc* and
94 *Aphanizomenon* are found in the rhizosphere that play a significant role in maintaining soil health
95 by acting as both bioprotective and bio-stimulant agents²⁶.

96 Rhizosphere microbiome is sensitive and quickly responds to any change in its surrounding²⁴. The
97 physio-chemical properties of soil, type of vegetation and stage of the plant influence the diversity
98 and abundance of microbes in rhizosphere.

99 **3.2 Rhizosphere Microbes in Ecosystem Services**

100 Rhizosphere microbes play a substantial role in various ecological services such as soil formation,
101 decomposition of organic matter, biogeochemical cycles and degradation of soil contaminants²⁷.
102 The ecosystem services performed by microbes are categorized into regulating, supporting and
103 provisioning services and are summarized in Supplementary Table 1.
104 Soil bioremediation is an important regulating service performed by the rhizosphere microbes.
105 Genera of bacteria including *Acinetobacter*, *Burkholderia*, *Pseudomonas*, *Flavobacterium*, *Bacillus*,
106 *Azotobacter*, and fungi such as *Aspergillus* are capable of degrading an array of soil
107 contaminants^{28,29}. Many species of mycorrhizal fungi are also involved in remediating soil pollutants
108 such as organic hydrocarbons³⁰. Microbes either use the organic pollutants as the carbon source, or
109 degrade the toxic and complex compounds into simpler and lesser toxic forms³¹. Their efficiency
110 and strategy are dependent on pollution load. A consortium of PAH degrading microbes removed
111 97.2% of pyrene from soil and inhibited its accumulation in host tissues³². Similarly, up to 90% of
112 biodegradation of lindane (organochlorine pesticide) using *Paracoccus* sp. NITDBR1 has been
113 reported³³. Rhizospheric microbes are also known to efficiently degrade diesel-based contaminants.
114 Diesel oil-contaminated soil was effectively mediated by an artificial consortium
115 containing *Alcaligenes xylosoxidans*, *Pseudomonas fluorescens*, *Pseudomonas*
116 *putida*, *Stenotrophomonas maltophilia* and *Xanthomonas* sp³⁴.

117 **4. Plant–microbes–rhizosphere system under PAH stress**

118 **4.1 Physicochemical properties of soil**

119 Soil across the globe is under constant threat of PAH which tend to accumulate on its particles. The
120 degree of adsorption and desorption of these pollutants depends on redox conditions, pH, organic
121 ligands and inorganic ions³⁵.

122 The molecular weight and ring structure of PAH govern their fate in the soil as well as influence soil
123 characteristics^{36,38}. The HMW tend to alter the physical characters of soil such as pore size, rate of
124 filtration and aeration as they bind more firmly to the particles as compared to LMW PAH. Fine soil

125 particles have lesser inter-particle space and a porosity-mediated effect that restricts the mobility of
126 HMW PAH due to their greater hydrophobicity³⁷. As a result, they tend to completely choke soil
127 pores or partially replace them leading to reduced aeration and filtration capacity of soil.

128 Presence of PAH, affects physico-chemical properties of soil such as pH, CEC, humic acid, carbon
129 and nitrogen content^{39,40}. PAH also influence physical characteristics of soil such as grain size,
130 water-holding capacity and porosity, leading to reduced aeration and choking of soil pores³⁷. Fine
131 grained particles i.e. silt and clay are more susceptible to PAH binding⁴¹.

132 Alternately, properties of soil such as its size, pH and associated organic carbon, also determine the
133 extent of PAH effect³⁸. Fine soil particles provide lesser scope of PAH movement resulting in
134 persistent toxicity and enduring effects³⁷. The carbon-rich organic and fine-grained acidic soil is
135 more susceptible to PAH contamination and is therefore difficult to bioremediate⁴². Prolonged
136 exposure and subsequent accumulation of PAH in the soil and by extension rhizosphere,
137 significantly lowers its quality and adversely affects the rate of seed germination, growth and
138 eventually plant health⁴³.

139 In spite of their ubiquitous presence across soil in different parts of the world, there is limited
140 information on the influence of PAH on soils and its processes. There is a large temporal and spatial
141 variability of types and concentration of PAH, and therefore more region-specific studies are
142 required to provide sustainable remediation solutions.

143 **4.2 Impact on rhizo-microbiota**

144 Rhizosphere microbes are highly sensitive to any change in the physico-chemical or edaphic
145 properties of soil. Under the influence of pollutants, a shift in the diversity and growth patterns of
146 the microbial population takes place. Biological activities in the rhizosphere such as microbial
147 biomass and associated enzymatic activities are susceptible to organic pollutants including PAH⁴⁴.
148 The type of PAH influence community profile of rhizospheric microbiota. Studies have been focused

149 on understanding the effect of PAH on microbial diversity, density and metabolic activity⁴⁵. At high
150 concentration, PAH are detrimental to their growth or even toxic to soil microbes. Contamination of
151 PAH in rhizosphere may even decrease the diversity of some microbial populations and increase the
152 density of aromatic ring dioxygenase-expressing bacteria (ARDB) due to an increase in
153 concentration of organic carbon¹¹. This could be the result of increased root exudation or
154 accumulation of PAH around the roots due to its transport processes inside PAH accumulated plants.
155 Soil microflora is highly sensitive to any soil perturbation and therefore known to be pollution
156 indicators^{46,47}. PAH stress in soil can lead to altered microbial diversity, activity and succession⁴⁸.
157 Influence on microbiome depends upon the type and molecular weight of the PAH⁴⁹. For example,
158 pyrene contaminated soils affected diversity and abundance of bacteria more than of fungi.
159 Similarly, the density of gram-negative bacteria and AM Fungi increased around *Echinacea*
160 *purpurea* grown in PAH-contaminated soil⁵⁰. PAH mediated change in microbiome is due to altered
161 carbon and nitrogen cycling, and accelerated metabolism of carbon in soil⁵¹. These studies indicate
162 that PAH may negatively affect the ecological balance of the rhizosphere. High PAH contamination
163 lowers the capacity of soil to degrade contaminants due to decreased microbial activity^{42,51}. Any
164 change in soil microbial diversity and population dynamics may alter the delicate balance of nutrient
165 cycling. Moreover, any shift in rhizospheric community structure and function has direct
166 implications on the growth of plants as well.

