

Health issues related to N pollution in water and air

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Nitrates enter human body through drinking water, food and air. Ingested nitrates converted to nitrite by microflora lead to methaemoglobinemia, increased free oxide radicals that predispose cells to irreversible damage and effects like cancer, increased infant mortality, abortions, birth defects, recurrent diarrhoea, recurrent stomatitis, histopathological changes in cardiac muscles, alveoli of lungs and adrenal glands, deterioration of immune system of the body. When inhaled, NO_x can cause unconsciousness, vomiting, mental confusion, congestion and inflammation of the respiratory tract, pulmonary oedema, genetic mutations, and adversely affect development of the foetus and decrease fertility.

Keywords: Cytochrome b₅ reductase, methaemoglobinemia, nitrate, nitrite.

NATURALLY occurring nitrate levels in surface and groundwater are generally a few milligrams per litre. Higher nitrate levels are found in groundwater due to water percolating through nitrate-rich rocks and also due to an excessive use of chemical fertilizers. The WHO report of 2004 maintains that extensive epidemiological data support limiting the value of nitrate-nitrogen to 10 mg/l or as nitrate to 50 mg/l for human consumption¹, whereas IS-10500 prescribes² maximum permissible limits in drinking water as 45 mg of NO₃/l².

Sources

The main sources contributing to nitrate content of natural waters are atmosphere, geological features, anthropogenic sources, atmospheric nitrogen fixation and soil nitrogen. Oxides of nitrogen are generated through lightning and reach the surface water with rain. Recent reports indicate that atmospheric contributions amount to 25% of the total load of nitrate³. Direct discharge from septic

tanks, sewage and industrial effluent are other contributors. Excessive use of chemical fertilizers is one of the main sources of nitrate in water.

Out of the total human nitrate intake³, fruit and vegetables account for 70%, drinking water 21%, and meat and meat products 6%. The common nitrate-rich vegetables are lettuce, spinach, beetroot, celery, egg plant, beet, banana, strawberry, tomatoes and peas. Preservatives used in the food industry are significant nitrate sources. Cooking in aluminum utensils enhances reduction of nitrates to nitrite³, and hence increases the toxicity.

Nitrates known as endogenous nitrate are also produced in the body. A major pathway for endogenous nitrate production is conversion of arginine by macrophages to nitric oxide and citrulline, followed by oxidation of the nitric oxide to nitrous anhydride, and then reaction of nitrous anhydride with water to yield nitrite. Gastrointestinal infections and non-specific diarrhoea increase endogenous (non-bacterial) nitrate synthesis, probably induced by activation of the mammalian reticuloendothelial system^{3,4}.

Hydrogeological investigations show that nitrate levels are high in sandy soil than in clayey soil, because of low water-holding capacity and high permeability of pollutants like chloride and nitrate.

Kinetics and metabolism

About 20% of ingested nitrate is reduced to nitrite by nitrate-reducing microflora present in the saliva⁵ at the base of the tongue⁶. The factors which influence oral microflora and hence reduction of nitrate are nutritional status, infection, environmental temperature and age (more in elderly)⁵.

Ingested nitrate is reduced to nitrite by nitrate-reducing microflora in the stomach (under favourable conditions, viz. pH ≥ 4) and upper part of the intestine. Conditions favouring high stomach pH are achlorhydria⁷, atrophic gastritis^{6,8}, artificially fed infants, or patients using anti-acid or similar drugs, e.g. Omeprazole^{9,10}.

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Nitrate intake by humans through leafy vegetables

Being a rich source of nutrients and antioxidants, leafy vegetables occupy an important place in the human diet. Some vegetable species are known to accumulate high concentration of nitrate under heavy fertilization. Vegetables are the major source of the daily intake of nitrate by human beings, supplying about 72–94% of the total intake. Part of this nitrate-N is converted to nitrite and N-nitroso compounds that have detrimental effects on human health.

The Joint Expert Committee on Food and Agriculture in the World Health Organization (WHO)¹ established the Acceptable Daily Intake (ADI) of nitrate as 0–3.7 mg kg⁻¹ body wt. The US Environmental Protection Agency's reference dose for nitrate is 7.0 mg kg⁻¹ body wt per day². In 1995, the European Commission's Scientific Committee for Food (SCF) established the ADI of nitrate as 3.65 mg kg⁻¹ body wt (equivalent to 219 mg day⁻¹ for a person weighing 60 kg)³. Assuming a 60 kg body wt⁴, the ingestion of 100 g of fresh vegetables with nitrate concentration of 2500 mg kg⁻¹ fr wt exceeds the ADI for nitrate by about 13%. For a real assessment, however, nitrate content as well as average daily consumption amount in other sources need to be considered. Interestingly, at least 50% of the nitrate can be removed by cooking vegetables in water (with low nitrate concentration).

The nitrate content in samples of chenopodium and spinach being sold to Indian consumers has been found to be as high as 4451 and 4293 mg kg⁻¹ fr wt⁵. Studies on genotypes of spinach have shown enormous intraspecific variation of nitrate content. One genotype at the three-week stage of plant growth, and six genotypes at the six-week stage, exceeded the ADI limit. Petioles possessed several times higher level of nitrate than leaf lamina. All of the genotypes studied showed diurnal variation in nitrate accumulation with a minimum at noon^{4,5}.

Nitrate content in leafy vegetables possesses a significant reverse relationship with nitrate reductase (NR) activity, the first enzyme in the nitrate assimilatory pathway. It is held that NR expression level is largely responsible for different nitrate accumulation patterns in different cultivars of a given species. Thus nitrate accumulation in plants can be significantly reduced by over expression of NR genes in high nitrate-accumulating genotypes. In addition, it is also important to enhance the expression of NR so that nitrate does not accumulate in the green tissue after converting into nitrite. Over expression of these genes will reduce the nitrate concentration in vegetables to safe limits for human consumption. Moreover, a careful selection of vegetable genotypes based on their relative levels of nitrate content and NR activity, harvesting young plants at noon time, and removal of petioles from leaves could minimize the dietary intake of nitrate through leafy vegetables.

