- 7. Tracy, R. B., Chedin, F. and Kowalczykowski, S. C., Cell, 1997, 90, 205-206.
- 8. Tracy, R. B., Baumohl, J. K. and Kowalczykowski, S. C., Genes Dev., 1997, 11, 3423-3431.
- 9. Rayssignier, C., Thaler, D. S. and Radman, M., Nature, 1989, 342, 396-401.
- 10. Beattie, K. L., Wiegand, R. C. and Radding, C. M., J. Mol. Biol., 1977, 116, 783-803.
- 11. Hsich, P., Camerini-Otero, C. S. and Camerini-Otero, D., Proc. Natl. Acad. Sci., 1992, 89, 6492-6496.
- 12. Adzuma, K., Genes Dev., 1992, 6, 1679-1694.
- 13. Karthikeyan, G., Wagle, M. D. and Rao, B. J., FEBS Lett., 1998, 425, 45-51.
- 14. Worth, Jr. L., Clark, S., Radman, M. and Modrich, P., Proc. Natl. Acad. Sci., 1994, 91, 3238-3241.
- 15. Dixon, D. A. and Kowalczykowski, S. C., Cell, 1991, 66, 361-371.
- 16. Dixon, D. A. and Kowalczykowski, S. C., Cell, 1993, 72, 87-96.
- 17. Anderson, D. G. and Kowalczykowski, S. C., Cell, 1997, 90, 77-86.
- 18. Dutreix, M., J. Mol. Biol., 1997, 273, 105-113.

Received 19 August 1998; revised accepted 1 December 1998

Lability of sex differentiation in fish

T. J. Pandian* and R. Koteeswaran

School of Biological Sciences, Madurai Kamaraj University, Madurai 625 021, India

The processes of sex determination and differentiation are labile in teleosts and are amenable for manipulations by ploidy during fertilization, hormone during hatching, temperature during the juvenile stage and other environmental or surgical factors during the adult stage.

IN oviparous teleosts early embryonic events, namely insemination, second polar body extrusion and first mitotic cleavage are manipulable and render 37 different types of ploidy induction possible; such ploidy inductions during early embryonic stages result in the production of all-male, all-female or all-sterile population¹. However, the scope for ploidy alterations to regulate sex determination is restricted to early embryonic stages alone. A large number of previous publications have attempted to precisely delineate the optimum (labile) period, during which it is possible to successfully induce ploidy^{2,3}. The process of sex differentiation in teleosts is also labile⁴, rendering hormonal induction of sex reversal possible in 37 gonochoristic species and 13 hermaphroditic species⁵; hormonal manipulations during the labile period result in the production of monosex population; again, the labile period is restricted mostly to just before and after hatching stages. Thus sex determination and differentiation in fish are labile and can

be reversed by manipulating ploidy, and hormone during fertilization and hatching stages, respectively.

A series of publications by Strussmann et al. and others' have recently documented thermal lability of sex determination in a number of teleosts; for instance, fish exposed to colder or warmer temperature from hatchling to juvenile stage lead to the production of all-female or all-male progenies^{8,9}; hence, thermal induction may serve as a third technique to regulate the sex of teleosts. A number of others, such as social and surgical factors may also induce sex reversal in adults. In many coral fish and in the freshwater Chinese paradise fish Macropodus opercularis, hierarchy and aggressive behaviour have led to the formation of a definite social organization and any manipulation to alter the social structure lead to sex reversal¹⁰⁻¹². Besides, it has long been known that gonadectomy induces sex reversal in a few teleosts¹³. For instance, female Betta splendens developed testes after ovariectomy and became functional male¹⁴. Therefore, the processes of sex determination and differentiation are labile in teleosts, rendering manipulations of ploidy during fertilization, hormone during hatching, temperature during juvenile, and surgical and social during adult stages. This communication reports the amenability of teleosts to sex regulation almost throughout life by manipulating ploidy, hormone, temperature and other selected environmental factors.