167 Soil enzymes are sensitive indicators of function and degradation potential of soil. Enzymes such as
168 urease, alkaline phosphatase, polyphenol oxidase and dehydrogenase are used to determine PAH
169 load through an assessment of the rhizo-microbiota⁵³. There is a positive correlation between the
170 presence of different PAH and the activity of dehydrogenase and urease⁵⁴.

171 **4.3 Plant response to PAH stress**

172 Increasing PAH concentration in soil is of concern due to their tendency to bioaccumulate and
173 subsequently bio-magnify through absorption by plants. They not only affect germinability, growth,
174 physiology and metabolic behaviour of plants, but also resource partitioning^{57,58}. Exposure to low
175 PAH concentration increased plant weight and reduced root area⁵⁵. However, high PAH
176 contamination in soil may completely inhibit plant growth and have phytotoxic effects, making them
177 prone to other stressors^{56,59}. Growth in high PAH soil leads to deformed trichomes, chlorosis, white
178 spots, impaired root growth and development⁵⁴. PAH even have the ability to penetrate cell
179 membranes and eventually decrease water content, nutrient utilization, inhibit photosynthetic
180 activity and electron transport in plants⁵⁸. LMW PAH even can cause phytotoxic effect in plants
181 such as impaired growth and development of plants⁵⁹.

182 Uptake of PAH by plants is governed by their concentration, water solubility, physicochemical state
183 and even the soil type³⁶. Additionally, their molecular weight is key determinant of their uptake.
184 HMW PAH binds more firmly to soil particles making their removal through physical means a
185 challenge⁶¹. PAH uptake and accumulation in plant, primarily a passive process, is limited due to
186 their high partition coefficients in soil^{62,63}. Root and shoot concentration factor, and transpiration
187 stream concentration factor also determine their uptake and availability in plants⁶⁴.

188 Rhizosphere significantly influences PAH uptake by plants through direct and indirect
189 influences^{63,65}. Active rhizosphere facilitates PAH degradation, and decreases its bioavailability^{66,67}.
190 Factors such as microflora, root exudates, soil type, nutrients available and pollutant load influence
191 the rhizosphere effect⁶⁸. Aromatic compounds that are homologous to PAH as well as surfactant
192 molecules released by plant roots generates rhizosphere effect that might contribute to an increase
193 in microbial activity and rhizodegradation of PAH in contaminated soil^{69,70}.

194 Exposure to PAH induces oxidative stress in plants leading to increased levels of reactive oxygen
195 species (ROS)⁷¹. *Arabidopsis thaliana* exposed to high phenanthrene concentration leads to an
196 increase in ROS load which may surpass the capacity of the plant's antioxidant systems⁷².

197 A significant amount of PAH is accumulated in leaves, especially in cuticle and is dependent on
198 morphological factors of leaf such as size, surface to volume ratio and wax content, as well as
199 chemical properties such as lipophilicity and volatility of PAH⁷³. Adsorption and intake of PAH not
200 only increase toxicity in plants owing to bioaccumulation, but also make them available in
201 vegetables and fruits⁷⁴. Temperature is yet another important factor that determines accumulation of
202 PAH in plants. High-temperature conditions enable PAH to exist in gas phase and low-temperature
203 condition facilitate its accumulation in leaves.

204 The effect of PAH on plant growth and metabolism is species-specific. For example, the biomass of
205 *Aeschynomene indica* increased whereas, the biomass of *Panicum bisulcatum* decreased to
206 approximately 70% when both species were applied with same the PAH concentration^{68,75}. They not
207 only affect the growth and metabolism of plants, but also their endophyte population. Plant
208 endophytes are highly sensitive to PAH contamination and result in an increase in specific PAH
209 degrading endophytes in them. Therefore, endophytes can be explored for better PAH
210 biodegradation in an eco-friendly manner.

211 **4.4 PAH stress on the rhizosphere-plant-microbial relationship**

212 Active interaction between plants and rhizo-microbiota has been long recognized and is the fulcrum
213 of biological activity in the rhizosphere. Microbes are dependent on plant exudates and other
214 associated rhizodeposits for energy, and in turn facilitates critical processes in nutrient cycling,
215 production of growth promoters and xenobiotic degradation. Plants under abiotic stress such as
216 contaminants influence soil properties through root exudates⁷⁶. This, in turn, affects the
217 rhizosphere's microbial communities and eventually alters plant performance. This is known as
218 plant-soil feedback mechanism. Thus, rhizosphere-plant-microbes share an intimate relationship
219 which not only symbiotic but is more of a cyclic relationship, wherein what happens in one has a
220 direct bearing on the other, and ultimately affects all the three components. The health of rhizosphere

221 and its microbial populations are important for plant health and productivity. Plants and rhizospheric
222 microbes are a combined unit of the soil ecosystem, neither of them can be studied and considered
223 separately.

224 Rhizosphere microbiome have the capability to degrade pollutants such as PAH⁷⁷. Mere presence of
225 rhizosphere accelerates the rate of PAH dissipation and degradation^{75,78}. Interaction between plant-
226 soil-microbiome is greatly influenced by plant species, physiology and photosystems, and life
227 stages^{79,80}. Rhizosphere of grasses provide better degradation of PAH due to their fibrous root
228 systems⁸¹. However, in a comparative study, legumes were better in removal of PAH when compared
229 to grasses⁷⁵.

230 This three-component rhizosphere-plant-microbial system quickly responds to any change in the
231 physico-chemical properties and composition of soil. The addition of carbon-based pollutants such
232 as PAH significantly influence all three components. The effect of PAH on this system is not
233 restricted to morpho-physiological changes in plants, and its influence can be seen on plant-microbe
234 signalling, nutrient allocation and resource partitioning⁸². Increased carbon allocation to roots and
235 rapid utilization by microbes in *Phragmite australis* under hydrocarbon stress⁸³. Studies have shown
236 that plant hormones such as ethylene, Brassinosteroids, ethane, expansin, cytokinin, cytochrome
237 P450, glutathione-S-transferase and endogenous abscisic acid play a significant role in the signalling
238 pathway of PAH induced stress⁸⁴. Glutathione S-transferase and ABC transporter play key role in
239 transfer of PAH from rhizosphere into plant tissue⁸⁵. PAH can affect the plant-rhizosphere-microbial
240 interaction is a two-way process (Figure 2). PAH accumulated in plants affect its microbiota and
241 physiology. This in turn, affect the lipophilic composition of root exudates, which facilitate PAH
242 degradation, the presence of glucose, pyruvate and acetate increase PAH accumulation in
243 rhizosphere. Presence of such compounds in root exudates in rhizosphere lowered the expression of
244 PAH degrading genes such as nahG in *Pseudomonas fluorescens* HK44 present in rhizosphere

245 resulting in increased bacterial biomass⁸⁶. Therefore, direct or indirect effect of PAH on plants,
246 microbes and rhizosphere may alter their natural interactions temporarily or permanently.

