

Figure 4. Electron impact-mass spectra of (a) Compound I (2,6,10-dodecatrien-1-ol,3,7,11-trimethyl (Z, E) and (b) standard for compound I isolated from male preputial gland.

opposite sexes, 3 animals were used in each set for behaviour analysis. Fresh samples were used in each trial. The behaviour was assessed for 15 min with these compounds and the solvents alone as control. The responders were of the same and the opposite sex. The time taken for visiting each fraction was recorded and subjected to statistical analyses.

The extracted male preputial gland contains 2,6,10-dodecatrien-1-ol,3,7,11-trimethyl (Z, E), di-n-octyl phthalate and 1,2,benzenedicarboxylic acid, diisooctyl ester (Figures 1-4). These chemical substances have their unique functions. The first two compounds are involved in the attraction of the opposite sex and the third compound is involved in the attraction of the same sex (Table 1). Thus, the preputial gland of male rat contains three different compounds with distinct social functions.

- 1. Dominic, C. J., Ad. Bios., 1986, 4, 71-86.
- 2. Balakrishnan, M. and Alexander, K. M., Indian Rev. Life Sci., 1985, 5, 277-313.
- 3. Kannan, S. and Archunan, G., Rev. Clenc. Biomed., 1997, in press.
- 4. Asa, C. S., Mech, L. D. and Seal, U. S., Anim. Behav., 1984, 33, 1034-1036.
- 5. Kannan, S. and Archunan, G., Acta Physiologica, 1997, in press.
- 6. Thody, A. J. and Dijkstra, H., J. Endocrinol., 1978, 77, 397-403.
- 7. Karl, S. O., Miller, V. and Hoffmann, D. M., J. Mamm., 1992, 73, 299-302.
- 8. Brouette-Lahlou, I., Amouroux, R., Chastrette, F., Cosnier, J., Stoffelsma, J. and Vernet-Maury, E., J. Chem. Ecol., 1991, 17, 1343-1354.
- 9. de Catanzaro, D., Zacharias, R. and Muir, C., J. Reprod. Fertil., 1996, 106, 269-274.
- 10. Andreolini, F., Jemiolo, B. and Novotny, M., Experientia, 1987, 43, 998-1002.
- 11. Brahmachary, R. L., Podder-Sarkar, M. and Dutta, J., Nature, 1990, 344, 26.
- 12. Rasmussen, L. E. L., Lee, T. D., Roelofs, W. L., Zhang, A. and Daves, G. D., *Nature*, 1996, 379, 684.

- 13. Tannoudji, J. C., Einhorn, J. and Signoret, J. P., Physiol. Behav., 1994, 56, 955-961.
- 14. Ferkin, M. H. and Seamon, J. O., Can. J. Zool., 1987, 65, 2931-2937.

ACKNOWLEDGEMENTS. We thank Bharathidasan University for providing facilities and SPIC Science Foundation, Chennai for their contribution in the analytical work. We gratefully acknowledge CSIR for financial assistance.

Received 10 November 1997; revised accepted 12 February 1998

Effect of chronic atenolol treatment on responses to noradrenaline and terbutaline in isolated ventricle from hypertensive and hyperthyroid rats

Praveen Bhugra and O. D. Gulati*

Department of Pharmacology, Pramukhswami Medical College, Karamsad 388 325, India

The present investigation was undertaken to study the effect of chronic treatment with atenolol on noradrenaline and terbutaline in hypertensive and hyperthyroid rats. The maximum increase in force of contraction of ventricle by noradrenaline in control preparations chronically treated with atenolol was reduced, while that produced by terbutaline was not affected. The pD₂ values of both the agonists were increased. In DOCA-saline hypertensive preparations, chronic treatment with atenelol did not produce any change in noradrenaline-induced positive inotropic responses; however, the maximal response to terbutaline was reduced. While the pD2 value of noradrenaline was decreased, that of terbutaline was increased. Chronic treatment with atenolol in hyperthyroid rats did not produce any significant effect on the maximal contractions of ventricle with either noradrenaline or terbutaline. However, the pD2 value of noradrenaline was reduced while that of terbutaline was unaffected. It is concluded that the beneficial effects of atenolol in hypertension and hyperthyroidism may be related to reduction in the number of beta-receptors in the heart.

