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2 ***Sāncipāt*: A Popular Writing-base of Manuscript of Early Assam**

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8 *Sāncipāt* was a popular manuscript writing-base of early and medieval Assam. Tens of thousands  
9 of *Sāncipāt* manuscripts still exist in Assam, some of them centuries old, without fading of ink and  
10 miniature painting in spite of its harsh hot and humid climate. Traditionally, *Sāncipāt* was made  
11 of bark of *Sānci* tree through an arduous procedure. Here we report a scientific study of  
12 physicochemical properties of traditionally prepared model and old *Sāncipāt* folios at different  
13 stages of preparation using FT-IR, XRD and SEM-EDX spectra, weight loss during degumming,  
14 tensile strength, gloss index and antifungal properties. Comparison of data with freshly prepared  
15 *Sāncipāt* showed intact internal structure and strength in the old *Sāncipāt*. An antifungal property  
16 observed in *Sāncipāt* is attributed to *Tutia* used during degumming of the bark, and two pigments,  
17 *viz.*, *Hengul* and *Hāitāl* applied as thin coating and border, respectively. Partial degumming,  
18 coating with fatty pulse, application of *Hāitāl* and *Hengul*, and repeated pressing, smoothening  
19 and drying together impart strength and glossiness to *Sāncipāt*.

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21 **Keywords:** *Sāncipāt* manuscripts, *Hengul-Hāitāl*, medieval writing-base, antifungal activity,  
22 degumming

23

## 24 **Introduction**

25 India possesses more than five million ancient manuscripts, making it the largest repository of  
26 manuscript wealth in the world<sup>1,2</sup>. These manuscripts are treasures of information for research of  
27 Indian intellectual heritage and need adequate conservation<sup>3</sup>. Scientific study of manuscripts and  
28 manuscript-making traditions may unravel interesting traditional knowledge and immensely help  
29 in their conservation. Different types of writing-bases were used in ancient and medieval world<sup>4</sup>  
30 <sup>7</sup>, such as papyrus in ancient Egypt and medieval Europe, animal skin in medieval Europe, hand-  
31 made paper in China. *Tālapatra* (Palm leaf) in south and southeast Asia including and *Bhurjapatra*  
32 (bark of *Betula utilis*, Himalayan Birch) in north India. Interestingly, Assam, a northeastern state  
33 of India, had a rich heritage of manuscript writing on a unique type of writing base, *Sāncipāt*, made  
34 of bark of *Sānci* tree (*Aquilaria malaccensis* Lamk. syn. *A. agallocha* Roxb.), during early and  
35 medieval period (Figure 1)<sup>8-10</sup>. Tens of thousands of *Sāncipāt* manuscripts still exist in Assam,  
36 most of them without losing their herbal ink and luster for centuries in a harsh and humid climate  
37 of Assam<sup>11</sup>. The first record of *Sāncipāt* manuscript is in *Harshacharitam* by Banabhatta, a  
38 biography of king *Harshavardhana* (606-647AD) of Kanouj, now in North India<sup>12,13</sup>. A bundle of  
39 manuscripts, made of *Sāncipāt* with a reddish yellow color of *Hāitāl* (yellow orpiment), was gifted  
40 to *Harshavardhana* by king *Kumar Bhaskaravarman* (595-650AD) of Kamrupa, then Assam. This  
41 unique tradition of writing *Sāncipāt* manuscript, started by 7<sup>th</sup> century which is assumed to be  
42 continued till the early 20<sup>th</sup> century<sup>14,15</sup>, predominated over other contemporary writing bases like  
43 *Tulāpāt* (hand-made paper) and *Tālapatra*, but now is almost extinct<sup>16</sup>. The tradition gained a new  
44 momentum during the Vaishnavite movement of Assam lead by the Vaishnavite Saint Srimanta  
45 Sankardev during the fifteenth century, carried forward to a great popularity by Vaishnavite Satras  
46 of Assam, and also patronized by Ahom and Koch kingdom<sup>16,17</sup>.

47

<Figure 1>

48 Due to its hot and humid climate, Assam is a hub of fungi and insects which feed on  
49 cellulosic materials. Interestingly, it is a general observation that the survival ability of properly  
50 made *Sāncipāt* manuscript is greater than other types of manuscript found in Assam where fungi  
51 and insects easily destroy manuscripts. Thus, although the choice of a writing base usually depends  
52 on availability of the material, the unique writing base of *Sāncipāt* seems to have been chosen due  
53 to its greater survival ability over others in its harsh hot and humid climate in addition to  
54 availability of *Sānci* tree in Assam<sup>8,17</sup>. The preparation of *Sāncipāt* from *Sānci* bark involves  
55 repeated smoothening, pressing and drying; partial degumming; polishing with fatty pulse paste;  
56 application of yellowish coating with *Hātāl* (HgS, *Cinnabar*) and a border with *Hengul* (As<sub>2</sub>S<sub>3</sub>,  
57 *Yellow Orpiment*), and punching a hole at the center for tying bundles of *Sāncipāt* which have been  
58 reported on several historical and cultural contexts<sup>17,18</sup>. This arduous process of preparation of  
59 *Sāncipāt* through several steps involving some interesting ingredients thought to be responsible  
60 for its special properties, viz., strength, glaze, durability and ability to retain ink and pigment.  
61 There are well-researched methods for conservation of manuscripts made of papyrus<sup>19</sup>, paper<sup>20</sup>,  
62 parchment<sup>21</sup> and *Tālpātra*<sup>22</sup> but absolutely no scientific method is available for conservation of  
63 *Sāncipāt*, which is different from other cellulose based writing bases. Thus, a customized scientific  
64 method of conservation is urgently needed to protect tens of thousands of *Sāncipāt* manuscripts.