247 **5. Significance of plant-microbe-soil in alleviating PAH stress**

248 Due to their high concentration and prevalence, particularly in some parts of the world, PAH make
249 way into the food chain, augmenting at each trophic level. Their effective and eco-friendly
250 mitigation is thus, the need of the hour. PAH removal through chemical and photolytic oxidation,
251 volatilization and sedimentation are costly and environmentally unsustainable³¹. Conversely,
252 phytoremediation and microbial remediation, although cost-effective and ecologically, sustainable
253 and are dependent on the physicochemical properties of soil, microbial profile and plant
254 communities present in the rhizosphere environment and may show incomplete and slow
255 degradation rates. Till now, studies have focused on microbe-assisted phytoremediation which uses
256 either a single microbe or a consortium for remediation. However, the mechanism of action of
257 interact a single microbe or consortia within the rhizosphere is not completely understood.
258 Moreover, the rhizospheric ecosystem of higher plants has unique eco-biological characteristics
259 which can be altered due to addition of such microbe(s). It may also be difficult to provide enough
260 microbial population for complete biodegradation of the PAH.

261 Rhizoremediation is the use of plant and rhizospheric microbiota for the efficient degradation of
262 PAH. The application of Plant Growth Promoting Rhizobacteria (PGPR) provides bidirectional
263 benefit by efficiently degrading PAH and improving plant growth and development, which in turn
264 help rhizospheric microbes to grow and survive in the rhizosphere⁸⁷. Various studies have confirmed
265 that degradation of PAH through rhizoremediation is far more efficient than other remediation
266 techniques due to the exploitation of useful interactions of the host plant and their respective rhizo-
267 microbiota. For example, >70% of pyrene degradation was documented in vegetated soil compared

268 to that in the unplanted soil conditions (which was <40%) indicating that rhizo-microbiota along
269 with host plant accelerated the degradation process⁸⁸.

270 Rhizo-microbiota of specific host plant may include microbes which are significant in plant growth
271 and development, and a subpopulation possessing PAH degrading genes such as ARDB. Such type
272 of healthy rhizo-microbiota may be created either naturally through microbial interaction within
273 rhizosphere or artificially via microbiome engineering. The latter includes establishment of more
274 efficient microbiome artificially into the rhizosphere which can degrade PAH completely. *In-situ*
275 microbiome engineering may be achieved through augmentation (elevating the levels of specific
276 microbial community via supplementing certain type of supplements), reduction (establishment of
277 non-conducive environment for undesirable microbial function) or bio-inoculation (application of
278 mixture of microbes involved in degradation)⁸⁹. Plant-rhizosphere-microbial interactions need to be
279 explored for better remediation of PAH-contaminated soil. Some of the beneficial interactions are
280 summarized in Table 1.

281 **6. Conclusion and future prospects**

282 Prolonged exposure of soil to high PAH levels increases the risk of their bioaccumulation and
283 magnification in food chain. PAH stress adversely affects the plant-soil-microbial interaction,
284 ultimately disrupting several of the ecosystem functions and services. To come up with possible
285 solution, it is imperative that this tripartite interaction is completely understood. Being a dynamic
286 system, it is in a constant state of flux and shows several regional and local variation. Since most of
287 PAH emissions are anthropogenic and associated with economic development, there seems to be no
288 imminent near future solution for this menace. To reduce toxic effects of PAH and minimize the
289 damage to ecosystem and health, sustainable solutions are required. Tapping the potential of soil
290 microbes which can efficiently degrade PAH seems to be the best possible sustainable solution.
291 Rhizoremediation is a promising tool in establishing a healthy host plant microbial zone and creating

292 an efficient PAH degradation system. However, diversity of rhizosphere microbes involved in
293 rhizoremediation and their functional genes are still not well understood. Exploring modern
294 molecular technologies such as genomics, metabolomics, transcriptomics and proteomics for
295 understanding the biochemistry of PAH degrading microbes will provide ample knowledge of
296 potential genes, enzymes involved in degradation process. This will lead to development of
297 genetically modified microbe or consortia and provide effective and sustainable solution for soil
298 remediation from PAH stress.

299

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304 **Conflict of interest**

305 The authors declare that they have no conflict of interest.

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317 **References**

- 318 1. Chen, X. et al., Past, present, and future perspectives on the assessment of
319 bioavailability/bioaccessibility of polycyclic aromatic hydrocarbons: A 20-year systemic review
320 based on scientific econometrics. *Sc. of the Total Environ.*, 2021, **774**, 145585.
- 321 2. Shen, H. et al., Global atmospheric emissions of polycyclic aromatic hydrocarbons from 1960 to
322 2008 and future predictions. *Environ. Sci. Technol.*, 2013, **47**, 6415-6424.
- 323 3. Zhang, Y. and Tao, S., Global atmospheric emission inventory of polycyclic aromatic hydrocarbons
324 (PAHs) for 2004. *Atmos. Environ.*, 2009, **43**, 812-819.
- 325 4. Kumar, A., Ambade, B., Sankar, T. K., Sethi, S. S., and Kurwadkar, S., Source identification and
326 health risk assessment of atmospheric PM_{2.5}-bound polycyclic aromatic hydrocarbons in
327 Jamshedpur, India. *Sustainable cities and society*, 2020, **52**, 101801.
- 328 5. Liu, R., Dai, Y., and Sun, L., Effect of rhizosphere enzymes on phytoremediation in PAH-
329 contaminated soil using five plant species. *PLoS One*, 2015, **10**, e0120369.
- 330 6. Patel, A. B., Shaikh, S., Jain, K. R., Desai, C., and Madamwar, D., Polycyclic Aromatic
331 Hydrocarbons: Sources, Toxicity, and Remediation Approaches. *Front. Microbiol.*, 2020, **11**,
332 562813.
- 333 7. Kuppusamy, S., Thavamani, P., Venkateswarlu, K., Lee, Y. B., Naidu, R., and Megharaj, M.,
334 Remediation approaches for polycyclic aromatic hydrocarbons (PAHs) contaminated soils:
335 Technological constraints, emerging trends and future directions. *Chemosphere*, 2017, **168**, 944-
336 968.
- 337 8. Cristaldi, A., Conti, G. O., Jho, E. H., Zuccarello, P., Grasso, A., Copat, C., and Ferrante, M.,
338 Phytoremediation of contaminated soils by heavy metals and PAHs. A brief review. *Environ.*
339 *Technol. Innov.*, 2017, **8**, 309-326.