BETA₁- and beta₂-adrenoceptors co-exist in the hearts of most species, with the beta₁-adrenoceptor predominating functionally under normal physiological conditions¹⁻³. Chronic administration of agonist or antagonist receptor ligands results in decrease (down-regulation) and increase (up-regulation) respectively in ligand-binding

^{*}For correspondence. Present address: Ambalal Sarabhai Enterprises, Wadi Wadi, Baroda 390 007, India.

densities. Prolonged treatment with beta-receptor blocking agent, propranolol, can lead to an increase in the beta-receptor number in various tissues, which may contribute to the development of withdrawal symptoms if discontinuation of drug therapy is abrupt⁵. However beta-adrenoceptor up-regulation is not an automatic consequence of antagonist exposure and in some systems beta-adrenoceptor decreases⁶. This downregulation may contribute to the pharmacological activity of these agents, but the mechanism of action remains to be defined⁷. Cardiac beta-adrenoceptor function may be altered in hypertension⁸. The decrease in betafunctions response may be attributed partly to the downregulation of beta₁-adrenoceptors in hypertension. Little is known about functional beta2-adrenoceptors in hypertension⁹. Clinically hyperthyroidism may be associated with systolic hypertension. The intention of the present study was to see if the hypertension or hyperthyroidstate-induced changes in responses to agonists could be reversed by chronic treatment with atenolol.

Male Wistar rats (250-350 g) were sacrificed by a sharp blow on the head and cutting of the neck blood vessels. The heart was rapidly removed and placed in a petri dish containing oxygenated, prewarmed (35°C) Ringer Locke physiological salt solution of the composition (mM): NaCl – 15.4; KCI – 5.6; CaCl₂ – 2.2; NaHCO₃ – 6.0; glucose – 11.1. The solution always contained guanethidine $(1 \times 10^{-5} \text{ M})$ and atropine $(1 \times 10^{-6} \,\mathrm{M})$. The free wall of the right ventricle was excised and the intact right ventricle was suspended in a 30 ml organ bath containing Ringer Locke physiological salt solution $(35 \pm 0.5^{\circ}C, bubbled with oxygen, 1 g$ tension). After equilibration for 45 min, during which the solution was changed every 15 min, the ventricle was driven by square wave pulses (5 PPS, 5 ms 1 ms delay, supramaximal voltage 50 V). The electrodes were attached to stimulator research model SS-48 (Recorders and Medicare System, Chandigarh). Isometric tension was recorded using the force displacement transducer (Ft 1312).

Concentration-response curves of noradrenaline and terbutaline were obtained non-cumulatively on a 15 min cycle. The maximal increase in contractile force of ventricle for each concentration of noradrenaline and terbutaline was measured.

Groups of 5-10 normal or 3-4 hypertensive rats received similar chronic treatments with atenolol (3 mg/kg) dissolved in triple-glass-distilled-water and administered orally via a Ryles tube once daily for 28 days¹¹. Another group of rats received simultaneously the same dose of L-thyroxine (0.75 mg/kg in 0.001 N NaOH in 0.9% NaCl; subcutaneously) and atenolol (3 mg/kg dissolved in triple-glass-distilled-water orally via a Ryles tube) once daily for 7 days.

Male albino rats weighing about 100 g were kept on a diet high in sodium chloride and drinking water was

replaced by 2% sodium chloride solution ad lib. When the rats attained a weight of about 250 g, they were given deoxycorticosterone acetate (DOCA) dissolved in sesame seed oil in a dose of 10 mg/kg, subcutaneously twice weekly for 42 days¹².

To check whether hypertension had been produced by the DOCA-saline treatment schedule, blood pressure of rats was recorded¹³. Following confirmation of induction of hypertension, groups of 3-4 rats received chronic treatment with drugs as discussed above.

Hyperthyroidism was induced by subcutaneous injection of 0.75 mg/kg L-thyroxine sodium in alkaline saline solution (0.001 N NaOH in 0.9% NaCl) daily for 7 days¹⁴.

Rectal temperature and heart weights of L-thyroxine-treated animals were recorded before the start of the treatment and before sacrifice. Serum was prepared from blood samples collected from the common carotid artery of rat exsanguinated on the day of the experiment and stored at -20°C. Total serum thyroxine (T₄), tri-iodothyronine (T₃) and thyroxine stimulating hormone (TSH) levels were determined by enzyme immunoassay (EIA) with a commercially available *in vitro* 'diagnostic kit' (Bio Me'rieux, France) on semi-autoanalyser (SEAC CH-100; Ames marketed by Miles India, Baroda).