1. Speijers, G. J. A. (ed.), Nitrate. In *Toxicological Evaluation of Certain Food Additives and Contaminants in Food. Food Additive Series 35*, WHO, Geneva, 1996, pp. 325–360.
2. Mensinga, T. T., Speijers, G. J. A. and Meulenbelt, J., Health implications of exposure to environmental nitrogenous compounds. *Toxicol. Rev.*, 2003, **22**, 41–51.
3. Scientific Committee on Food, Opinion on nitrate and nitrite. Annex 4 to Document III/5611/95, European Commission (ed.), Brussels, 1995, p. 20.
4. Anjana, Umar, S. and Iqbal, M., Nitrate accumulation in plants, factors affecting the process, and human health implications. A review. *Agron. Sustain. Dev.*, 2007, **26**, 45–57.
5. Anjana, Umar, S., Iqbal, M. and Abrol, Y. P., Are nitrate concentrations in leafy vegetables within safe limits? *Curr. Sci.*, 2007, **92**, 355–360.

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The nitrite is readily and completely absorbed from both the stomach and the upper small intestine. The absorbed nitrite is then rapidly distributed throughout the tissues. It is rapidly oxidized to nitrate in the blood, with the formation of methaemoglobin.

The fate of nitrate is also associated with a metabolic pathway, which produces N-nitroso compound. This is a multiple-step process¹¹. First, nitrate is converted into nitrite after consumption. Second, the nitrite reacts with natural or synthetic organic compounds (such as secondary amines or amides) in the food or water to form new combinations, called N-nitroso compounds (either nitros-

amines or nitrosamides). More than one hundred of these N-nitroso compounds have been tested for carcinogenicity in animals¹², and 75–80% of them have been found to be carcinogens^{4,13,14}. IARC¹⁵ concluded that 11 N-nitroso compounds were carcinogenic in man. The most common N-nitroso compounds are dimethylnitrosamine (DMN), N-methylmethanamine (DMA), trimethylamine (TMA) and trimethylamine oxide (TMAO). Nitrite has been shown to cross the placenta and cause the formation of foetal methaemoglobinemia in rats¹⁶. The half-life of nitrate in the body after ingestion¹⁸ is approximately 5 h¹⁷.

Metabolism of ingested nitrate in human body at cellular level

Ingestion of inorganic or organic nitrates will result in increased oxidation of haemoglobin to methaemoglobin and increased production of nitric oxide^{19,20} (Figure 1). The conversion of nitrite to nitric oxide is non-enzymatic^{20,21}. The oxidation of haemoglobin to methaemoglobin results in the formation of the superoxide radical by the transfer of a single electron. The enzyme superoxide dismutase present in the erythrocytes, catalyses the conversion of superoxide radical (O_2^-) to H_2O_2 and O_2 . H_2O_2 is decomposed by glutathione peroxidase or catalase, both also present in erythrocytes^{22,23}. Once the rate of oxidation of haemoglobin increases sufficiently in erythrocytes and overwhelms the protective and reductive capacities (e.g. cytochrome b_5 reductase system, etc.) of the cells^{24,25}, there is increased production of reactive free radicals of nitric oxide (NO^*) and oxygen (O_2^*)²².

Fate of free radical nitric oxide

Haemoglobin scavenges nitric oxide through the high-affinity ferrous sites on heme to form S-nitrosothiol, whose affinity to nitric oxide is 8000 times higher than that for oxygen²⁶ by binding at β -93 cysteine residue on the globin chain. As haemoglobin binds oxygen in the lungs, its binding affinity to S-nitrosothiol is increased. As haemoglobin releases oxygen at the periphery, its affinity for S-nitrosothiol is reduced and nitric oxide is released in the tissues²⁶. The thiol group of S-nitrosothiol essentially protects nitric oxide from being scavenged by the binding site on heme. Thus in addition to carrying oxygen, haemoglobin acts as a carrier of nitric oxide. The enhanced release of nitric oxide from S-nitrosohaemoglobin in the hypoxic tissue in turn reduces regional vascular resistance.

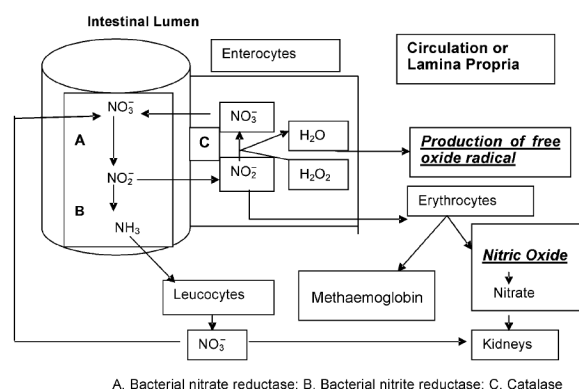


Figure 1. Metabolism of ingested nitrate in human body at cellular level.

Nitric oxide is a biogenic messenger, an endothelial-derived relaxing factor (EDRF)^{25,26} and activates the guanylyl cyclase system²⁷ [converts guanosine triphosphate (GTP) to 3',5' cyclic guanosine monophosphate (cGMP)], raising the cGMP pool and therefore inducing *inter alia* vasodilatation²⁷ by lowering intracellular calcium ion²¹.