- 340 9. Fismes, J., Perrin-Ganier, C., Empereur-Bissonnet, P., and Morel, J. L., Soil-to-Root Transfer and
341 Translocation of Polycyclic Aromatic Hydrocarbons by Vegetables Grown on Industrial
342 Contaminated Soils. *J. Environ. Qual.*, 2002, **31**, 1649-1656.
- 343 10. Oguntimehin, I., Eissa, F., and Sakugawa, H., Negative effects of fluoranthene on the
344 ecophysiology of tomato plants (*Lycopersicon esculentum* Mill). Fluoranthene mists negatively
345 affected tomato plants. *Chemosphere*, 2010, **78**, 877-884.
- 346 11. Liste, H. H. and Prutz, I., Plant performance, dioxygenase-expressing rhizosphere bacteria, and
347 biodegradation of weathered hydrocarbons in contaminated soil. *Chemosphere*, 2006, **62**, 1411-
348 1420.
- 349 12. Kumar, M. et al., Remediation of soils and sediments polluted with polycyclic aromatic
350 hydrocarbons: To immobilize, mobilize, or degrade? *J. Hazard. Mater.*, 2021, **420**, 126534.
- 351 13. Krauss, M., and Wilcke, W., Polychlorinated naphthalenes in urban soils: analysis,
352 concentrations, and relation to other persistent organic pollutants. *Environ. Pol.*, 2003,**12**, 75-89.
- 353 14. Hussain, K., Hoque, R. R., Balachandran, S., Medhi, S., Idris, M. G., Rahman, M., and Hussain,
354 F. L., Monitoring and risk analysis of PAHs in the environment. In *Handbook of Environ. Mater.*
355 *Management* (ed. Hussain C. M.), Springer International Publishing, 2018, pp.1-35.
- 356 15. Kanaly, R. A. and Harayama, S., Biodegradation of high-molecular-weight polycyclic aromatic
357 hydrocarbons by bacteria. *J. Bacteriol.*, 2000, **182**, 2059-2067.
- 358 16. Qu, Y. et al., Potential sources, influencing factors, and health risks of polycyclic aromatic
359 hydrocarbons (PAHs) in the surface soil of urban parks in Beijing, China. *Environ. Pol.*, 2020, **260**,
360 114016.
- 361 17. Duan, Y. et al., Characteristics of polycyclic aromatic hydrocarbons in agricultural soils at a
362 typical coke production base in Shanxi, China. *Chemosphere*, 2015, **127**, 64-69.

- 363 18. Berg, G. and Smalla, K., Plant species and soil type cooperatively shape the structure and
364 function of microbial communities in the rhizosphere. *FEMS Microbiol. Ecol.*, 2009, **68**, 1-13.
- 365 19. van Loon, L. C., Plant responses to plant growth-promoting rhizobacteria. In *New perspectives*
366 *and approaches in plant growth*, Springer, Dordrecht, 2007, 243-254.
- 367 20. Lagos, L., Maruyama, F., Nannipieri, P., Mora, M. L., Ogram, A., and Jorquera, M. A. et al.,
368 Current overview on the study of bacteria in the rhizosphere by modern molecular techniques: A
369 mini-review. *J. Soil Sci. Plant Nutr.*, 2015, **15**, 504-523.
- 370 21. Ward, N. L. et al., Three genomes from the phylum Acidobacteria provide insight into the
371 lifestyles of these microorganisms in soils. *Appl. Environ. Microbiol.*, 2009, **75**, 2046-2056.
- 372 22. Liu, F., Mo, X., Kong, W., and Song, Y., Soil bacterial diversity, structure, and function of
373 *Suaeda salsa* in rhizosphere and non-rhizosphere soils in various habitats in the Yellow River Delta,
374 China. *Sc. of the Total Environ.*, 2020, **740**, 140144
- 375 23. Qiao, Q. et al., Characterization and variation of the rhizosphere fungal community structure of
376 cultivated tetraploid cotton. *PLoS One*, 2019, **14**, e0207903.
- 377 24. Storey, S., Ashaari, M. M., Clipson, N., Doyle, E., and De Menezes, A. B., Opportunistic bacteria
378 dominate the soil microbiome response to phenanthrene in a microcosm-based study. *Front.*
379 *Microbiol.*, 2018, **9**.
- 380 25. Gai, J. P., Christie, P., Feng, G., and Li, X. L., Twenty years of research on community
381 composition and species distribution of arbuscular mycorrhizal fungi in China: A review.
382 *Mycorrhiza*, 2006, **16**, 229-239.
- 383 26. Lee, S. M. and Ryu, C. M., Algae as New Kids in the Beneficial Plant Microbiome. *Front. Plant*
384 *Sci.*, 2021, **12**, 599742.