Microscopic examination of the ventricular tissue was performed according to the method of Helper¹⁵.

Noradrenaline (± arterenol; NA); atropine sulphate and guanethidine (1-[2-guanidinoethy1] octa hydroazocine) monosulphate were obtained from Sigma. Terbutaline was obtained form Astra-IDL. The following drugs were received as free gifts; atenolol (Cadila, Ahmedabad); deoxycorticosterone acetate (Infar, Bombay); L-thyroxine sodium (Glaxo, Calcutta).

Polyethylene glycol 400 (E. Merck, Bombay), glycerine I. P. (Metro, Wadhwan City), sesame seed oil (Ahmed Mills, Bombay) were obtained and used as solvents of the drugs.

Only one agonist was used for getting concentration-response curve in a given preparation. Contractile force was expressed as gram of tension developed. The pD₂ values were determined from the EC₅₀ (ref. 16). The results are expressed as mean \pm SEM and analysed by the Student's t test for obtaining the level of significance¹⁷.

Noradrenaline $(3.51 \times 10^{-7} \text{ M} \text{ to } 1.4 \times 10^{-6} \text{ M})$ and terbutaline $(3.03 \times 10^{-6} \text{ M} \text{ to } 9.11 \times 10^{-6} \text{ M})$ produced concentration-dependent increase in the force of contraction in electrically driven isolated right ventricle. The maximum increase in force of contraction by noradrenaline in control preparations chronically treated with atenolol was found to be reduced significantly (P < 0.01). The response to terbutaline was, however, not altered significantly. The pD₂ values of both the agonists were increased (P < 0.01) (Table 1).

Table 1. Effect of atenolol on the maximal contractile force and pD2 values obtained in rat isolated ventricle with noradrenaline and terbutaline in control, DOCA-saline hypertensive and hyperthyroid rats

Treatment group	Noradrenaline			Terbutaline		
	Maximum contractile force (g)	n	pD ₂ value	Maximum contractile force (g)	n	pD ₂ value
Control	4.07 ± 0.19	6	6.33 ± 0.04	0.81 ± 0.14	5	5.26 ± 0.01
Chronic atenolol	$2.79 \pm 0.11*$	5	6.94 ± 0.01*	0.99 ± 0.09^{NS}	3	$6.32 \pm 0.02*$
DOCA-saline	3.96 ± 0.19^{NS}	3	$6.90 \pm 0.02*$	2.07 ± 0.01*	3	$5.40 \pm 0.06 *$
DOCA-saline + chronic atenolol	4.02 ± 0.08^{NS}	3	6.45 ± 0.04*	1.74 ± 0.10**	3	5.74 ± 0.03*
L-thyroxine	$2.01 \pm 0.16*$	3	$7.00 \pm 0.00 *$	$1.74 \pm 0.11*$	3	5.55 ± 0.07*
L-thyroxine + chronic atenolol	2.07 ± 0.00^{NS}	3	6.32 ± 0.00*	1.92 ± 0.03^{NS}	3	5.66 ± 0.06^{NS}

Values are listed as ± SEM

Table 2. Effect of chronic treatment with atenolol on various somatic parameters measured following altered thyroid state in rats

		Treatment group		
Variable	Control	L-thyroxine	L-thyroxine + atenolol	
Body weight (g)	250.00 ± 0.00	216.66 ± 10.54*	250.00 ± 12.91**	
Rectal temperature in °C	37.81 ± 0.03	39.56 ± 0.15*	$38.34 \pm 0.11*$	
Heart weight (mg 100 g ⁻¹ body wt.)	362.00 ± 7.00	$594.00 \pm 1.80*$	518.00 ± 29.2*	
Serum thyroxine (T ₄) (mcg %)	11.3 ± 0.30	$38.26 \pm 0.35*$	36.33 ± 1.43^{NS}	
Serum tri-iodothyronine (T ₃) (ng/ml)	1.23 ± 0.08	8.68 ± 0.50*	$7.76 \pm 0.10**$	
Serum TSH (µ/ml)	3.83 ± 0.03	$0.24 \pm 0.02*$	0.10 ± 0.00^{NS}	

Values are listed as ± SEM

In DOCA-saline hypertensive preparations, noradrenaline-induced increase in force of contraction was not the maximal contraction with noradrenaline and inaltered significantly, while terbutaline showed a significant increase compared to control. The pD₂ values of both the agonists showed an increase compared to the control. Chronic treatment with atenolol did not produce any change in noradrenaline-induced positive inotropic responses, however, the maximum positive inotropic effect of terbutaline was reduced (P < 0.05) compared to DOCA-saline control. The pD₂ value of noradrenaline showed a decrease while pD₂ value of terbutaline showed an increase (P < 0.01) (Table 1).

