

monographic publications on *Iris* cultivation, including that of Mathew¹⁵, Kohlein¹⁶ and Service¹⁴ have no report of its subsequent cultivation in England or elsewhere after its publication by Dykes in 1913.

1. Dykes, W. R., In *The Genus Iris*, Cambridge University Press, London, 1913, vol. 134, t.31.
2. Dykes, W. R., *Gard. Chron.*, 1908, **43**, 396.
3. Biswas, K., In *Plants of Darjeeling and Sikkim Himalayas*, Superintendent Government Printing, Calcutta, 1966.
4. Hara, H., Tuyama, T., Murata, G., Kanai, H. and Togashi, M., In *Spring Flora of Sikkim Himalaya*, Hoikusha Publishing Co Ltd, Osaka, Japan, 1963.
5. Srivastava, R. C., In *Flora of Sikkim* (eds Hajra, P. K. and Verma, D. M.), Botanical Survey of India, Calcutta, 1996, vol. 1, pp. 136–137.
6. Hara, H., In *Flora of Eastern Himalaya*, University of Tokyo, Japan, 1966.
7. Hara, H., In *Flora of Eastern Himalaya (2nd Report)*, University of Tokyo Press, Japan, 1971.
8. Ohashi, H., In *Flora of Eastern Himalaya (3rd Report)*, University of Tokyo Press, Japan, 1975.
9. Hara, H., Stearn, W. T. and Williams, L. H. J., In *An Enumeration of the Flowering Plants of Nepal*, British Museum, London, 1978, **1**, 63–64.
10. Press, J. R., Srestha, K. K. and Sutton, D. A., In *Annotated Checklist of the Flowering Plants of Nepal*, The Natural History Museum, London, 2000.
11. Noltie, H., In *Flora of Bhutan*, Royal Botanic Garden, Edinburgh, 1994, vol. 3, pp. 111–121.
12. Waddick, J. W. and Zhao, Y. T., In *Iris of China*, Timber Press, Oregon, 1992.
13. Zhao, Y. T., Noltie, H. and Mathew, B., In *Flora of China* (eds Zheng-yi, W. and Raven, P.), Science Press and Missouri Botanical Garden, 2000, vol. 24, pp. 340–355.
14. Service, N., In *A Guide to Species Irises – Their Identification and Cultivation* (eds Species Group of the British Iris Society), Cambridge University Press, Cambridge, 1997, pp. 17–108.
15. Mathew, B., In *The Iris*, BT Batsford Ltd, London, 1981.
16. Kohlein, P. F., In *Iris* (English translation by M. C. Peters), Christopher Helm, London, 1987.

ACKNOWLEDGEMENTS. I thank Dr M. Sanjappa, Director, Botanical Survey of India, Kolkata and Prof. S. P. Rath, Utkal University, Bhubaneswar for guidance and suggestions, and Dr H. J. Noltie for sharing his field experience on Sikkim.

Received 1 July 2005; accepted 22 August 2005

D. D. BAHALI

*Conservation Biology and Molecular Taxonomy Laboratory,
National Botanical Research Institute,
Rana Pratap Marg,
Lucknow 226 001, India
e-mail: ddbahali@yahoo.com*

Fishmeal extract agar – a medium to inhibit swarming of *Proteus* spp.

Swarming on appropriate solid medium is characteristic of *Proteus* spp. It is the result of migration of a group of cells (swarm cells) from the edge of the developing micro-colony to an uninoculated area of the medium producing thin films of concentric rings of growth¹. The swarming of *Proteus* may make it difficult to isolate in pure culture other pathogens which may be present in clinical specimens. Several anti-swarming agents have been described. Many of these inhibit the growth of certain pathogenic bacteria¹. Fishmeal-based media were used for growing *Entamoeba histolytica*² and for the isolation and antibiotic susceptibility testing of medically important bacteria^{3,4}. We report here the use of fishmeal extract agar to inhibit swarming of *Proteus* spp., without affecting the growth of other pathogenic bacteria which may be present along with *Proteus* in clinical specimens.

Fishmeal extract agar (FMEA) was prepared as previously described². Five grams of fishmeal was boiled in 100 ml of distilled water and filtered through Whatman no. 1 filter paper. The volume was made up to 100 ml and pH was ad-

justed to 7.4. Agar (Hi Media) was added to a concentration of 2.0%, the medium was sterilized by autoclaving at 121°C for 15 min and poured into sterile petri plates. The plates were dried at 37°C for 1 h before inoculation. FMEA with sodium chloride was prepared by incorporating 0.5% sodium chloride into the former.

Clinical isolates of 25 *P. mirabilis* and 15 *P. vulgaris*, together with 40 swarming strains of *Proteus* spp. were inoculated on FMEA and nutrient agar (NA). The latter served as control. The swarming strains of *Proteus* were also inoculated on FMEA incorporated with 0.5% sodium chloride. Smears were prepared from growth on FMEA, FMEA with 0.5% sodium chloride and NA after 2 and 4 h, stained by Gram's stain and microscopically examined.