- 385 27. Singh, M., Awasthi, A., Soni, S. K., Singh, R., Verma, R. K., and Kalra, A., Complementarity
386 among plant growth promoting traits in rhizospheric bacterial communities promotes plant growth.
387 *Sci. Rep.*, 2015, **5**, 1-8.
- 388 28. Gupta, A., Gupta, R., and Singh, R. L., Microbes and Environment. In *Principles and Application*
389 *of Environ. Biotechnol. for a Sustain. Future*, Springer, Singapore, 2017, pp. 43-84.
- 390 29. Eilers, K. G., Lauber, C. L., Knight, R., and Fierer, N., Shifts in bacterial community structure
391 associated with inputs of low molecular weight carbon compounds to soil. *Soil Biol. Biochem.*, 2010,
392 **42**, 896-903.
- 393 30. dos Santos, J. J. and Maranhão, L. T., Rhizospheric microorganisms as a solution for the recovery
394 of soils contaminated by petroleum: A review. *J. Environ. Manage.*, 2018, **210**, 104-113.
- 395 31. Haritash, A. K. and Kaushik, C. P., Biodegradation aspects of Polycyclic Aromatic Hydrocarbons
396 (PAHs): A review. *J. Hazard. Mater.*, 2009, **169**, 1-15.
- 397 32. Singha, L. P. and Pandey, P., Rhizobacterial community of *Jatropha curcas* associated with pyrene
398 biodegradation by consortium of PAH-degrading bacteria. *Appl. Soil Ecol.*, 2020, **155**, 103685.
- 399 33. Sahoo, B., Ningthoujam, R., and Chaudhuri, S., Isolation and characterization of a
400 lindane degrading bacteria *Paracoccus* sp. NITDBR1 and evaluation of its plant growth promoting
401 traits. *Int. Microbiol.*, 2019, **22**, 155-167.
- 402 34. Szulc, A., Ambrożewicz, D., Sydow, M., Ławniczak, Ł., Piotrowska-Cyplik, A., Marecik, R., and
403 Chrzanowski, Ł. The influence of bioaugmentation and biosurfactant addition on bioremediation
404 efficiency of diesel-oil contaminated soil: Feasibility during field studies. *J. Environ. Manage.*,
405 2014, **132**, 121-128.

- 406 35. Biswas, B., Qi, F., Biswas, J. K., Wijayawardena, A., Khan, M. A. I., and Naidu, R., The fate of
407 chemical pollutants with soil properties and processes in the climate change paradigm—a review.
408 *Soil Syst.*, 2018, **2**, 51.
- 409 36. Abdel-Shafy, H. I., and Mansour, M. S., A review on polycyclic aromatic hydrocarbons: source,
410 environmental impact, effect on human health and remediation. *Egyptian J. of petroleum*, 2016, **25**,
411 107-123.
- 412 37. Singh, S. K., and Haritash, A. K., Polycyclic aromatic hydrocarbons: soil pollution and
413 remediation. *Int. J. of Environ. Sci. and Technol.*, 2019, **16**, 6489-6512.
- 414 38. Ahangar, A. G., Sorption of PAHs in the Soil Environment with Emphasis on the Role of Soil
415 Organic Matter: A Review. *World Appl. Sci. J.*, 2010, **11**, 759-765.
- 416 39. Irha, N., Slet, J., and Petersell, V., Effect of heavy metals and PAH on soil assessed via
417 dehydrogenase assay. *Environ. Int.*, 2003, **28**, 779-782.
- 418 40. Włóka, D., Kacprzak, M., Grobelak, A., Grosser, A., and Napora, A., The Impact of PAHs
419 Contamination on the Physicochemical Properties and Microbiological Activity of Industrial
420 Soils. *Polycycl. Aromat. Compd.*, 2015, **35**, 372-386.
- 421 41. Magi, E., Bianco, R., Ianni, C., and di Carro, M., Distribution of polycyclic aromatic hydrocarbons
422 in the sediments of the Adriatic Sea. *Environ. Pol.*, 2002, **119**, 91-98.
- 423 42. Maletić, S. P., Beljin, J. M., Rončević, S. D., Grgić, M. G., and Dalmacija, B. D., State of the art
424 and future challenges for polycyclic aromatic hydrocarbons in sediments: sources, fate,
425 bioavailability and remediation techniques. *J. of Hazard. Mater.*, 2019, **365**, 467-482.
- 426 43. Tang, J. C., Wang, R. G., Niu, X. W., Wang, M., Chu, H. R., and Zhou, Q. X., Characterisation
427 of the rhizoremediation of petroleum-contaminated soil: Effect of different influencing factors.
428 *Biogeosciences*, 2010, **7**, 3961-3969.

- 429 44. Labud, V., Garcia, C., and Hernandez, T., Effect of hydrocarbon pollution on the microbial
430 properties of a sandy and a clay soil. *Chemosphere*, 2007, **66**, 1863-1871.
- 431 45. Li, S., Hu, S., Shi, S., Ren, L., Yan, W., and Zhao, H., Microbial diversity and metaproteomic
432 analysis of activated sludge responses to naphthalene and anthracene exposure. *RSC Adv.*, 2019, **9**,
433 22841-22852.
- 434 46. Maliszewska-Kordybach, B., Klimkiewicz-Pawlas, A., Smreczak, B., and Janusauskaite, D.,
435 Ecotoxic Effect of Phenanthrene on Nitrifying Bacteria in Soils of Different Properties. *J.*
436 *Environ. Qual.*, 2007, **36**, 1635-1645.
- 437 47. Hou, J., Xu, X., Yu, H., Xi, B., and Tan, W., Comparing the long-term responses of soil
438 microbial structures and diversities to polyethylene microplastics in different aggregate fractions.
439 *Environ. Int.*, 2021, **149**, 106398.
- 440 48. Wu, Y., Zeng, J., Zhu, Q., Zhang, Z., and Lin, X., PH is the primary determinant of the bacterial
441 community structure in agricultural soils impacted by polycyclic aromatic hydrocarbon pollution.
442 *Sci. Rep.*, 2017, **7**, 40093.
- 443 49. Yi, M., Zhang, L., Li, Y., and Qian, Y., Structural, metabolic, and functional characteristics of
444 soil microbial communities in response to benzo[a]pyrene stress. *J. Hazard. Mater.*, 2022, **431**.
- 445 50. Liu, K. et al., Response of Rhizosphere Microbial Community in High-PAH-Contaminated Soil
446 Using *Echinacea purpurea* (L.) Moench. *Appl. Sci. (Switzerland)*, 2022, **12**, 2973.
- 447 51. Zhang, L., Yi, M., and Lu, P., Effects of pyrene on the structure and metabolic function of soil
448 microbial communities. *Environ. Pol.*, 2022, 119301.
- 449 52. Mueller, K. E., and Shann, J. R., PAH dissipation in spiked soil: impacts of bioavailability,
450 microbial activity, and trees. *Chemosphere*, 2006, **64**, 1006-1014.