Fate of free oxide radical

In a normal cell, O_2^- will be scavenged by the enzyme superoxide dismutase and H_2O_2 , which is a product of the reaction, and by glutathione peroxidase and catalase^{28,29}. Any O_2^- that escapes this mechanism should react with other cell constituents, possibly causing irreversible cell damage. This mechanism is likely to become more significant if O_2^- is produced in abnormally high amount (e.g. excessive nitrate ingestion) or if any of the protective mechanisms are defective^{23,26}. Thus increased consumption of nitrate will lead to (a) increased production of nitrite³⁰, (b) enhanced absorption of sodium from the intestinal lumen²⁸, (c) excess NO^* (free radical nitric oxide) generation having vasodilatory effect^{21,22,26,27} and (d) increased production of O_2^- , which will react with other cell constituents, possibly causing irreversible cell damage^{27,31} (Figure 1).

Excretion

The major part (70–75%) of the ingested nitrate is eventually excreted in the urine as nitrate, ammonia or urea within 24 h. The renal excretion is predominantly tubular³² and occurs more in the first five hours³³. Faecal excretion is negligible. A small amount is also excreted by exhalation through the lungs in the form of oxides of nitrogen (nitric oxide, nitrous oxide, nitrogen, etc.). Little nitrite is excreted^{3,13} in the urine.

Acute toxic effects

Toxic doses of nitrate ingestion in humans have been reported as 2–5 g of NO_3^- (equivalent to 33–150 mg NO_3^- /kg body wt)³⁴. Human lethal doses of 4–50 g NO_3^- (equivalent to 67–833 mg NO_3^- /kg body wt) have been reported.

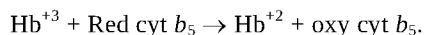
The acute toxicity symptoms occur in the form of cyanosis, severe gastroenteritis with abdominal pain, blood in the urine and faeces, dyspepsia, mental depression, headache and weakness.

Chronic toxic effects

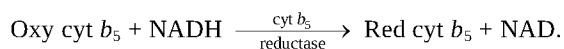
The following effects of long-term exposure to non-lethal doses have been reported.

Methaemoglobinemia

Nitrates in drinking water have been reported to cause methaemoglobinemia, mainly in infants up to 6 months of age. Nitrate, after conversion to nitrite, oxidizes ferrous ion of haemoglobin to ferric state, forming methaemoglobin. The methaemoglobin formed is normally reduced³⁵ by reduced cytochrome b_5 :



This reaction thus forms oxidized cytochrome b_5 , which is further regenerated by getting reduced by the enzyme cytochrome b_5 reductase in the presence of NADH to form red cyt b_5 , making it again available for reducing methaemoglobin.



In conditions where either the consumption of nitrate is more than the tolerable limit or cytochrome b_5 reductase system is not fully developed (infants)³⁵, the cytochrome b_5 reductase system gets exhausted, causing methaemoglobinemia. It has been reported that most cases of methaemoglobinemia occur with 90 mg nitrate ion/l or more^{25,29,36}. Recently, methaemoglobinemia has been reported in all age groups³⁷ and was more severe in infants and in older age groups (>45 years). Infants are more susceptible to nitrate toxicity because of relatively higher stomach pH (2.0–5.0), which permits growth of nitrate-reducing organisms such as coliforms, *E. coli*, *Pseudomonas fluorescens*, *B. subtilis*, *Staph. Albus*, etc., relatively higher consumption of water per unit weight of body, presence of foetal haemoglobin which readily gets oxidized to methaemoglobin and poorly developed cytochrome b_5 reductase system.

Repeated boiling of nitrate-rich water for feeding, high-nitrate water used for preparing dried milk powder, weaning with nitrate-rich vegetables, e.g. spinach and viral diarrhoea in children cause increase in nitrate toxicity. Toxicity to infants with pregnant mothers ingesting high nitrates is possible because of reported transplacental passage of nitrite. Methaemoglobin level in the blood more than 10% of the total haemoglobin, manifests as clinical cyanosis and causes cellular anoxia. When these levels exceed 25–50%, it causes cyanosis, dyspnoea, headache and disorientation. Levels more than 60% may be lethal³⁶.

Cytochrome b_5 reductase adaptation

Gupta *et al.*³⁸ reported that methaemoglobinemia was more pronounced in infants and elderly persons, in comparison to children and adolescents. It was observed that cytochrome b_5 reductase activity increases with increased

nitrate ingestion. This adaptation peaks at about 95 mg nitrate ion/l nitrate concentration and gets exhausted by 200 mg nitrate ion/l, thus making people more prone to toxic effects. This adaptation was more active in children and adolescents in comparison to infants and elderly.

Infant mortality rate

A study on African mothers and other studies also^{39,40} reported an increase in infant deaths with increasing exposure of pregnant mothers and infants to nitrate. This may have been either due to undetected toxic methaemoglobinemia or due to malformations and weaknesses in the infant caused by foetal nitrate exposure.

Nitrate, nitrite, nitrosamines and cancer

Nitrate acts as a 'procarcinogen', meaning that it reacts with other chemicals (amines and amides) to form carcinogenic compounds (N-nitroso compounds). Formation of N-nitroso compounds depends upon the presence of nitrate, nitrate-reducing microbial population and conditions favourable for chemical nitrosation⁴¹.

In animal or human studies, N-nitroso compounds have been associated with 15 different types of cancers, including tumours in the bladder, stomach, brain, esophagus, bone and skin, kidney, liver, lung, oral and nasal cavities, pancreas, peripheral nervous system, thyroid, trachea, acute myelocytic leukaemia, and T and B cell lymphoma^{14,15,42}. More than one hundred of these N-nitroso compounds have been tested for carcinogenicity in animals, and 75–80% of them have been found to be carcinogens¹⁴. IARC¹⁵ concluded that 11 N-nitroso compounds were carcinogenic in man. In humans, the organs thought to be more at risk from cancer are the stomach, esophagus, nasopharynx and bladder.