Table 1 lists selected representative species, in which sex reversal has been successfully accomplished by manipulating one or more of the following: ploidy, hormone, temperature and environmental factors. The scope for ploidy manipulation is indeed very strictly restricted to a limited period of few seconds and minutes during fertilization¹; likewise, the duration of the labile period is also very much restricted to a few minutes just before and after hatching, when the immersion technique is chosen for hormonal induction, or a few days immediately following hatching in ornamental fish¹⁵, or a few months in foodfish like carps¹⁶ and salmon¹⁷, when dietary administration is chosen for hormonal induction of sex reversal; rarely, Poecilia reticulata has been shown to be amenable for hormonal induction of sex reversal during embryogenesis, just before and after hatching and post-maturity stage^{15,18-20}; yet, the optimum period for hormonal induction of sex reversal is mostly restricted to the hatchling stage. Likewise, the optimum stage for successful sex reversal by manipulating thermal, or any other environmental factor is now shown to be restricted to the juvenile or adult stage. In more than 60% of the selected representatives species, successful sex reversal has been induced by ploidy and/or hormonal manipulation(s) by different authors. In hermaphroditic species like Monopterus albus, sex is spontaneously reversed in adults²¹; such spontaneous sex reversal during adult stage is recorded in hermaphrodites characterized by polyandrous or polygynous¹¹ social system; a manipula-

^{*}For correspondence. (e-mail: mathavan@pronet.xlweb.com)

Table 1. Lability of sex differentiation of selected fish; the lability provides scope for (1) ploidy (during and after fertilization), (2) hormonal (during and after hatching), (3) thermal (during juvenile stage) or (4) environmental (4a: spontaneous; 4b: density dependent or 4c: surgical – during adult stage) manipulations. +* = all ? brood; +** = all or brood

Family	Species	I	2	3		4		
					a	b	c	Ref.
Siluridae	Clarias gariepinus	+	+	-	_			40, 41
	C. lazera	_	+	+		_	_	8, 42
	Ictalurus punctatus	-	+	+	_	-	_	7
Anabantidae	Betta splendens	+	+	~	_	+	+	14, 15, 43, 44
	Trichogaster trichopterus	-	-	~		_	+	45
	Macropodus opercularis	-}-	+	~	_	+		46, 47
Cichlidae	Oreochromis mossambicus	+	+	+•	_	_	_	48, 49
	O. aureus	+	+	+**	_	_	_	49-51
	O. niloticus	+	+	+	_	_	_	52, 53
	Tilapia macrocephalus		_		_		+	13
Salmonidae	Oncorhynchus mykiss	+	+		_	-	_	54, 55
	O. masou	+	+		_	_		56, 57
Poeciliopsis	Poeciliopsis lucida	~-	_	+	_	_	_	58
Poecilidae	Poecilia reticulata	~	+		-		_	18, 19
Cyprinidae	Cyprinus carpio .	+	+			-	_	16, 59
	Brachydanio rerio .	+			-	_	_	60, 61
	Oryzias latipes	+	+	+*	-	-	+	62, 63
	Misgurnus anguillicaudatus	+	_	⊹ *		100	_	63, 64
Paralichthyidae	Paralichthys olivaceus	+	+	+*/+		-	_	3, 65, 66
Atherinidae	Odontesthes bonariensis	+	+	+	_		_	6, 22, 63
	Patagonia hatcheri		_	+	_	_	_	9