- 451 53. Margesin, R., Determination of Enzyme Activities in Contaminated Soil. In *Monitoring and*
452 *Assessing Soil Bioremediation*, Springer, Berlin, Heidelberg, 2005, pp. 309-320.
- 453 54. Shen, G., Lu, Y., Zhou, Q., and Hong, J., Interaction of polycyclic aromatic hydrocarbons and
454 heavy metals on soil enzyme. *Chemosphere*, 2005, **61**, 1175-1182.
- 455 55. Calabrese, E. J. and Blain, R. B., Hormesis and plant biology. *Environ. Pol.*, 2009, **157**, 42-48.
- 456 56. Chaîneau, C. H., Morel, J. L., and Oudot, J., Phytotoxicity and Plant Uptake of Fuel Oil
457 Hydrocarbons. *J. Environ. Qual.*, 1997, **26**, 1478-1483.
- 458 57. Maliszewska-Kordybach, B., Klimkiewicz-Pawlas, A., Smreczak, B., and Stuczyński, T.,
459 Relationship between soil concentrations of PAHs and their regional emission indices. *Water Air*
460 *Soil Pol.*, 2010, **213**, 319-330.
- 461 58. Kreslavski, V. D. et al., Effects of polyaromatic hydrocarbons on photosystem II activity in pea
462 leaves. *Plant Physiol. and Biochem.*, 2014, **81**, 135-142.
- 463 59. Henner, P., Schiavon, M., Druelle, V., and Lichtfouse, E., Phytotoxicity of ancient gaswork soils.
464 Effect of polycyclic aromatic hydrocarbons (PAHs) on plant germination. In *Org. Geochem.* 1999,
465 **30**, 963-969.
- 466 60. Smreczak, B., and Maliszewska-Kordybach, B., Seeds germination and root growth of selected
467 plants in PAH contaminated soil. *Fresenius Environ. Bulletin*, 2003, **12**, 946-949.
- 468 61. García-Alonso, S., Pérez-Pastor, R. M., Sevillano-Castaño, M. L., Escolano, O., and García-
469 Frutos, F. J., Influence of particle size on the quality of pah concentration measurements in a
470 contaminated soil. *Polycycl. Aromat. Compd.*, 2008, **28**, 67-83.
- 471 62. Trapp, S., Kästner, M., Adam, I. K. U., Rein, A., and Gosewinkel, K., Methods for improvement
472 of PAH degradation by substrate amendments, solvent and phytoremediation. *Project Molecular*

- 473 *Approaches and Metagenomic Investigations for Optimizing Clean-up of PAH-contaminated Sites*
474 *(MagicPAH), EU FP, 2014, 7.*
- 475 63. Su, Y. H. and Zhu, Y. G., Uptake of selected PAHs from contaminated soils by rice seedlings
476 (*Oryza sativa*) and influence of rhizosphere on PAH distribution. *Environ. Pol.*, 2008, **155**, 359-
477 365.
- 478 64. Briggs, G. G., Bromilow, R. H., and Evans, A. A., Relationships between lipophilicity and root
479 uptake and translocation of non-ionised chemicals by barley. *Pestic. Sci.*, 1982, **13**, 495-504.
- 480 65. Marchal, G., Smith, K. E. C., Mayer, P., Wollesen De Jonge, L., and Karlson, U. G., Impact of
481 soil amendments and the plant rhizosphere on PAH behaviour in soil. *Environ. Pol.*, 2014, **188**,
482 124-131.
- 483 66. Pignatello, J. J., and Li, J., bioavailability of PAHs to native soil bacteria promoted by nutrient
484 addition. In *AGU Fall Meeting Abstracts*, 2006, **2006**, pp. H33I-05.
- 485 67. Fu, P. P., Xia, Q., Sun, X., and Yu, H., Phototoxicity and environmental transformation of
486 polycyclic aromatic hydrocarbons (PAHs)-light-induced reactive oxygen species, lipid
487 peroxidation, and DNA damage. *J. Environ. Sci. Health C. Environ. Carcinog. Ecotoxicol. Rev.*,
488 2012, **30**, 1-41.
- 489 68. Ma, B., He, Y., Chen, H. hai, Xu, J. ming, and Rengel, Z., Dissipation of polycyclic aromatic
490 hydrocarbons (PAHs) in the rhizosphere: Synthesis through meta-analysis. *Environ. Pol.*, 2010, **158**,
491 855-861.
- 492 69. Singer, A. C., Crowley, D. E., and Thompson, I. P., Secondary plant metabolites in
493 phytoremediation and biotransformation. *Trends Biotechnol.*, 2003, 123-130.

- 494 70. Goel, G., Pandey, P., Sood, A., Bisht, S., Maheshwari, D. K., and Sharma, G. D., Transformation
495 of pWWO in *Rhizobium leguminosarum* DPT to Engineer Toluene Degrading Ability for
496 Rhizoremediation. *Indian J. Microbiol.*, 2012, **52**, 197-202.
- 497 71. Shen, Y., Li, J., Gu, R., Yue, L., Wang, H., Zhan, X., and Xing, B., Carotenoid and superoxide
498 dismutase are the most effective antioxidants participating in ROS scavenging in phenanthrene
499 accumulated wheat leaf. *Chemosphere*, 2018, **197**, 513-525.
- 500 72. Liu, H., Weisman, D., Ye, Y. B., Cui, B., Huang, Y. H., Colón-Carmona, A., and Wang, Z. H., An
501 oxidative stress response to polycyclic aromatic hydrocarbon exposure is rapid and complex in
502 *Arabidopsis thaliana*. *Plant Sci.*, 2009, **176**, 375-382.
- 503 73. Mukhopadhyay, S., Dutta, R., and Das, P., A critical review on plant biomonitors for determination
504 of polycyclic aromatic hydrocarbons (PAHs) in air through solvent extraction techniques.
505 *Chemosphere*, 2020, **251**, 126441.
- 506 74. Singh, L. and Agarwal, T., Polycyclic aromatic hydrocarbons in diet: Concern for public health.
507 *Trends Food Sci. Technol.*, 2018, **79**, 160-170.
- 508 75. Lee, S. H., Lee, W. S., Lee, C. H., and Kim, J. G., Degradation of phenanthrene and pyrene in
509 rhizosphere of grasses and legumes. *J. Hazard. Mater.*, 2008, **153**, 892-898.
- 510 76. Rohrbacher, F. and St-Arnaud, M., Root exudation: The ecological driver of hydrocarbon
511 rhizoremediation. *Agronomy*, 2016, **6**, 19.
- 512 77. Muratova, A., Hübner, T., Tischer, S., Turkovskaya, O., Möder, M., and Kuschik, P., Plant -
513 Rhizosphere-microflora association during phytoremediation of PAH-contaminated soil. *Int. J.*
514 *Phytoremediation*, 2003, **5**, 137-151.