Hyperthyroid state in rats produced significant increase in heart weight and hyperthermia (Table 2). Serum T₃ and T₄ levels were increased while TSH level was reduced significantly (P < 0.01). Chronic treatment with atenolol produced significant (P < 0.01) reduction in the heart weight and rectal temperature, without affecting T₄ and TSH levels. There was, however, slight but significant decrease in T₃ level after atenolol treatment (Table 2).

Chronic treatment with L-thyroxine reduced (P < 0.01)creased that with terbutaline; the pD2 values were increased (P < 0.01) with both the agonists. Chronic treatment with atenolol in hyperthyroid rats did not produce any significant effect on the maximal contractions of ventricle with noradrenaline and terbutaline. However the pD₂ value of noradrenaline was reduced (P < 0.01), while that of terbutaline was unaffected (Table 1).

Microscopic examination of stained ventricular tissues of DOCA-saline-treated rats showed focal aggregates of inflammatory cells (b). Atenolol treatment had no effect in some (cc₁) and restored the profile in others (cd₂) (Figure 1).

The results confirm the co-existence of beta₁- and beta2-adrenoceptors in rat ventricle. This also strongly supports the view that in the rat right ventricle, as in human hearts, beta2-adrenoceptors can mediate a positive inotropic effect, but these effects are smaller compared to beta₁-adrenoceptor-mediated effects¹⁸.

^{*}P < 0.01 and **P < 0.05, significant difference; NS, no significant difference. Comparisons are made between (i) chronic atenolol/DOCA-saline/L-thyroxine and control (ii) chronic DOCA-saline and chronic DOCA-saline + atenolol (iii) L-thyroxine and chronic L-thyroxine + atenolol.

^{*}P < 0.01 and **P < 0.05, significant difference; NS, no significant difference. Comparisons are made between (i) chronic L-thyroxine and control (ii) chronic L-thyroxine treatment and chronic L-thyroxine + atenolol treatment.