65 A scientific study of the traditional knowledge involved in the recipe and the characteristics  
66 of *Sāncipāt* may unravel the secrets of using the unique writing base in preference to others or the  
67 secrets behind its survival for centuries with undiminished glaze in an unfavorable climate. The  
68 study may also help in deeper understanding of the craftsmanship and technology that was used  
69 and provide clues for its appropriate method for conservation of *Sāncipāt* manuscripts for which

70 there does not exist any specific conservation method. The present work was aimed at carrying out  
71 a systematic scientific study of the traditional knowledge involved in *Sāncipāt* manuscript through  
72 characterization of the folio at various stages of preparation by the traditional recipe through  
73 physicochemical and biochemical analysis with a focus on the possible factors of its survival for  
74 centuries retaining the ink and glaze in a hot and humid climate. In order to avoid destructive  
75 method, we prepared fresh model *Sāncipāt* by the traditional method under supervision of one of  
76 a few traditional practitioners. We also had it in our mind to find some clues for customized  
77 conservation of *Sāncipāt* manuscripts.

78

## 79 **Experimental**

80 *Sānci* bark was obtained from Debajit Gogoi from Golaghat, Assam, who grows *Sānci* tree crop  
81 for extracting a valuable perfume oil from its stem. The *Sānci* bark strips of about 2m long, 15cm  
82 wide and 3mm thick were collected from a 17-years old *Sānci* tree (Figure 2). The mineral  
83 pigments, viz., *Hāitāl*, *Hengul* and *Tutia* (blue vitriol,  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ ) were obtained from  
84 Kamakhya Bhandar, Nagaon, Assam (Figure 2). XRD spectra of *Hāitāl*, *Hengul* and *Tutia*  
85 indicates them to be  $\text{As}_2\text{S}_3$ ,  $\text{HgS}$  and  $\text{CuSO}_4 \cdot 5\text{H}_2\text{O}$ , respectively. Potato dextrose broth (PDB) and  
86 Agar were purchased from Himedia, Mumbai. Distilled water was used to prepare solutions used  
87 in degumming process.

88

<Figure 2>

89 Scanning electron micrograph (SEM) and energy dispersive X-ray (EDX) spectra were  
90 recorded on a Jeol machine, JSM 6390LV. Fourier transform-infrared (FT-IR) spectra were  
91 recorded on a Perkin Elmer spectrophotometer, (Frontier MIR FIR). X-ray diffraction (XRD)  
92 spectra were recorded on a D8 FOCUS (Software: XRD COMMANDER 2, DIFFRAC.EVA.)

93 spectroscope and PCPDF-WIN software was used to analyze them. Tensile strength was measured  
94 using a universal testing machine, Zwick-Z010. Glossiness of surface was measured with digital  
95 glossmeter (S.C. Dey & Co., Kolkata).

#### 96 **Preparation of Sāncipāt by traditional method**

97 *The major steps traditional method of preparation of a Sāncipāt folio, in brief, involves removal*  
98 *of a strip of bark of several feet length and several inches width from a mature Sānci tree, drying*  
99 *in sunlight the strip rolled inside out, smoothening, cutting into size of folios, partial degumming*  
100 *using tutia, drying and smoothening again, application of a primer of a paste fatty pulse, and*  
101 *application of a coating of Hāitāl and a boarder of Hengul before writing.*<sup>17,18,19</sup> Sāncipāt folios  
102 were prepared for this study under supervision of a practicing expert of the tradition following  
103 traditional procedure as briefed below. Sāncipāt at various stages of preparation can be seen in  
104 Figure 3. The outer scale of the Sānci bark was cleaned using a knife, rolled inside out and is dried  
105 in the sunlight for 10 days. The strip was dampened, cleaned, smoothened and then dried again.  
106 Then the strips were cut into pieces of about 12 inches long and 4 inches wide. These pieces were  
107 analyzed as raw Sānci bark. The pieces were soaked in 500 ml water in a plastic tray in presence  
108 of 0.1M Tutia (CuSO<sub>4</sub>), 10 crushed seeds of Konibih (*Croton Triglium*) and 30g of crushed leaves  
109 of Chalkuwari (*Aloe Barbadensis*) (Figure 2) for 12h for partial degumming. These materials were  
110 collected locally. Both surfaces of the strip were again scrapped with a small knife and smoothened  
111 by rubbing with a Ghila, a hard seed with red color, nicker bean, *Entada scandens* Benth (Figure  
112 2). Then, a thin coat of a fine paste of a skinned fatty pulse, called Matimah (*Phaseolus radiatus*  
113 Linn.), was applied on the strip to fill up any cracks or wrinkles on the surface of the strip and then  
114 sundried. The dry strips were smoothened again by rubbing with a Ghila. Then, a thin coating of  
115 yellow Hāitāl pigment was applied on both sides of the strip. For this, the paint is prepared by