- 515 78. Liu, S., Luo, Y., Cao, Z., Wu, L., and Wong, M., Effect of ryegrass (*Lolium multiflorum* L.) growth
516 on degradation of benzo[a]pyrene and enzyme activity in soil. *J. Food Agric. Environ.*, 2013, **11**,
517 247-253
- 518 79. Bouasria, A., Mustafa, T., De Bello, F., et al., Changes in root-associated microbial communities
519 are determined by species-specific plant growth responses to stress and disturbance. *Eur. J. Soil*
520 *Biol.*, 2012, **52**, 59-66.
- 521 80. D'Orazio, V., Ghanem, A., and Senesi, N., Phytoremediation of pyrene contaminated soils by
522 different plant species. *Clean (Weinh)*, 2013, **41**, 377-382.
- 523 81. Thomas, F. and Cébron, A., Short-term rhizosphere effect on available carbon sources,
524 phenanthrene degradation, and active microbiome in an aged-contaminated industrial soil. *Front.*
525 *Microbiol.*, 2016, **7**, 92.
- 526 82. Xiang, L., Harindintwali, J. D., Wang, F., et al., Integrating biochar, bacteria, and plants for
527 sustainable remediation of soils contaminated with organic pollutants. *Environ. Sci. &*
528 *Technol.*, 2022, **56**, 16546-16566.
- 529 83. Nie, M., Yang, Q., Jiang, L. F., Fang, C. M., Chen, J. K., and Li, B., Do plants modulate biomass
530 allocation in response to petroleum pollution? *Biol. Lett.*, 2010, **6**, 811-814.
- 531 84. Zhao, C. et al., Regulation of endogenous phytohormones alters the fluoranthene content in
532 *Arabidopsis thaliana*. *Sci. of the Total Environ.*, 2019, **688**, 935-943.
- 533 85. Ahammed, G. J. et al., Brassinosteroids induce plant tolerance against phenanthrene by enhancing
534 degradation and detoxification in *Solanum lycopersicum* L. *Ecotoxicol. and Environ. Safety*, 2012,
535 **80**, 28-36.

- 536 86. Kamath, R., Schnoor, J. L., and Alvarez, P. J. J., Effect of Root-Derived Substrates on the
537 Expression of nah-lux Genes in *Pseudomonas fluorescens* HK44: Implications for PAH
538 Biodegradation in the Rhizosphere. *Environ. Sci. Technol.*, 2004, **38**, 1740-1745.
- 539 87. Azaizeh, H., Castro, P. M. L., and Kidd, P., Biodegradation of Organic Xenobiotic Pollutants in
540 the Rhizosphere. *Organic Xenobiotics and Plants: From Mode of Action to Ecophysiology*, 2011,
541 191-2015.
- 542 88. Liste, H. H. and Alexander, M., Accumulation of phenanthrene and pyrene in rhizosphere soil.
543 *Chemosphere*, 2000, **40**, 11-14.
- 544 89. Kotoky, R., Rajkumari, J., and Pandey, P., The rhizosphere microbiome: Significance in
545 rhizoremediation of polyaromatic hydrocarbon contaminated soil. *J. Environ. Manage.*, 2018, **217**,
546 858-870.
- 547 90. Dominguez, J. J. A., Bacosa, H. P., Chien, M. F., and Inoue, C., Enhanced degradation of polycyclic
548 aromatic hydrocarbons (PAHs) in the rhizosphere of sudangrass (*Sorghum × drummondii*).
549 *Chemosphere*, 2019, **234**, 789-795.
- 550 91. Zhao, X., Miao, R., Guo, M., and Zhou, Y., Effects of Fire Phoenix (a genotype mixture of *Festuca*
551 *arundinacea* L.) and *Mycobacterium* sp. on the degradation of PAHs and bacterial community in
552 soil. *Environ. Sci. and Pol. Res.*, 2021, **28**, 25692-25700.
- 553 92. Kong, F. Xin, Sun, G. Dong, and Liu, Z. Pei, Degradation of polycyclic aromatic hydrocarbons in
554 soil mesocosms by microbial/plant bioaugmentation: Performance and mechanism. *Chemosphere*,
555 2018, **198**, 83-91.
- 556 93. Bisht, S. et al., Utilization of endophytic strain *Bacillus* sp. SBER3 for biodegradation of
557 polyaromatic hydrocarbons (PAH) in soil model system. *Eur. J. Soil Biol.*, 2014, **60**, 67-76.

- 558 94. Bisht, S., Pandey, P., Kaur, G., Aggarwal, H., Sood, A., Sharma, S., Kumar, V. and Bisht, N.S.,
559 Biodegradation of naphthalene and anthracene by chemo-tactically active rhizobacteria of *Populus*
560 *deltoides*. *Brazilian J. of Microbiol.*, 2010, **41**, 922-930.
- 561 95. Muratova, A., Dubrovskaya, E., Golubev, S., Grinev, V., Chernyshova, M., and Turkovskaya, O.,
562 The coupling of the plant and microbial catabolisms of phenanthrene in the rhizosphere of *Medicago*
563 *sativa*. *J. Plant Physiol.*, 2015, **188**, 1-10.
- 564 96. Yu, X. Z., Wu, S. C., Wu, F. Y., and Wong, M. H., Enhanced dissipation of PAHs from soil using
565 mycorrhizal ryegrass and PAH-degrading bacteria. *J. Hazard. Mater.*, 2011, **186**, 1206-1217.
- 566 97. Li, W. et al., Combination of plant-growth-promoting and fluoranthene-degrading microbes
567 enhances phytoremediation efficiency in the ryegrass rhizosphere. *Environmental Sci. and Pol.*
568 *Res.*, 2021, **28**, 6068-6077.
- 569 98. Zhang, J., Lin, X., Liu, W., Wang, Y., Zeng, J., and Chen, H., Effect of organic wastes on the plant-
570 microbe remediation for removal of aged PAHs in soils. *J. Environ Sci. (China)*, 2012, **24**, 1476-
571 1482.
- 572 99. Dhote, M., Kumar, A., Jajoo, A., and Juwarkar, A., Assessment of hydrocarbon degradation
573 potentials in a plant-microbe interaction system with oil sludge contamination: A sustainable
574 solution. *Int. J. Phytoremediation*, 2017, **19**, 1085-1092.
- 575 100. Wołejko, E., Jabłońska-Trypuć, A., Wydro, U., Butarewicz, A., and Łozowicka, B., Soil biological
576 activity as an indicator of soil pollution with pesticides – A review. *Appl. Soil Ecol.*, 2020, **147**,
577 103356.
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- 580