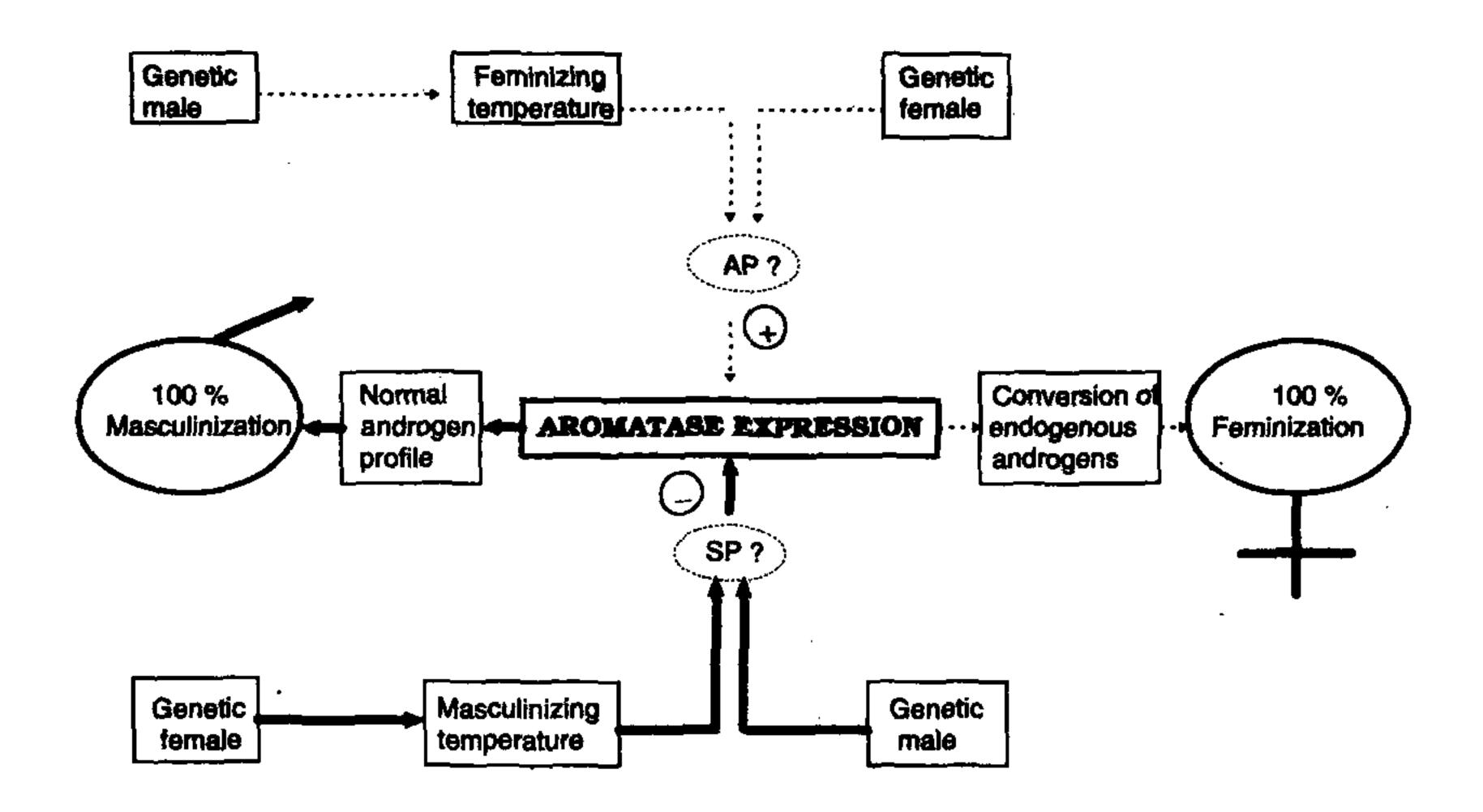


Figure 1. Suggested pathways through which thermal treatment generates all male/female progenies in fish. (AP = Activator proteins; SP = suppressor proteins; ···> Feminizing pathway; → Masculinizing pathway).

tion altering social structure of these hermaphrodites leads to sex reversal in adults of one or more individuals.

Thermal manipulations to induce sex reversal in teleosts have been explored only in recent years; it has thus far been possible to successfully accomplish sex reversal by thermal induction in about 11 species only. While documenting the thermolabile sex determination in the atherinid fish, Odontesthes bonariensis, Struss-

mann et al.^{6,9,22} who reared the hatchlings at 17°C obtained 100% females and 100% males when the juveniles were reared at 25°C for 48 days. A very similar thermal labile sex determination has also been demonstrated to occur in Clarias lazera⁸. It is not yet known how such a thermal treatment can reverse the sex of a genotypic male or female. However, it is known that such thermal treatment induces specific proteins²³, which may modify the expression and hence, the activity

of aromatase, which is a 'turn-key' enzyme in sex diferentiation; its full expression may reduce normal androgenic profile in genetic males, resulting in 100% female progeny; alternatively the reduction of its activity maintains normal endogenous androgen profile in genetic females resulting in 100% male progeny²³⁻³⁵ (Figure 1).

Although ploidy and hormonal induction of sex reversal have been widely practised now for over 20 years in many commercially important species, it has increasingly become apparent that these techniques used for sex reversal result in stunted growth (e.g. Oryzias latipes¹⁷), sterility (e.g. Poeicilia reticulata²⁰), intersexuality (e.g. P. reticulata²⁰) and/or low fecundity (e.g. P. reticulata¹⁵), when the reversed individual is fertile (see also Pandian and Sheela⁵); incidentally, these techniques also require skilled labour, costly chemicals and may involve consumer resistance, when the residual level of the administered hormone is high. Thermal manipulation to reverse the sex may prove to be the cheapest and easiest technique and can easily be practised for mass production by unskilled farmers or by using the equipment for industrial level production of monosex population^{36,37}.