116 mixing powdered *Hāitāl* in water using an herbal gum obtained from fruit of *Bel* (Stone apple,  
117 *Aegle Marmelos*). The pieces are again sundried and smoothed with a *Ghila*. A red boarder of  
118 about 1 cm width of red *Hengul* pigment is also applied along the boundaries on both sides of the  
119 strip which is ready for manuscript writing and illustration. The freshly prepared *Sāncipāt* was  
120 found to be physically strong, smooth, and glazing. The samples were powdered and dried in  
121 desiccator for 7 days prior to physicochemical analysis.

122 <Figure 3>

123

#### 124 **Physicochemical characterization at various stages of preparation of *Sāncipāt***

125 In each step of preparation of *Sāncipāt* from a raw *Sānci* bark, changes in physical and chemical  
126 properties have been analyzed through various techniques. The effect of smoothing of fibrous  
127 lignocellulose structure is apparent from SEM images (Figure 4). The EDX analysis showed the  
128 atomic percentage of carbon, nitrogen, and oxygen in cleaned raw *Sānci* bark as 44.66%, 10.87%,  
129 43.15%, respectively, commensurate with its lignocellulosic composition<sup>24</sup> (Figure 4). The ratio  
130 changed to 54.08%, 10.20% and 34.90%, respectively, in freshly prepared *Sāncipāt* due to removal  
131 of some lignin during degumming and addition of proteins and fats in the coating of fatty pulse.  
132 The atomic percentage of carbon, nitrogen, and oxygen obtained for an old damaged *Sāncipāt* were  
133 46.69%, 8.16% and 44.06%, respectively. The elements of Mg, Si, S, Cl, K, Ca, As, Pb were  
134 detected in trace amounts in all three samples. The amount of arsenic was found to be highest in  
135 the freshly prepared *Sāncipāt* which probably comes from *Hāitāl*. Cu was absent in the raw *Sānci*  
136 bark but observed in the freshly prepared *Sāncipāt* and the old *Sāncipāt* which can be attributed to  
137 *Tutia* used during partial degumming.

138 <Figure 4>

139 FT-IR spectra of raw *Sānci* bark and *Sānci* barks at three stages of preparation also  
140 indicated presence of lignin, cellulose and hemicellulose (Figure 5). In traditional method of  
141 preparation of *Sāncipāt*, only a partial degumming is done retaining the fibrous cellulosic part in  
142 order to retain the basic structure and shape of the bark. As expected, the FT-IR spectra of the old  
143 manuscript folio was also found to be similar to that of the freshly prepared one.

144 <Figure 5>

145 Similar largely amorphous XRD patterns were obtained for both freshly prepared *Sāncipāt*  
146 and old *Sāncipāt* samples (Figure 5). However, characteristic peaks observed at  $14.1^\circ$  (-1 0 1) and  
147  $22.8^\circ$  (0 0 2) and  $41.6^\circ$  (2 4 0),<sup>25</sup> suggest them to be native cellulose,  $(C_6H_{12}O_6)_x$ , system-  
148 monoclinic,<sup>21</sup>(PDF no. 030289; ID:03-0289)<sup>25</sup>. A broad peak observed in the range between  $21.90^\circ$   
149 to  $22.80^\circ$  for freshly prepared *Sāncipāt* which was flattened in old *Sāncipāt* may be attributed to a  
150 slight change in cellulose structure.

151 The tensile strength (longitudinal) values of cleaned raw *Sānci* bark, traditionally  
152 degummed *Sānci* bark and freshly prepared *Sāncipāt* after drying are shown in Figure 6.  
153 Elongation at break was same for all three samples and was approximately 7.8%. The ultimate  
154 tensile strength of the cleaned raw *Sānci* bark considerably increased from 20MPa to 48MPa and  
155 78MPa in the degummed and freshly prepared *Sāncipāt*, respectively. The Young's modulus (Y/X)  
156 calculated from three points on the three curves at (X=3.32, Y=9.97), (X=3.32, Y=28.63) and  
157 (X=3.32, Y=48.01) were found to be 3.00, 8.62 and 14.46 for cleaned raw *Sānci* bark, degummed  
158 *Sānci* bark and freshly prepared *Sāncipāt*, respectively. Thus, the tensile elasticity of the *Sānci*  
159 strip is significantly increased during preparation of *Sāncipāt*. The degumming and application a  
160 coating of paste of fatty pulse with stone apple gum have probably contributed to the toughness of  
161 *Sāncipāt*.