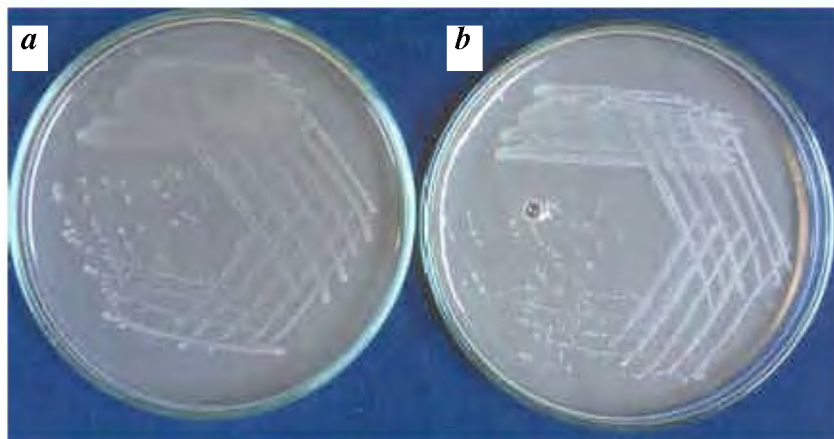
All 40 strains of *Proteus* showed swarming on NA. None of these swarmed on FMEA and FMEA incorporated with 0.5% sodium chloride (Table 1). Microscopy of growth on FMEA and FMEA with 0.5% sodium chloride after 2 and 4 h did not reveal swarm cells. Similar examination of growth on NA showed swarm cells.

The swarming of *Proteus* may make it difficult to isolate in pure culture, other bacterial pathogens in clinical specimens from polymicrobial infections. Many ways of inhibiting swarming have been described. Restricting movement of *Proteus* cells by increasing agar concentration to 3–4%; preventing formation and interfering with structure and activity of flagella by incorporating into media polyvalent H antisera; ethanol, boric acid, bile salts, detergents or by retarding the growth rate by incorporating growth inhibitors such as sulphonamides, neomycin, chloral hydrate, barbiturates, sodium azides or purine bases. Many of the anti-swarming agents are toxic and prevent the growth of delicate pathogens. Others may interfere with the colonial morphology of the organisms or lyse red blood cells in the medium, making the recognition and detection of hemolytic organisms difficult¹.

Our study showed that *Proteus* spp. does not swarm on FMEA. It is known that the omission of sodium chloride from the medium prevents spreading of *Proteus* colonies⁵. Inhibition of swarming of *Proteus* spp. on FMEA was regardless of in-

Table 1. Anti-swarmling nature of fishmeal extract agar (FMEA) compared with nutrient agar (NA)

Medium	No. of <i>Proteus</i> strains tested		No. of <i>Proteus</i> strains that showed swarming	
	<i>P. mirabilis</i>	<i>P. vulgaris</i>	<i>P. mirabilis</i>	<i>P. vulgaris</i>
NA	25	15	25	15
FMEA	25	15	Nil	Nil
FMEA with 0.5% sodium chloride	25	15	Nil	Nil

**Figure 1.** Growth of *Proteus mirabilis* on FMEA (a) and on FMEA with 0.5% sodium chloride (b) after 24 h of incubation at 37°C.

corporation of sodium chloride in the medium. However, growth was better on FMEA incorporated with 0.5% sodium chloride (Figure 1 a, b). After 24 h incubation at 37°C, colonies of both *P. mirabilis* and *P. vulgaris* on FMEA with sodium chloride were about 1–2 mm, circular, convex, smooth, translucent and non-swarmling (Figure 1 b).

Formation of multinucleate, multiflagellate, nonseptate elongated cells, 20–80 µm in length, known as swarm cells is essential for the swarming activity of *Proteus*. Swarm cell were not formed by *Proteus* spp. on FMEA. Various factors such as glutathione, viscous environment or inhibition of flagellar rotation were reported to trigger swarm cell formation, resulting in swarming¹. It appears that FMEA may be deficient in such factors which trigger swarm cell formation.

Fishmeal has already been used as a base in formulating medium for cultivation of *E. histolytica*², bacteria³, and for anti-

biotic susceptibility testing⁴. Fishmeal extract obviates the need to incorporate yeast extract, beef extract and peptone when used to grow bacteria.

Fishmeal used in FMEA is the ground cake obtained after oil extraction from pooled sardines and used as fertilizer. Fishmeal is non-hygroscopic and cost-effective (Rs 16/kg) than peptone (Rs 1600/kg).

FMEA is therefore a cost-effective, simple medium that may be effectively used for isolation in pure culture of bacterial pathogens from clinical samples containing *Proteus* spp.

1. Senior, B. W., In *Topley and Wilson's Microbiology and Microbial Infections* (eds Collier, L., Ballows, A. and Duerden, B. I.), Arnold, New York, 1998, vol. II, 9th edn, pp. 1035–1050.
2. Subbannayya, K., Babu, H., Kumar, A., Rao, K. N. A. and Shivananda, P. G., *Indian J. Microbiol.*, 1983, **23**, 126–127.

3. Anita, M., Subbannayya, K. and Shivananda, P. G., *Indian J. Pathol. Microbiol.*, 1985, **28**, 329–332.
4. Subbannayya, K., Raghunath, P. and Arjuna Rao, V., *Indian J. Exp. Biol.*, 2002, **40**, 960–962.
5. Collie, J. and Marr, W., In *Mackie and McCarty's Practical Medical Microbiology* (eds Collie, J. G. et al.), Churchill Livingstone, Edinburgh, 1989, vol. II, 13th edn, p. 115.

Received 23 June 2005; revised accepted 26 September 2005

K. SUBBANNAYYA*
J. UDAYALAXMI

Department of Microbiology,
Kasturba Medical College,
Mangalore 575 001, India.

*For correspondence.
e-mail: dr_s_kotigadde@yahoo.co.in