It is known that the primordial germ cells of teleosts are of extra gonadal origin, and retain their bipotentiality to differentiate into male or female germ cells until sexual maturation³⁸. A large number of teleosts are known to retain this bipotentiality, even after sexual maturity³⁹. Thus the ability to retain bipotentiality, a unique characteristic of many teleosts may ultimately be responsible for the observed lability of sex differentiation throughout the life of the teleost species.

It must, however, be mentioned that not a single teleost species has thus far been subjected to all the 4 methods of manipulations to reverse the sex from fertilization to adult stage, although such a possibility is suggested by the present communication.

- 1. Pandian, T. J. and Koteeswaran, R., Hydrobiologia, 1998, 384, 167-243.
- 2. Varadaraj, K. and Pandian, T. J., Proceedings of the International Congress, Vancouver, 1988, pp. 531-535.
- 3. Tabata, K., Bull. Kyogo. Pref. Fish Exp., 1991, 28, 1-134.
- 4. Francis, C. R., Q. Rev. Biol., 1992, 67, 1-17.
- 5. Pandian, T. J. and Sheela, S. G., Aquaculture, 1995, 138, 1-22.
- 6. Strussmann, A., Moriyama, S., Hanke, E. F., Cota, C. J. C. and Takashima, F., J. Fish. Biol., 1996, 48, 643-651.
- Patino, R., Davis, K. B., Schoore, J., Uguz, C., Strussmann, C. A., Parker, N. C. and Simco, B. A., J. Exp. Zool., 1996, 276, 209-218.
- 8. Castelli, M., in Genetics and Evolution of Aquatic Organisms, (ed. Beaumont, A. R.), Chapman and Hall, London, 1994, pp. 509-519.
- 9. Strussmann, C. A., Saito, T., Usui, M., Yamada, H. and Taka-shima, F., J. Exp. Zool., 1997, 278,167-177.
- 10. Fishelson, L., Nature, 1970, 227, 90-91.
- 11. Fricke, H. and Fricke, S., Nature, 1977, 266, 830-832.
- 12. Koteeswaran, R. and Pandian, T. J., 1999, unpublished observations.