162 The averages of glossiness of inner side of the folios at three stages of preparation are  
163 shown in Figure 6 along with the average of glossiness of three folios of a damaged old manuscript  
164 at three different angles of measurements. It has been observed that the glossiness of the folio is  
165 increased with the angle of measurement as expected. It was interesting to note an increase in the  
166 glossiness gradually in each stage of the preparation. The improvement in gloss index from stage  
167 1 to 3 may be attributed to increase in reflection due to filling of the pores and cracks on the surface  
168 by the coating of fatty pulse and the coating of *Hāitāl*. However, the glossiness of the old  
169 manuscript was much lower than stages 2 and 3 of the fresh ones as expected. The gloss index of  
170 both inner and outer sides of freshly prepared *Sāncipāt* were also compared to examine if there  
171 was any difference between the two sides and found to be almost identical (Figure SI-1).  
172 Interestingly, it was not possible to distinguish the inner and outer sides of an old manuscript.

173 The effect of the duration of soaking of *Sāncipāt* in water presence of 0.1M CuSO<sub>4</sub> on  
174 weight loss or partial degumming of the *Sāncipāt* was studied by varying the duration up to 24h  
175 [figure SI-2]. The weights were recorded before soaking, and after soaking and drying without  
176 scrapping with knife. It is interesting to note a linear relation of percent weight loss, calculated as  
177 (final weight of strip/initial weight of strip) × 100%, with time (SI-2). The weight loss may be  
178 attributed to a gradual loss of hemicellulose and lignin of *Sānci* strip with time during the process.

179

#### 180 **Antifungal activity:**

181 For testing antifungal activity of *Sānci* bark and *Sāncipāt*, three filamentous fungi, viz., *Aspergillus*  
182 *niger*, *Candida albicans* and *Fusarium oxysporum*, were sub-cultured. A standard procedure was  
183 used for antifungal activity test on *Sānci* strip at four different stages of the preparation<sup>26</sup>. The  
184 *Sāncī* manuscript was cut into small pieces having the equal surface area for the experiment. For



185 preparation of potato dextrose broth (PDB) solution, 5g of PDB was dissolved in 50 mL distilled  
186 water followed by autoclave at 121°C and 15 pounds for 15 min. Moreover, potato dextrose plates  
187 were prepared with PDB and 1.8 g of agar. All the fungal strains were seeded to 50 mL of PDA  
188 containing conical flasks and incubated for 7 days at approximately 20°C. For the antifungal test,  
189 100 µL of fungal inoculums were spread carefully in the PDA plates and placed all small pieces  
190 of the samples. The samples along with controls were incubated in static incubator and their growth  
191 were monitored systematically. Water was used as a negative control whereas antimycotic solution  
192 from Himedia at 10µg/mL were used as a positive control. All the experiments were performed in  
193 triplicates.

194 The results of antifungal test on samples of cleaned raw *Sānci* bark, degummed *Sānci* bark,  
195 *Sānci* bark after application of *Hāitāl* and *Sānci* bark after application of both *Hāitāl* and *Hengul*  
196 (Figure 6). The growth of the fungi takes place within 48-72h. The observations of antifungal  
197 activity in open control, closed negative control and closed positive control were made after 168h  
198 (SI-3). After 2h, it was observed that all three fungi grew equally in the open control. *Aspergillus*  
199 *niger*, being the most voracious fungus among the three, grew most rapidly in the negative control.  
200 On the other hand, *Candida albicans* grew most rapidly in the positive control. The fungal growths  
201 on the open positive and negative control have been observed to increase after one week also.  
202 There was no fungus growth on any of the samples before incubation. Interestingly, we observed  
203 growth of all three fungi only in the petri plates for cleaned raw *Sānci* bark after one week of  
204 incubation [SI-3]. But no growth of any of the fungi was observed on the degummed *Sānci* bark,  
205 *Sānci* bark after application of *Hāitāl* and *Sānci* bark after application of both *Hāitāl* and *Hengul*.

206 <Figure 6>

207 Fungus feed on cellulosic materials and so *Sāncipāt* is likely to be damaged by fungi in a  
208 hot and humid climate that exists during most of the year in Assam. On the other hand, the three  
209 minerals, viz., *Tutia*, *Hāitāl* and *Hengul*, are also reported to have antifungal properties<sup>16</sup>. The  
210 absence of any growth of all three fungi on the samples of degummed *Sānci* bark, *Sānci* bark after  
211 application of *Hāitāl* and *Sānci* bark after application of both *Hāitāl* and *Hengul* is expected to be  
212 attributed to antifungal properties of CuSO<sub>4</sub> used during degumming of *Sānci* bark and application  
213 of coating of *Hāitāl* and *Hangul*. The antifungal property of *Sāncipāt* may be additive, synergistic,  
214 or cumulative in nature. Interestingly, the authors have witnessed in various libraries, museums  
215 and personal possessions that *Sāncipāt* manuscripts with coating of *Hāitāl* and *Hengul* are never  
216 destroyed by fungus or insects while those without the coatings are mostly damaged by fungi or  
217 insects (SI-4). It is interesting to note the traditional knowledge of antifungal properties of these  
218 minerals and their use for protection of *Sāncipāt* manuscripts from fungus in Assam during the  
219 medieval period. This indicates that application of a thin coat of *Hāitāl* on *Sāncipāt* manuscripts  
220 on the free spaces of the cleaned manuscript folios to be a good option for conservation of the  
221 manuscripts followed by a thin coating of wood sap in a traditional way to avoid future  
222 contamination during reading. Though *Hāitāl* is known to be toxic to fungi and insects, it is used  
223 traditionally in make-up of dancer in Kathakali dance and for lightening of dark skin. However,  
224 its prolific use in cosmetics has been reported to pose risk to human health<sup>27</sup> and care must be  
225 taken while handling and applying toxic *Hāitāl* using gloves and masks.