Human epidemiology studies reported an increase in stomach cancer rates with consumption of water with high nitrate, especially if exposed during the first ten years of life⁴³. People having low gastric acidity are more prone to gastric cancer, as it favours formation of N-nitroso compounds^{9,10,44}. Association of other cancers with nitrate ingestion are bladder cancer⁴⁵, non-Hodgkin's lymphoma⁴⁶ and colon cancer⁴⁷. It is interesting to note that patterns as observed with colon cancer have not been observed with rectal cancer. It is probably because the N-nitroso compounds, which are formed in the digestive tract, have less contact time in the rectum than in the colon⁴⁸.

Respiratory system

An increase in asthmatic attacks³⁶ is reported with high air-borne nitrate concentrations. High percentages (40–82) of cases of acute respiratory tract infection with his-

tory of recurrence have been reported in children drinking high-nitrate water⁴⁹. These findings were further substantiated⁵⁰ in animal experiments, significant changes in lungs were observed with congestion, presence of inflammatory cells, breakdown of alveoli, frequent purulent bronchial exudates, interstitial round cell infiltration and fibrosis at certain areas, when animals were fed with water containing >100 mg/l of nitrate.

Cardiovascular system

Earlier onset of hypertension has been reported with high nitrate ingestion⁵¹. In an animal study it was reported that high nitrate ingestion is associated with changes in form of small foci of inflammatory cells and fibrosis in cardiac muscles. Diffuse interstitial cellularity with pronounced degenerative foci was also noted. Thinning and dilatation of intramural coronary arteries⁵⁰ has also been reported. The changes in cardiac tissue even in animal studies are important, in view of the side/adverse effects related to the use of nitrate-containing drugs for the management of cardiac disorders and increasing drug tolerance of these nitrate-containing drugs⁵².

Gastrointestinal system

Recurrent diarrhoea has been reported with high nitrate ingestion, especially in children⁵³. It has been suggested³⁰ that increased consumption of nitrate leads to (a) increased production of nitrite, (b) enhanced absorption of sodium from the intestinal lumen, (c) excess NO^{*} generation, having vasodilatory effect and (d) increased production of O₂⁻, which will react with other cell constituents, possibly causing irreversible cell damage. These changes in enteric mucosa cause hyperemia and oedema in the enteric mucosa and later on possibly cause irreversible mucosal damage and therefore, provide high-risk conditions favourable for recurrent diarrhoea. These observations were further substantiated by animal studies which revealed the pathological changes in intestine and colon, and the changes were progressive as the nitrate content of the ingested water increased⁵³. These findings are of interest since infants and children are supplemented with ORS, which if prepared with nitrate-rich water, during diarrhoea will be an aggravating factor for nitrate toxicity¹⁹. Recurrent stomatitis⁵⁴ was another problem reported in people using high nitrate-containing drinking water. The recurrent stomatitis was well correlated with increased cytochrome *b*₅ reductase activity following high nitrate ingestion.

Abortions

Health effects associated with ingestion of nitrate-contaminated water have included stillbirth, low birth weight

and slow weight gain and even death of the animals affected⁵⁵. Spontaneous abortions have been reported in animal studies⁵⁵, as well as in humans^{39,40,56}.

Birth defects

The risk of birth defects could be due to a single high dose of nitrate early in the pregnancy that later, having profound effects on long-term foetal development. In studies on rats and hamsters multiple birth defects, including malformations of the eye, central nervous system and musculoskeletal system were observed when a single dose of *N*-ethyl-*N*-nitrosourea, a nitrosamine, was given to the mother⁵⁷. In humans, increased rates of anencephaly⁵⁸, birth defects of the central nervous system and musculoskeletal system have been reported.

Diabetes

A positive correlation between high nitrate levels in drinking water and increased incidence of type-1 diabetes was observed^{59,60}. High nitrate ingestion during pregnancy has been shown to be associated with increased incidence of type-1 diabetes in male offspring⁶¹. High nitrate ingestion causes increased production of free radicals which are toxic to pancreatic beta cells^{49,54,60}. Some studies indicated no relationship between nitrites and nitrates in drinking water and increased incidence of type-1 diabetes⁶². Hence the association of dietary nitrites with diabetes remains tenuous and further research needs to be supported.

Adrenal gland

In humans, high nitrate ingestion causes a decreased production of adrenal steroids as reflected by the decreased concentration of 17-hydroxysteroid and 17-ketosteroids in urine⁶³. Results of animal studies on rabbits also reported the same results⁶⁴.

Thyroid function and morphology

The thyroid gland contains an iodine-trapping transport mechanism which is accomplished by a membrane protein, the sodium-iodine symporter, which provides sufficient iodine substrate for hormone formation. This trapping mechanism for iodine is shared by other monovalent anions, including pertechnetate, perchlorate, thiocyanate and nitrate. It has been reported that relative potency of perchlorate for inhibiting iodine uptake is 15 and 240 times greater than that of thiocyanate and nitrate respectively, on a molar concentration basis in the serum⁶⁵. Due to inhibition of this trapping mechanism, chronic nitrate exposure causes an inhibition in the accu-

mulation of iodine in the thyroid gland. Consequently, it may result in thyroid malfunction causing higher relative risk of goiter⁶⁶ in children, more volume^{67,68} and weight^{69,70} of thyroid gland in children as well as in adults, and higher frequency of hypoechogenicity⁶⁸ of thyroid gland. Histomorphological changes reported are retention of lobular architecture, prominent vascular congestion, follicular hyperplasia, a vacuolization and an increase in the colloidal volume of the follicles⁷⁰. Investigators have shown that a low dose or short-term nitrate intake causes a decrease in thyroid radioiodine uptake⁷¹. Whereas Eskiocak *et al.*^{69,70} reported that high dose and long-term nitrate exposure results in an increase in the thyroid radioiodine uptake. These findings suggest that the effect of nitrate on thyroid iodine uptake is dose-dependent and the inhibition of thyroid iodine uptake may be stronger at higher amounts of nitrate.