- 13. Levy, M. and Aronson, L. R., Anat. Rec., 1975, 122, 39-55.
- 14. Lowe, T. P. and Larkin, J. R., J. Exp. Zool., 1975, 191, 25-32.
- 15. Kavumpurath, S. and Pandian, T. J., Indian J. Exp. Biol., 1993, 31, 16-20.
- 16. Ali, P. K. M. M. and Rao, G. P. S., Aquaculture, 1989, 76, 157-167.
- 17. Yamazaki, F., J. Fish Res. Bd. Canada, 1976, 33, 948-958.
- 18. Kavumpurath, S. and Pandian, T. J., Aquaculture, 1993, 116, 83-89.
- 19. Kavumpurath, S. and Pandian, T. J., Aquaculture, 1993, 116, 183-186.
- 20. Takahashi, H., Bull. Fac. Fish, Hokkaido Univ., 1975, 26, 223-234.
- 21. Chan, S. T. H., in *Handbook of Sexology* (eds Money, J. and Musaph, H.), Elsevier, Amsterdam, 1977, pp. 91-105.
- 22. Strussmann, C. A., Ng, B., Takashima, F. and Oshiro, T., Prog. Fish Cult., 1993, 55, 83-89.
- 23. Usha, K. S. and Swamynathan, S. K., J. Biosci., 1996, 21, 103-121.
- 24. Desvages, G. M., Girondot, M. and Pieau, C., Gen. Comp. Endocrinol., 1993, 92, 54-61.
- 25. Fitzpatrick, M. S., Gale, W. I. and Schreek, C. B., Gen. Comp. Endocrinol., 1994, 95, 399-408.
- 26. Jeyasuria, P., Roosenburg, W. M. and Place, A. R., J. Exp. Zool., 1994, 270, 95-111.
- 27. Pieau, C. Girondot, M., Richard-Mercier, G., Desvages, G. M., Dorizzi, M. and Zabroski, P., J. Exp. Zool:, 1994, 270, 86-94.
- 28. Piferrer, F., Zanuy, S., Carrilo, M., Solar, I. I., Devlin, R. H. and Donaldson, E. M., J. Exp. Zool., 1994, 270, 255-262.
- 29. Rhen, T. and Lang, J. W., Gen. Comp. Endocrinol., 1994, 96, 243-254.
- 30. Smith, C. A. and Joss, J. M. P., Gen. Comp. Endocrinol., 1994, 93, 232-235.
- 31. Smith, C. A. and Joss, J. M. P., J. Exp. Zool., 1994, 270, 219-224.
- 32. Wibbels, T. and Crews, D., J. Endocrinol., 1994, 141, 295-299.
- 33. Chardard, D., Desvages, G., Pieau, C. and Dournon, C., Gen. Comp. Endocrinol., 1995, 99, 100-107.
- 34. Crews, D., Cantu, A. R., Bergeron, J. M. and Rhen, T., Gen. Comp. Endocrinol., 1995, 100, 119-127.
- 35. Fitzpatrick, M. S., Gale, W. L., Slater, C. H. and Schreck, C. B., in Proceedings of the Fifth International Symposium on the Reproductive Physiology of Fish (eds Goetz, F. and Thomas, P.), Fish Symp'95, Austin, Texas, 1995, p. 308.
- 36. Benfey, T. J., Bosa, P. G., Richardson, N. L. and Donaldson, E. M., Aquacult. Eng., 1988, 7, 147-154.
- 37. Recoubratsky, A. V., Gomelsky, B. I., Emelyanov, O. V. and Pankratyeva, E. V., Aquaculture, 1992, 108, 13-19.
- 38. Reinboth, R., Differentiation, 1983, 23, S82-S86.
- 39. Reinboth, R., Environ. Biol. Fish, 1988, 28, 249-259.
- 40. Hurk, R. V., Richter, C. J. J. and Dommerholt, J. J., Aquaculture, 1989, 83, 179-191.
- 41. Volckaert, F. A. M., Galbusera, P. H. A., Hellemant, B. A. S., Vanderheute, C., Vanstaen, D. and Ollivier, F., Aquaculture, 1994, 128, 221-233.
- 42. Liu, S., Yao, Z. and Wang, Y., J. Exp. Zool., 1996, 276, 432-438.
- 43. Kavumpurath, S. and Pandian, T. J., *Isr. J. Aquacult.*, 1992, 44, 111-119.
- 44. Eberhard, S., Zeitsch. Indukdt. Abstam. Vererbungs, 1943, 81, 363-373.
- 45. Johns, L. S., Liley, N. R. and Seghens, B. H. (cited in Hoar, W. S., 1969), in *Fish Physiology* (eds Hoar, W. S. and Randal, D. J.), Academic Publishers, New York, 1969, pp. 1-72.
- 46. Gervai, J. and Csanyi, V., Theor. Appl. Genet., 1984, 68, 481-485.

- 47. Minu, N., MSc, Dissertation, School of Biological Sciences, Madurai Kamaraj University, Madurai, 1995.
- 48. Varadaraj, K., Ph D Thesis, Madurai Kamaraj University, Madurai, 1990.
- 49. Mair, G. C., Beardmore, J. A. and Skibinski, D. O. F., in Proceedings of Second Asian Fisheries Forum (eds Hirano, R. and Hanyu, I.), Tokyo, 1990, pp. 555-558.
- 50. Melard, C., Aquaculture, 1995, 130, 25-34.
- 51. Desperz, D., Melard, C. and Philippart, J. C., Aquaculture, 1995, 130, 35-41.
- 52. Kim, D. S., Chei, Y. H., Noh, C. H. and Nam, Y. K., J. Aquacult., 1995, 8, 295-306.
- 53. Baroiller, J. F., Chourrout, D., Fostier, A. and Jalabert, B., J. Exp. Zool., 1995, 273, 216-223.
- 54. Chourrout, D. and Quillet, E., *Theor. Appl. Genet.*, 1982, 72, 633-636.
- 55. Lincoln, R. F. and Scott, A. P., Aquaculture, 1983, 30, 375-380.
- 56. Piferrer, F., Benfey, T. J. and Donaldson, E. M., Aquacult. Living Res., 1994, 7, 127-131.
- 57. Piferrer, F., Benfey, T. J. and E. D. Donaldson, E. M., J. Fish Biol., 1994, 45, 541-553.