581 **Table 1:** Plant-microbial system for effective PAH degradation

S. No.	Plant-microbial System	PAH degraded	Percentage degradation	Genes involved	Reference
1.	<i>Sorghum x drummondii</i> and Sphingomonadales	Fluorene, Phenanthrene, Fluoranthene and Pyrene	98%	PAH- <i>RHD</i> α and <i>nidA</i>	90
2.	Fire Phoenix and <i>Mycobacterium</i> sp	Total PAH	40.3–53.7%	-	91
3.	<i>Orychopragmus violaceus</i> and <i>Rhodococcus ruber</i> Em1	Total PAH	17.85%	<i>alkB</i> and PAH- <i>RHD</i>	92
4.	<u><i>Populus deltoides</i></u> and <i>Bacillus</i> sp. SBER3	Anthracene and Naphthalene	>75%	-	93
5.	<i>Populus deltoides</i> and <i>Kurthia</i> sp., <i>Micrococcus varians</i> <i>Deinococcus radiodurans</i> and <i>Bacillus circulans</i>	Anthracene and Naphthalene	>85%		94

6.	Alfalfa (<i>Medicago sativa</i> L.) and <i>Ensifer meliloti</i> , <i>Pseudomonas kunmingensis</i> , <i>Rhizobium petrolearium</i> and <i>Stenotrophomonas</i> sp.	Phenanthrene	20-60%	-	95
7.	<i>Jatropha curcas</i> and <i>Pseudomonas aeruginosa</i> PDB1, <i>Pseudomonas fragi</i> DBC, <i>Klebsiella pneumoniae</i> AWD5, <i>Alcaligenes faecalis</i> BDB4 and <i>Acinetobacter</i> sp. PDB4	Pyrene	97.2%	<i>nod</i> , <i>nahR</i> , <i>nahAF</i> , <i>catA</i> , <i>kshA</i> , <i>hsaC</i>	100
8.	Ryegrass and <i>Acinetobacter</i> sp. or AMF	Phenanthrene and Pyrene	>90%		96
9.	Ryegrass (<i>Lolium multiflorum</i>) and <i>Arthrobacter pascei</i> strain (ZZ21) and/or <i>Bacillus cereus</i> strain (Z21)	Fluoranthene	74.9%		97
10.	Alfalfa (<i>Medicago sativa</i> L.) and <i>Bacillus</i> s and <i>Flavobacterium</i> sp.	Total PAH	25.8%		98
11.	<i>Vertiveria zizanioides</i> and <i>Bacillus</i> and <i>Pseudomonas</i> sp.	Total PAH	88-89%		99

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584 **Figure Legends**

585 **Figure 1:** Country-wise PAH contamination in soil.

586 Supplementary Table 2 titled “Country-wise distribution of PAH” with corresponding references has been attached as supplementary information.

587 The map has been prepared using mapchart.net.

588 **Figure 2:** Rhizosphere-plant-microbe system under PAH stress

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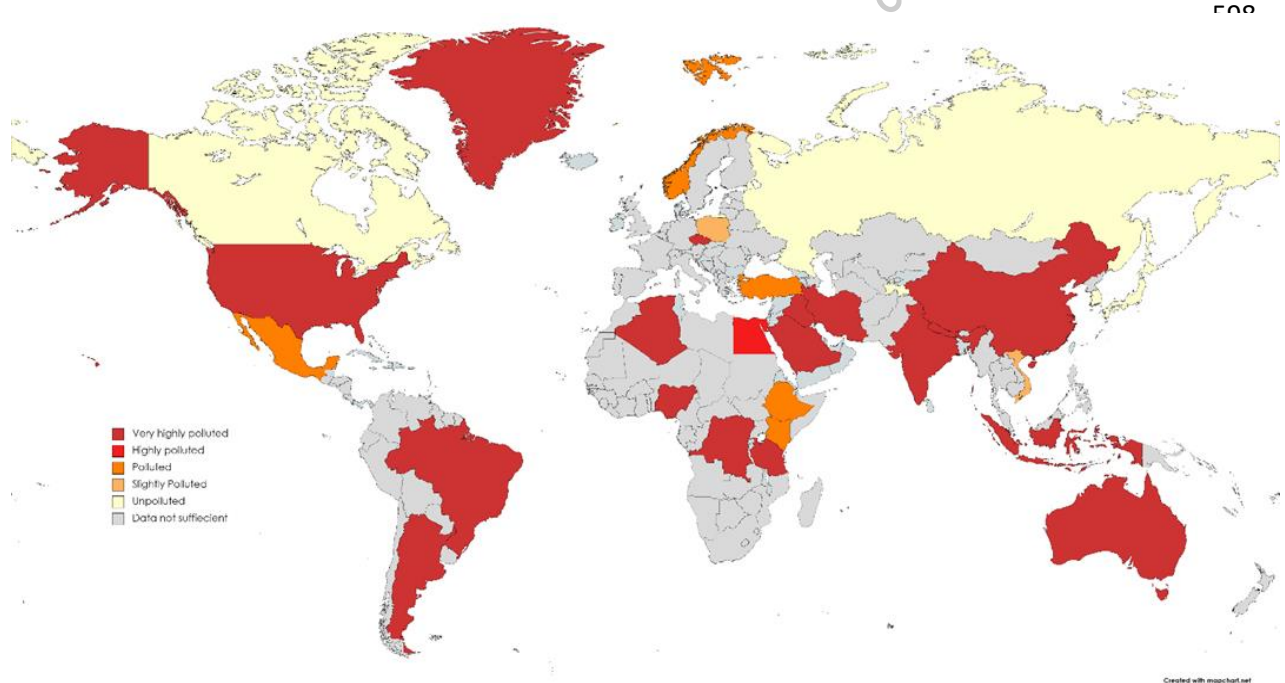
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608 Figure 1

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