Human studies⁶⁷ reported decrease in TSH levels, whereas others⁶⁸ reported an increase in TSH. A decrease in total and free T₃ and T₄ levels at high dose with long-term nitrate exposure^{69,70} has been reported, whereas some studies⁷¹ demonstrated that short-term nitrate administration may result in a significantly higher serum level of total T₃. These findings indicate that short or long-term nitrate exposure may be strongly responsible for the prominent change in thyroid hormone production.

Immunity

Studies on human immune system indicated that nitrate ingestion does not affect lymphocyte growth, but nitrite decreases proliferation of lymphocytes⁷². Fibroblast growth remains unaffected. A decreased production of Th1 cytokines (interleukin-2, interferon-gamma and tumour necrosis factor-beta), which are responsible for resistance to a variety of infectious diseases was noted. No effect on the production of the Th2 cytokine interleukin-10, which is responsible for disease susceptibility, was noted. Because nitrate/nitrite shifted the balance from a Th1 to a Th2 response in some individuals, exposure to these compounds may decrease such a person's responsiveness to infectious diseases. Animal studies also reported an immune suppression due to high nitrate ingestion⁷³.

Air pollution and nitrate toxicity

Inhalation of NO_x causes a wide variety of health and environmental impacts because of various compounds and derivatives in the family of nitrogen oxides, including nitrogen dioxide, nitric acid, nitrous oxide, nitrates and nitric oxide. It reacts to form nitrate particles, acid aerosols and contributes to formation of acid rain.

Health effects are related to levels of NO_x as well as the duration of exposure^{74,75}. Low levels of nitrogen oxides in the air irritate the eyes, nose, throat and lungs,

leading to cough and shortness of breath, tiredness and nausea. Breathing high levels of nitrogen oxides can cause rapid burning, spasms and inflammatory swelling of tissues in the throat and upper respiratory tract. High exposures may lead to pulmonary oedema, leading to hypoxemia and even death. Industrial exposure to nitrogen dioxide may cause genetic mutations, damage a developing foetus and decrease fertility in women. Industrial exposure to nitric oxide can cause unconsciousness, vomiting, mental confusion, and damage to the teeth. So far, there is no evidence that nitrogen oxides are potential carcinogens.

A review of studies published in the last decade has shown urban pollution to be an environmental cardiovascular risk factor⁷⁶. This link was significant for NO_x and PM10. A study of short-term effects of nitrogen dioxide on total, cardiovascular and respiratory mortality in 30 European cities found significant association between the two⁷⁷. Significant associations of daily changes in particle concentrations, nitrogen dioxide and carbon monoxide were found with hospitalization for respiratory diseases (COPD, pneumonia, asthma) and cardiovascular diseases⁷⁸. Exposure to indoor NO₂ at levels well below the Environmental Protection Agency (EPA) outdoor standard (53 ppb) was associated with respiratory symptoms among children with asthma. Each 20 ppb increase in NO₂ increased both likelihood of any wheeze or chest tightness and days of wheeze or chest tightness⁷⁹.

The EPA has established that the average concentration of nitrogen dioxide in ambient air in a calendar year should not exceed 0.053 parts of nitrogen dioxide per million parts of air (0.053 ppm). The Occupational Safety and Health Administration (OSHA) has set a limit of 25 ppm of nitric oxide in workplace air during an 8-h workday, 40-h work week. OSHA has also set a 15-min exposure limit of 5 ppm for nitrogen dioxide in workplace air.

Conclusion

Nitrate ingestion beyond permissible limit is toxic to human beings. The literature available on nitrate toxicity in humans is limited, except for reports documenting methaemoglobinemia in infants. Many studies indicated nitrate as a cause of cancer, but the finding is still controversial and no firm conclusions have been drawn. Other effects observed were increased infant mortality, abortions, birth defects, recurrent diarrhoea, recurrent stomatitis, early onset of hypertension, histopathological changes in cardiac muscles, alveoli of lungs and adrenal glands, recurrent respiratory tract infection in children, hypothyroidism and diabetes, and adverse effects on the immune system of the body. Recently, an adaptation system to nitrate ingestion has also been reported. This adaptation to an enzyme cytochrome *b*₅ reductase has been shown to be protective to human beings, but to a limited extent only.

Since denitrification of water is difficult and costly, it is recommended that some changes in habits and adoption of simple, preventive measures may be urgently introduced in Government campaigns, at least in high-nitrate belts. Indiscriminate use of nitrogenous fertilizers should be avoided. Promoting breast feeding up to the age of at least 6 months is an important strategy. Long-term use of antacids, H₂ receptor inhibitor or proton pump inhibitor should be avoided. If at all needed, it should be given with antioxidants. Health education system should be developed to make people aware of the toxic effects of nitrate ingestion and its prevention.

More detailed epidemiological health-related studies covering a large sample are required to provide insights to the nitrate toxicity on human beings, especially the exploration of pathophysiology, nitrate toxicity and role of free oxide radicals to yield better understanding of the various diseases caused by nitrates, and their prevention and treatment. It is highly desirable to study what can be the better acceptable standards for nitrate in drinking water and to develop an easy and cost-effective denitrification method.

Certain recommendations made by the Council on Scientific Affairs were adopted by the AMA House of Delegates, as AMA directives in 2004. (1) The AMA supports the current FDA and United States Department of Agriculture regulations, including current labelling requirements, for nitrites in food (Directive). (2) The AMA encourages continued research and surveillance of the safety of nitrite use in foods, with particular attention to its possible effects on type-1 diabetes (Directive).