- 58. Sullivan, J. A. and Schultz, R. J., Evolution, 1986, 40, 152-158.
- 59. Gomelsky, B. I., Emelyanov, O. V. and Recoubratsky, A. V., Aquaculture, 1992, 106, 233-237.
- 60. Kavumpurath, S. and Pandian, T. J., Aquacult. Fish. Manag., 1990, 21, 299-306.
- 61. Schwark, H., Aquaculture, 1993, 112, 25-37.
- 62. Naruse, K., Ijiri, K., Shima, A. and Egami, H., J. Exp. Zool., 1985, 236, 335-341.
- 63. Strussmann, C. A. and Patino, R., in Proceedings of the Fifth International Symposium on Reproductive Physiology of Fish (eds Goetz, F. W. and Thomas, P.), Austin, 1995, pp. 153-157.
- 64. Matsubara, K., Arai, K. and Suzuki, R., Aquaculture, 1995, 131, 37-48.
- 65. Abata, K., Fish Sci., 1995, 6, 199-201.
- 66. Yamamoto, E. and Masutani, R., Yoshoku, 1990, 27, 80-85.

ACKNOWLEDGEMENT. We gratefully appreciate the Indian Council of Agricultural Research, New Delhi for financial support.

Received 11 September 1998; revised accepted 20 November 1998

Influence of eyestalk ablation and 5-hydroxytryptamine on the gonadal development of a female crab, *Paratelphusa hydrodromous* (Herbst)

M. G. Ragunathan* and A. Arivazhagan

Department of Zoology, Post-Graduate Extension Centre, University of Madras, Vellore 632 004, India

The histology of the ovaries shows polymorphic oocytes in the stage I ovaries and much enlarged in the eyestalk-ablated saline-treated ovaries. The ovarian index has significantly increased in the eyestalk-ablated 5-hydroxytryptamine (5-HT)-treated crabs. The histology of the ovaries of the eye-stalk-ablated 5-HT-treated crabs shows much reduced nucleus and more cytoplasmic contents. The staining intensity is more. Histological study of the spermatheca at stage I reveals fused constituent layers whereas eyestalk-ablated 5-HT-treated crabs shows spermatheca having distinct constituent layers and the cross-section of the spermatheca shows rich secretory substances in the lumen and the staining intensity is more. The spermathecal index is significantly increased in the eyestalk-ablated 5-HT-treated crabs.

AFTER the discovery of gonad inhibiting hormone principles in the eyestalk of *Palaemon serratus* by Panouse¹, various groups of crustaceans have been shown to possess such inhibiting hormones in the X-organ, sinus gland complex². The surgical removal of the eyestalk in crustaceans in order to promote ovarian maturation has

*For correspondence: No. 3, Vinayagam Street, Velappadi, Vellore 632 001, India

been reported by many workers³⁻⁵. Eastman-Reks and Fingerman⁶ have stated that in general, the removal of both eyestalks in decapod crustaceans induces precocious yolk deposition and vitellogenesis in the ovary. In contrast, injection of eyestalk extracts or partially purified ovary inhibiting hormones results in the inhibition of ovarian maturation.

Richardson et al.⁷ have reported that the administration of 1.25×10^{-9} to 1.25×10^{-7} mole/crab of 5-hydroxytryptamine (5-HT) has showed an increased dose-dependant ovarian development in the fiddler crab, Uca pugilator found in the Florida coast, USA. Sarojini et al.⁸ have reported that 5-HT exerts an indirect stimulatory effect on the testis by triggering the release of gonad-stimulating hormone in U. pugilator.

Recently, Sharmila⁹ has reported that the administration of biogenic amines, namely 5-HT, octopamine and dopamine on the stage I female crab, *U. pugilator* exhibited interesting results. The crabs treated with 5-HT showed significant increase in the gonadal indices, in contrast to those who showed a significant decrease in the gonadal indices when treated with octopamine and dopamine.

From the literature cited above it is clear that studies have been carried out on the eyestalk ablation, treatment of various pharmacological drugs in affecting the reproduction in crustaceans. However, no work has been carried out in studying the combined action of eyestalk ablation and 5-HT treatment especially in a freshwater crab. Hence, the present work has been undertaken to study the concurrent effects of eyestalk ablation and 5-HT on the ovarian and spermathecal development of a female, freshwater crab, Paratelphusa hydrodromous (Herbst).

Paratelphusa hydrodromous used in the present study were collected from a lake in Thellur village which is