226

#### 227 **Conclusions:**

228 The present study reveals some interesting traditional knowledges associated with preparation of  
229 *Sāncipāt* related to their attractive look, physical properties, composition, adhesion of the ink on

230 it and longevity of the manuscript in harsh climate. Raw *Sānci* bark consists of native cellulose  
231 together with some hemicellulose and lignin devoid of nitrogenous compounds. The traditional  
232 degumming process is less drastic than common alkaline degumming which partially removes the  
233 unwanted hemicellulose and lignin leaving mostly the fibrous cellulosic part. The amount of  
234 carbon and oxygen in freshly prepared *Sāncipāt* is slightly higher than that of raw *Sānci* bark. The  
235 smoothness of surface, tensile strength and gloss index of *Sāncipāt* are increased during the  
236 preparation of *Sāncipāt*.

237 Antimicrobial test shows that raw *Sānci* bark has no antifungal properties but the *Sānci* bark  
238 after degumming in the presence of  $\text{CuSO}_4$ , and after application of a coating of *Hāitāl* and *Hengul*  
239 shows remarkable inhibition towards fungi. The antifungal property of *Sāncipāt* manuscript have  
240 been attributed to a synergistic effect of  $\text{CuSO}_4$  used during degumming and due to application of  
241 *Hāitāl* and *Hengul*. The findings may help in developing a customized method of conservation of  
242 old Sāncipāt manuscripts.

243

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#### 250 **References:**

- 251 1. Gaur, R.C. and Chakraborty, M., Preservation and access to Indian manuscripts : A knowledge  
252 base of Indian cultural heritage resources for academic libraries. *In International Conference*  
253 *on Academic Libraries*, Delhi, 2009, 5-8.
- 254 2. Quenzer, J., Bondarev, D. and Sobisch, J-U., Manuscript cultures: Mapping the field. De  
255 Gruyter, Berlin, 2014.
- 256 3. Fleming, B.J., The materiality of South Asian manuscripts from the University of Pennsylvania  
257 ms coll. 390 and the Rāmamālā library in Bangladesh. *Manuscript Studies*, 2016, **1**, 28-51.
- 258 4. Goswami, H., A note on Assamese manuscripts, in descriptive catalogue of Assamese  
259 manuscripts. Calcutta University, Kolkata, 1930.
- 260 5. Banerji, S.C., A companion to Sanskrit literature. Motilal Banarsidass Publication, 1989.
- 261 6. Agrawal, O.P., Conservation of manuscripts and paintings of South-East Asia. Trubner & Co.  
262 Ltd., London, 1984.
- 263 7. Fuchs, L.S., Fuchs, D., Hosp, M.K. and Jenkins, J.R., Oral reading fluency as an indicator of  
264 reading competence: A theoretical, empirical, and historical analysis. *Scientific Studies of*  
265 *Reading*, 2001, **5**, 239-256.
- 266 8. Goswami, B.R., Das, M.K., Das, P.P., Medhi, T., Ramteke, A., Hazarika, S. and Dutta, R.K.,  
267 Mahi: A unique traditional herbal ink of early Assam. *Curr Sci*, 2017, 591-595.
- 268 9. Goswami, B.R., Chamuah, N. and Dutta, R.K., A physicochemical characterisation of a  
269 medieval herbal ink, mahī, of Assam, India. *Coloration Tech*, 2018, **134**, 450-463.
- 270 10. Goswami, B.R., Das, D., Saikia, P. and Dutta, R.K., Role of saponins on enhancement of quality  
271 of Mahī, a traditional herbal ink of early Assam. *Dyes Pigments*, 2021, **188**, 109234.