1. WHO, Guidelines for drinking water quality, Geneva, 2004, vol. 1, p. 191.
2. IS-10500, Drinking water specification. Bureau of Indian Standards, New Delhi, 1995, vol. 3.
3. WHO, Health hazards from nitrate in drinking-water. Report on a WHO Meeting, Copenhagen, 5-9 March 1984, WHO Regional Office for Europe (Environmental Health Series No. 1), Copenhagen, 1985.
4. WHO, Toxicological evaluation of certain food additives and contaminants. In Forty-Fourth Meeting of the Joint FAO/WHO Expert Committee on Food Additives (JECFA), International Programme on Chemical Safety (WHO Food Additives Series 35), World Health Organization, Geneva, 1996.
5. Eisenbrand, G., Spiegelhalter, B. and Preussmann, R., Nitrate and nitrite in saliva. *Oncology*, 1980, **37**, 227-231.
6. Walker, R., The conversion of nitrate into nitrite in several animal species and man. In *Health Aspects of Nitrate and its Metabolites (Particularly Nitrite)*, Proceedings of an International Workshop, Bilthoven (The Netherlands), 8-10 November 1994, Council of Europe Press, Strasbourg, 1995, pp. 115-123.
7. Ruddell, W. S., Bone, E. S., Hill, M. J. and Walters, C. L., Pathogenesis of gastric cancer in pernicious anaemia. *Lancet*, 1978, **1**, 521-523.
8. Mirvish, S. S., Formation of N-nitroso compounds: Chemistry, kinetics, and *in vivo* occurrence. *Toxicol. Appl. Pharmacol.*, 1975, **31**, 325-351.

9. Farinati, F. *et al.*, Gastric antioxidant nitrites and mucosal lipoperoxidation in chronic gastritis and *Helicobacter pylori* infection. *J. Clin. Gastroenterol.*, 1996, **22**, 275-281.
10. Vermeer, I. T., Engels, L. G., Pachen, D. M., Dallinga, J. W., Kleinjans, J. C. and van Maanen, J. M., Intra-gastric volatile N-nitrosamines, nitrite, pH and *Helicobacter pylori* during long-term treatment with omeprazole. *Gastroenterology*, 2001, **121**, 517-525.
11. Choi, B. C. K., N-nitroso compounds and human cancer: A molecular epidemiologic approach. *Am. J. Epidemiol.*, 1985, **121**, 737.
12. Terblanche, A. P. S., Health hazards of nitrate in drinking water and possible means of denitrification. *Water SA*, 1991, **17**, 77-83.
13. RIVM, Integrated criteria document nitrate effects. Appendix to RIVM Report No. 758473012. Bilthoven, Rijksinstituut voor de Volksgezondheid en Milieuhygiëne, National Institute of Public Health and Environmental Protection (RIVM Report No. A758473012), 1989.
14. Drinking Water and Health, National Academy of Sciences, Safe Drinking Water Committee, Washington DC, 1977.
15. IARC, IARC monographs on the evaluation of carcinogenic risks of chemicals to man. Volume 17: Some N-nitroso compounds, World Health Organization, 1978.
16. El Nahas, S. M., Globus, M. and Vethamany-Globus, S., Chromosomal aberrations induced by sodium nitrite in bone marrow of adult rats and liver cells of transplacentally exposed embryos. *J. Toxicol. Environ. Health*, 1984, **13**, 643-647.
17. Wagner, D. A., Schultz, D. S., Deen, W. M., Young, V. R. and Tannenbaum, S. R., Metabolic fate of an oral dose of ¹⁵N-labeled nitrate in humans: Effect of diet supplementation with ascorbic acid. *Cancer Res.*, 1983, **43**, 1921-1925.
18. Waldman, S. A. and Murad, F., Cyclic GMP synthesis and function. *Pharmacol. Rev.*, 1987, **39**, 163-196.
19. Murray, K. F. and Christie, D. L., Dietary protein intolerance in infants with transient methemoglobinemia and diarrhoea. *J. Pediatr.*, 1993, **122**, 90-92.
20. Lowenstein, C. J., Dinerman, J. L. and Snyder, S. H., Nitric oxide: A physiological messenger. *Ann. Intern. Med.*, 1994, **120**, 227-237.
21. Smith, A. D., Datta, S. P. and Smith, G. H. (eds), Nitric oxide. In *Oxford Dictionary of Biochemistry and Molecular Biology*, Oxford University Press, Oxford, 1997, p. 451.
22. Winterbourn, C. C., McGrath, B. M. and Carrell, R. W., Reaction involving superoxide and normal and unstable haemoglobins. *Biochem. J.*, 1976, **155**, 493-502.
23. Sutton, H. C., Roberts, P. B. and Winterbourn, C. C., The rate of reaction of superoxide radical ion with oxyhemoglobin and methemoglobin. *Biochem. J.*, 1976, **155**, 503-510.
24. Bodansky, O., Methemoglobin and methemoglobin producing compounds. *Pharmacol. Rev.*, 1951, **3**, 144-195.
25. Jaffe, E. R., Methemoglobinemia. *Clin. Hematol.*, 1981, **10**, 99.
26. Hsia, C. C. W., Respiratory function of hemoglobin. *New Engl. J. Med.*, 1998, **338**, 239-247.
27. Berger, H. M., Moison, R. M. W. and Grobden, D. V. Z., Pathophysiology of respiratory distress syndrome. In *14th Recent Advances in Pediatrics* (ed. David, T. J.), Churchill Livingstone, New Delhi, 1997, 1st edn, pp. 117-119.
28. Roediger, W. E. W., Deakin, E. J., Radcliffe, B. C. and Nance, S., Anion control of sodium absorption in the colon. *Q. J. Exp. Physiol.*, 1986, **71**, 195-204.
29. Comly, H. H., Cyanosis in infants caused by nitrates in well water. *JAMA*, 1945, **129**, 112-116.
30. Cole, J. A. and Brown, C. M., Nitrate reduction to ammonia by fermentative bacteria: A short circuit in the biological nitrogen cycle. *FEMS Microbiol. Lett.*, 1980, **7**, 65-72.
31. Gupta, S. K., Fitzgerald, J. F., Chong, S. K. F., Croffie, J. M. and Garcia, J. G. N., Expression of inducible nitric oxide synthase (iNOS) mRNA in inflamed esophageal and colonic mucosa in pediatric population. *Am. J. Gastroenterol.*, 1998, **93**, 795-798.

32. Ellen, G., Schuller, P. L., Bruijns, E., Froeling, P. G. and Baadenhuijsen, H. U., Volatile N-nitrosamines, nitrate and nitrite in urine and saliva of healthy volunteers after administration of large amounts of nitrate. In *N-nitroso Compounds: Occurrence and Biological Effects* (eds Bartsch *et al.*), IARC Scientific Publ. No. 41, 1982, pp. 365–378.
33. Bartholomew, B. A. and Hill, M. J., The pharmacology of dietary nitrate and the origin of urinary nitrate. *Food Chem. Toxicol.*, 1984, **22**, 789–795.
34. Corre, W. J. and Breimer, T., Nitrate and nitrite in vegetables. In Literature Survey no. 39, Center for Agricultural Publishing Documentation, Wageningen, 1979.
35. Benz, E. J., Hemoglobinopathies, hemoglobin with altered oxygen affinity. In *Harrison's Principles of Internal Medicine 1* (eds Braunwald, E. *et al.*), McGraw-Hill, New York, 2001, 15th edn, p. 1524.
36. WHO, Nitrate, nitrite and N-nitroso compounds. In *Environmental Health Criteria 5*, Geneva, 1977.
37. Gupta, S. K., Gupta, R. C., Seth, A. K., Gupta, A. B., Bassin, J. K. and Gupta, A., Methemoglobinemia – A problem of all age groups in areas with high nitrate in drinking water. *Natl. Med. J. India*, 2000, **13**, 58–61.
38. Gupta, S. K., Gupta, R. C., Seth, A. K., Gupta, A. B., Bassin, J. K. and Gupta, A., Enzymatic adaptation of cytochrome *b₅* reductase activity and methemoglobinemia in areas with high nitrate concentration in drinking water. *Bull. WHO*, 1999, **77**, 749–753.
39. Fewtrell, L., Drinking-water nitrate, methemoglobinemia, and global burden of disease: A discussion. *Environ. Health Perspect.*, 2004, **112**, 1371–1374.
40. Center for Disease Control and Prevention, Spontaneous abortions possibly related to ingestion of nitrate-contaminated well water – LaGrange County, Indiana, 1991–1994. *Morb. Mortal Wkly Rep.*, 1996, **45**, 569–572.
41. Tannenbaum, S. R., Fett, D., Young, V. R., Land, P. D. and Bruce, W. R., Nitrite and nitrate are formed by endogenous synthesis in the human intestine. *Science*, 1978, **200**, 1487–1489.
42. Mirvish, S. S., The etiology of gastric cancer: Intra-gastric nitrosamide formation and other theories. *J. Natl. Cancer Inst.*, 1983, **71**, 631–647.
43. Xu, G., Song, P. and Reed, P. I., The relationship between gastric mucosal changes and nitrate intake via drinking water in a high risk population for gastric cancer in Moping county, China. *Eur. J. Cancer Prevent.*, 1992, **1**, 437–443.
44. Dallinga, J. W. *et al.*, Volatile N-nitrosamines in gastric juice of patients with various conditions of the gastrointestinal tract determined by gas chromatography – mass spectrometry and related to intra-gastric pH and nitrate and nitrite levels. *Cancer Lett.*, 1998, **124**, 119–125.
45. Weyer, P. J. *et al.*, Municipal drinking water nitrate level and cancer risk in older women: The Iowa women's health study. *Epidemiology*, 2001, **12**, 327–338.
46. Weisenburger, D., Environmental epidemiology of non-Hodgkins lymphoma in Eastern Nebraska. *Am. J. Ind. Med.*, 1990, **18**, 303–305.
47. Roos, A. J. D., Ward, M. H., Lynch, C. F. and Cantor, K. P., Nitrate in public water supplies and the risk of colon and rectum cancers. *Epidemiology*, 2003, **14**, 640–649.
48. Vander, A. J., Sherman, J. H. and Luciano, D. S., *The Digestion and Absorption of Food – Human Physiology*, McGraw-Hill Inc, New York, 1994, pp. 561–600.
49. Gupta, S. K., Gupta, R. C., Gupta, A. B., Seth, A. K., Bassin, J. K. and Gupta, A., Recurrent acute respiratory tract infection in areas having high nitrate concentration in drinking water. *Environ. Health Perspect.*, 2000, **108**, 363–366.
50. Gupta, S. K., Gupta, R. C., Seth, A. K., Gupta, A. B., Sharma, M. L. and Gupta, A., Toxicological effects of nitrate ingestion on cardio respiratory tissues in rabbit. *South Asian J. Prevent. Cardiol.*, 1999, **2**, 101–105.
51. Meiberg, J. B. M., Bruinenberg, P. M. and Harder, W., Effect of dissolved oxygen tension on the metabolism of methylated amines in *Hyphomicrobium X* in the absence and presence of nitrate: Evidence for aerobic denitrification. *J. Gen. Microbiol.*, 1980, **120**, 453–463.
52. USP DI, Nitrates Systemic, 1990, pp. 2033–2036.
53. Gupta, S. K., Gupta, R. C., Gupta, A. B., Seth, A. K., Bassin, J. K. and Gupta, A., Recurrent diarrhoea in areas with high nitrate in drinking water. *Arch. Environ. Health*, 2001, **56**, 369–374.
54. Gupta, S. K., Gupta, R. C., Seth, A. K., Gupta, A. B., Bassin, J. K., Gupta, D. K. and Sharma, S., Epidemiological evaluation of recurrent stomatitis, nitrates in drinking water and cytochrome *b₅* reductase activity. *Am. J. Gastroenterol.*, 1999, **94**, 1808–1812.
55. Committee on Nitrate Accumulation, Ag. Board, Division of Biology and Agriculture, National Research Council, Hazards of nitrate, nitrite, and nitrosoamines to man and livestock. In *Accumulation of Nitrate*, National Academy of Sciences, Washington DC, 1972, pp. 46–75.
56. Muhrer, M. E., Garner, G. B. and Pfander, W. H., The effect of nitrate on reproduction and lactation. *J. Anim. Sci.*, 1959, **15**, 1291–1292.
57. Druckrey, H., Ivankovic, S. and Preussmann, R., Teratogenic and carcinogenic effects in the offspring after single injection of ethylnitrosourea in pregnant rats. *Nature*, 1966, **210**, 1378–1379.
58. Croen, L. A., Todoroff, K. and Shaw, G. M., Maternal exposure to nitrate from drinking water and diet and risk for neural tube defects. *Am. J. Epidemiol.*, 2001, **153**, 325–331.
59. Van Maanen, J. M., Albering, H. J. and de Kok, T. M., Does the risk of childhood diabetes mellitus require revision of the guideline values for nitrate in drinking water? *Environ. Health Perspect.*, 2000, **108**, 457–461.
60. Kostraba, J. N., Gay, E. C., Rewers, M. and Hamman, R. F., Nitrate levels in community drinking waters and risk of IDDM. An ecological analysis. *Diabetes Care*, 1992, **15**, 1505–1508.
61. Helgason, T. and Jonasson, M. R., Evidence for a food additive as a cause of ketosis-prone diabetes. *Lancet*, 1981, **2**, 716–720.
62. Longnecker, M. P. and Daniels, J. L., Environmental contaminants as etiologic factors for diabetes. *Environ. Health Perspect. (Suppl. 6)*, 2001, **109**, 871–876.
63. Kuper, F. and Til, H. P., Subchronic toxicity experiments with potassium nitrite in rats. In *Health Aspects of Nitrate and its Metabolites (Particularly Nitrite)*. Proceedings of an International Workshop, Bilthoven (Netherlands), 8–10 November 1994, Strasbourg, Council of Europe Press, 1995, pp. 195–212.
64. Violante, A., Cianetti, A. and Ordine, A., Adrenal cortex function during subacute poisoning with sodium nitrite. *Quad. Sclavo Diagn. Clin. Lab.*, 1973, **9**, 907–920.
65. Tonacchera, M. *et al.*, Relative potencies and additivity of perchlorate, thiocyanate, nitrate, and iodide on the inhibition of radioactive iodide uptake by the human sodium iodide symporter. *Thyroid*, 2004, **14**, 1012–1019.
66. Vladeva, S., Gatseva, P. and Gopina, G., Comparative analysis of results from studies of goitre in children from Bulgarian villages with nitrate pollution of drinking water in 1995 and 1998. *Cent. Eur. J. Public Health*, 2000, **8**, 179–181.
67. Van Maanen, J. M. *et al.*, Consumption of drinking water with high nitrate levels causes hypertrophy of the thyroid. *Toxicol. Lett.*, 1994, **72**, 365–374.
68. Tajtakova, M. *et al.*, Increased thyroid volume and frequency of thyroid disorder signs in school children from nitrate polluted area. *Chemosphere*, 2006, **62**, 559–564.
69. Eskioçak, S., The effects of administration of chronic nitrate by drinking water on thyroid hormones, thyroidal radioiodine uptake and mass. Thesis in Biochemistry Speciality, Samsun, Turkey, 1995.