- 272 11. Mazumdar, N.R., Digital preservation of rare manuscripts in Assam. International Convention  
273 on Automation of Libraries in Education and Research, Pondicherry University, Pondicherry,  
274 2009.
- 275 12. Hermann, K. and Rothermund, D., A history of India. Routledge, London, 2004.
- 276 13. Cowell E.B and Thomas, F.W., The Harshacharita of Banabhatta. Royal Asiatic Society,  
277 London, 1897.
- 278 14. Neog, M., Early history of the vaisnava faith and movement in Assam. Motilal Banarsidass,  
279 New Delhi, 1965.
- 280 15. Datta, B., Folk painting in Assam. Tezpur University, Tezpur, 1998.
- 281 16. Nath, D., Religious tradition and social practices in Assam. Essays on popular religion, buddhist  
282 tradition and writing culture. DVS Publication, Guwahati, 2015.
- 283 17. Dutta, R.K., The science in the traditional manuscript-writing aids of Assam: Sancipat, mahi  
284 and hengul-haital. In *Religious traditions and social practices in Assam* (ed. Nath, D.), DVS  
285 Publication, Guwahati, pp 239-261.
- 286 18. Sharma, S.K. and Sharma, U., Discovery of north-east india: Geography, history, culture,  
287 religion, politics, sociology, science, education and economy. Assam. Mittal Publications, 2015.
- 288 19. Roger, B., Papyrus and preservation. *The Classical World*, Baltimore, US, 1998, **91**, 543-552.
- 289 20. Ogden, B., On the preservation of books and documents in original form. Report. Commission  
290 on Preservation and Access, Washington DC, 1989.
- 291 21. Yusopova, M.V., Conservation and restoration of manuscripts and bindings on parchment.  
292 *Resturator* 1980, **4**, 57-70.
- 293 22. Suryawanshi, D., Sinha, P. and Agrawal, O., Basic studies on the properties of palm leaf.  
294 *Resturator*, 1994, **15**, 65-78.

- 295 23. Gait, E.A., A history of Assam. Thacker, Spink & Company, London, 1906.
- 296 24. McQueen, C.M.A., Mortensen, M.N., Caruso, F., Mantellato, S. and Braovac, S., Oxidative  
297 degradation of archaeological wood and the effect of alum, iron and calcium salts. *Heritage Sci*,  
298 2020, **8**, 1-10.
- 299 25. Oreshkina, A., Kaziev, G., Steblevskii, A., Quinones, S.H., Peretokina, O. and De Ita, A.,  
300 Synthesis and physicochemical study of nonamolybdomanganate with the hexamminecadmium  
301 cation. *Russian J Inorg Chem*, 2013, **58**, 512-514.
- 302 26. Bhalodia, N.R. and Shukla, V., Antibacterial and antifungal activities from leaf extracts of  
303 cassia fistula l.: An ethnomedicinal plant. *J Adv Pharm Tech Res*, 2011, **2**, 104.
- 304 27. Mohammed, T., Mohammed, E. and Bascombe, S., The evaluation of total mercury and arsenic  
305 in skin bleaching creams commonly used in Trinidad and Tobago and their potential risk to the  
306 people of the Caribbean. *J Public Health Res*, 2017, **6**, 1097.

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313 **Figure captions: (main document)**

314

315 Figure 1. A *Sāncipāt* folio from Adi Dasham, a manuscript dated 1799 AD, preserved at Bengena  
316 Ati Satra, Majuli, Assam. The folio is coated with yellow *Hāitāl* with art on a wide boarder of  
317 *Hengul* and written with *Mahr*<sup>8-10</sup>, a traditional herbal ink of early Assam. The illustrations were  
318 drawn with different proportions of *Hengul*, *Hāitāl* and Indigo.

319  
320 Figure 2. A *Bholā Sānci* tree after removal of bark (a), *Sānci* bark strip rolled inside out for drying  
321 (b), *Sānci* bark after degumming (c), after fatty pulse polishing (d), after application of *Hāitāl* (e)  
322 and after drawing boarder with *Hengul* and writing with *Mahī* (f).

323  
324 Figure 3. Chalkuwari (*Aloe-vera*, *Aloe barbadensis*) (a) Koni Bih guti (*Crotton Triglium*) (b) and  
325 Ghila (c), Pieces of *Hāitāl* (d) and *Hengul* (e) and fine powder of *Hāitāl* (f) and fine powder of  
326 *Hengul* (g) and Tutia (h).

327  
328 Figure 4. SEM images of raw *Sānci* bark strip (a), freshly prepared *Sāncipāt* (b) and old *Sāncipāt*  
329 (c) at a resolution of 100  $\mu\text{m}$ , and EDX analysis of raw *Sānci* bark strip (d), freshly prepared  
330 *Sāncipāt* and old *Sāncipāt* (f).

331  
332 Figure 5. FT-IR spectra of freshly prepared *Sāncipāt* (a) and old *Sāncipāt* (b); XRD patterns of  
333 freshly prepared *Sāncipāt* (c) and old *Sāncipāt* (d); tensile strength (e) of cleaned raw *Sānci* bark  
334 (i), traditionally degummed *Sānci* bark (ii) and freshly prepared *Sāncipāt* (iii); and average gloss  
335 index (f) of *Sānci* bark at three stages of preparation: Stage 1 - cleaned raw *Sānci* bark, Stage 2 –  
336 after application of fatty pulse-paste, Stage 3 - freshly prepared *Sāncipāt*, and of old *Sāncipāt*  
337 measured at 20°, 60° and 80° angles.

338  
339 Figure 6. Results of antifungal test on samples of cleaned raw *Sānci* bark, degummed *Sānci* bark,  
340 *Sānci* bark after application of *Hāitāl* and *Sānci* bark after application of both *Hāitāl* and *Hengul*  
341 before (top) and after (bottom) incubation.