70. Eskiocak, S., Dundar, C., Basoglu, T. and Altaner, S., The effects of taking chronic nitrate by drinking water on thyroid functions and morphology. *Clin. Exp. Med.*, 2005, **5**, 66–71.
71. Szokeova, E. *et al.*, Effect of nitrates on active transport of iodine. *Vnitr. Lek.*, 2001, **47**, 768–771.
72. Ustyugova, I. V., Zeman, C., Dhanwada, K. and Beltz, L. A., Nitrates/nitrites alter human lymphocyte proliferation and cytokine production. *Arch. Environ. Contam. Toxicol.*, 2002, **43**, 270–276.
73. Porter, W. P., Jaeger, J. W. and Carlson, I. H., Endocrine, immune, and behavioral effects of aldicarb (carbamate), atrazine (triazine) and nitrate (fertilizer) mixtures at groundwater concentrations. *Toxicol. Ind. Health*, 1999, **15**, 133–150.
74. Committee of the Environmental and Occupational Health Assembly of the American Thoracic Society, Health effects of outdoor air pollution. *Am. J. Respir. Crit. Care Med.*, 1996, **153**, 3–50.
75. Bernstein, J. A., Alexis, N., Barnes, C., Bernstein, I. L., Bernstein, J. A. and Nel, A., Health effects of air pollution. *J. Allergy Clin. Immunol.*, 2004, **114**, 1116–1123.
76. Maitre, A., Bonnetterre, V., Huillard, L., Sabatier, P. and de Gaudemaris, R., Impact of urban atmospheric pollution on coronary disease. *Eur. Heart J.*, 2006, **27**, 2275–2284.
77. Samoli, E. *et al.*, Short-term effects of nitrogen dioxide on mortality: An analysis within the APHEA project. *Eur. Respir. J.*, 2006, **27**, 1129–1138.
78. Hinwood, A. L. *et al.*, The relationship between changes in daily air pollution and hospitalizations in Perth, Australia 1992–1998: A case-crossover study. *Int. J. Environ. Health Res.*, 2006, **16**, 27–46.
79. Belanger, K., Gent, J. F., Triche, E. W., Bracken, M. B. and Leaderer, B. P., Association of indoor nitrogen dioxide exposure with respiratory symptoms in children with asthma. *Am. J. Respir. Crit. Care Med.*, 2006, **173**, 297–303.
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