342 Figures:

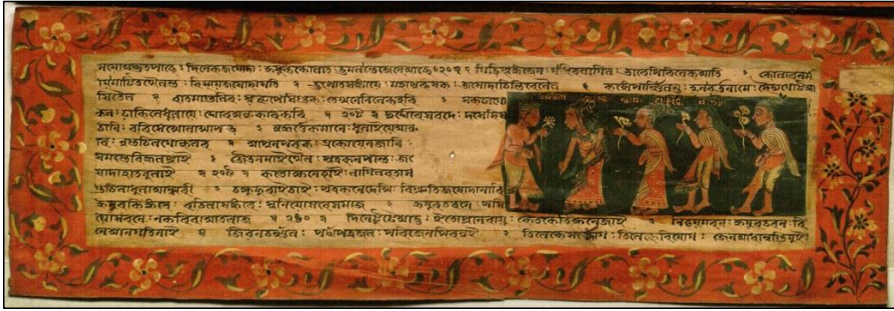
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Figure 1

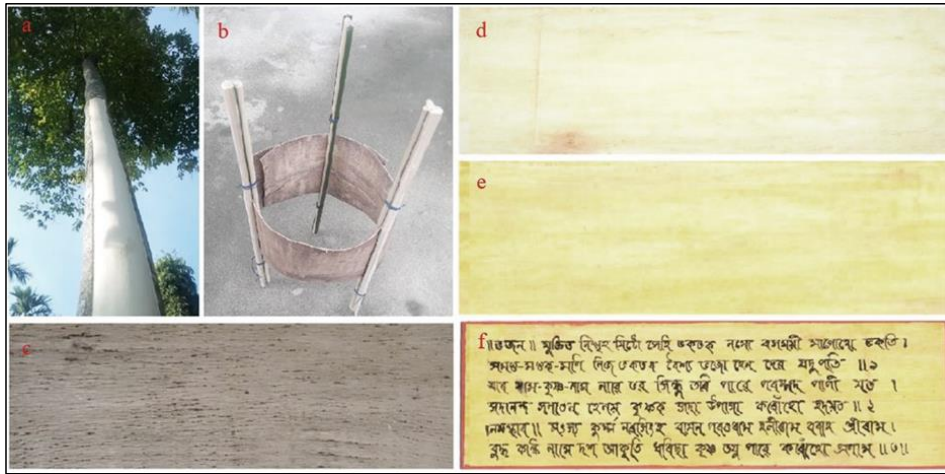


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Figure 2

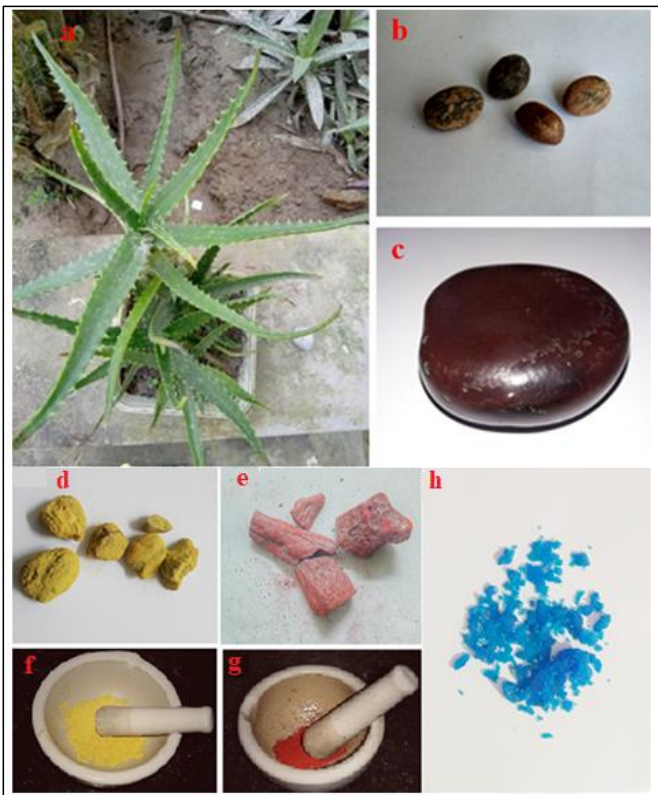


Figure 3

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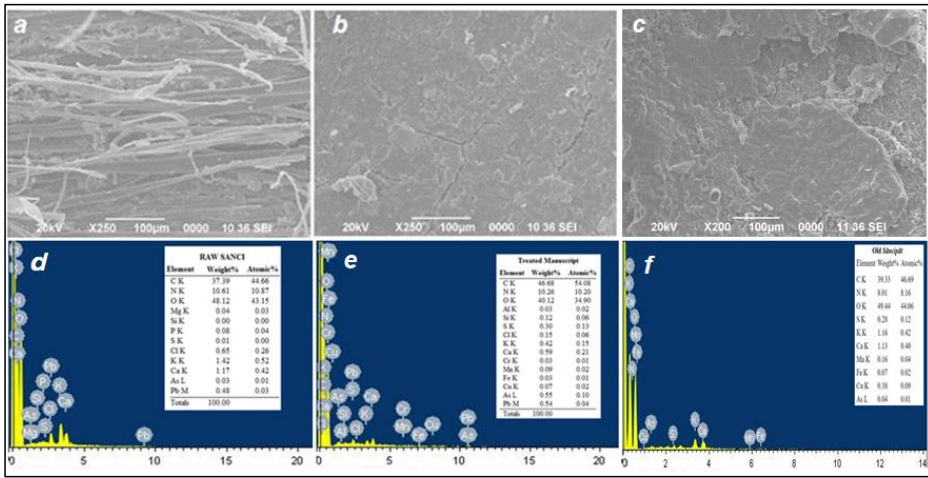
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Figure 4

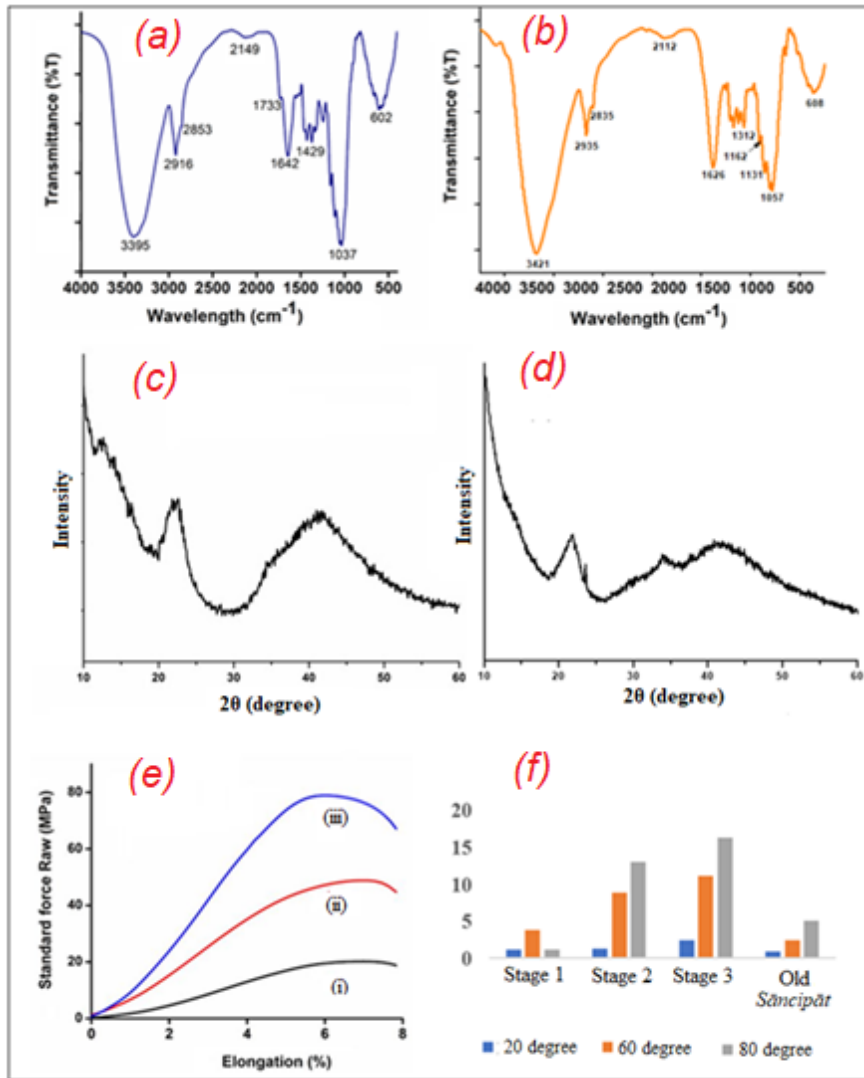


Figure 5

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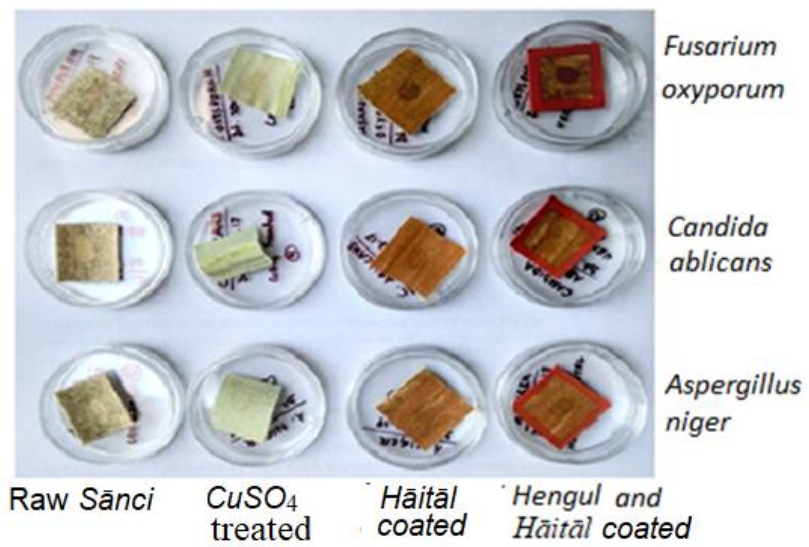
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Figure 6

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