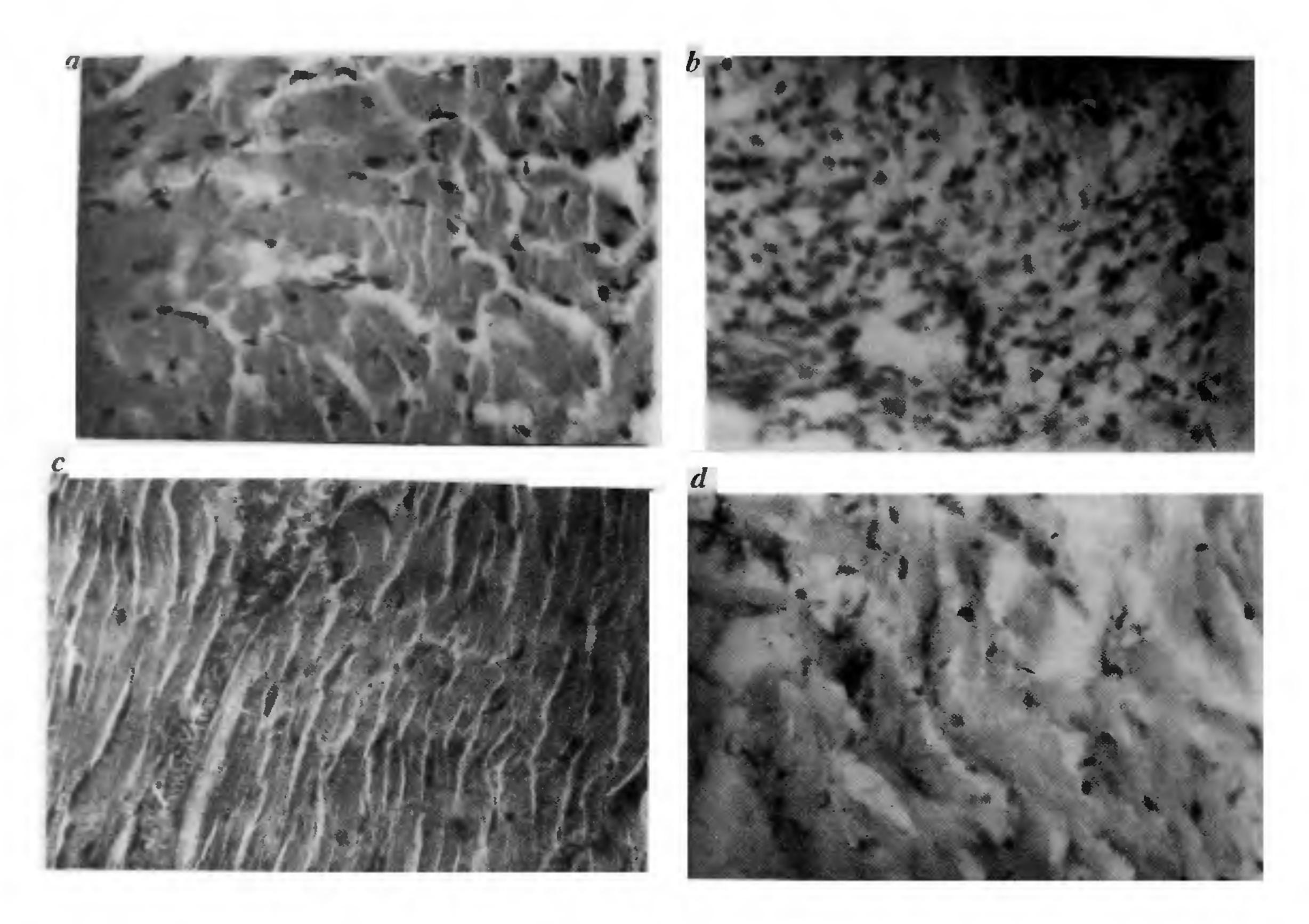


Figure 1. Photomicrograph (\times 47) of the cut section of the right ventricle from (a), normotensive rat depicting normal structure; (b), rat chronically treated for 42 days with DOCA-saline. The photomicrograph depicts focal aggregates of inflammatory cells; (c, d), in rats chronically treated for 42 days with DOCA-saline and then with atenolol for 28 days. The photomicrograph depicts (c) congestion of vessels (d) reversal of pathology to normal.

Atenolol, at a concentration which blocks beta₁-adrenergic activity, results in virtually no beta₂-antagonism^{19,20}. In the present study chronic atenolol treatment increased the pD₂ values of both noradrenaline and terbutaline, suggesting an increase in the beta-adrenoceptor density. The decrease in the maximal response with noradrenaline and no effect on the response to terbutaline may be due to down-regulation of the beta₁-receptors with compensatory up-regulation of the beta₂-receptors.

In hypertensive preparations there was an increase in the pD₂ value of noradrenaline, implying increased beta-adrenoceptor density; the maximal response was not changed. However with terbutaline, there was an increase in both the pD₂ value and the maximal response, suggesting an increase in the beta₂ receptors and increased calcium influx²¹. Chronic treatment with atenolol reduced the pD₂ value of noradrenaline with no change in the maximal contractile force of ventricle, however, the pD₂ value of terbutaline was increased with a decrease in the maximal contraction, possibly due

by interfering with transmitter turnover²² or uptake²³ or both. Reduction or reversal by atenolol of the process of arterial hypertrophy or hyperplasia associated with hypertension may be an alternate possibility²⁴. The restoration of tissue damage detected microscopically in some atenolol-treated preparations from hypertensive rats supports the above suggestion. Further work with specific radioligands would be necessary to confirm this suggestion.

Chronic L-thyroxine treatment increased the pD₂ value of noradrenaline and reduced the maximal response, implying increased beta-adrenoceptor density and decreased calcium influx. However with terbutaline, there was significant increase in the pD₂ value together with increased calcium influx. Simultaneous chronic treatment with L-thyroxine and atenolol decreased the pD₂ value of noradrenaline with no change in the pD₂ value of terbutaline and in the maximal contractile force of ventricle, suggesting that this may be due to a slight reduction in the beta₁-adrenoceptor with compensatory

increase in the beta₂-receptors. Apart from this, the significant somatic changes seen in rats treated simultaneously with L-thyroxine and atenolol, specially the heart weight and T₃ levels, may have played some vital role in the decrease of the affinity (pD₂ value) of adrenoceptors to noradrenaline.

It is concluded that the beneficial effects of atenolol in hypertension and hyperthyroidism may be related to the reduction in the number of beta₁-adrenoceptors in heart.

- 1. Juberg, E. N., Minneman, K. P. and Abel, P. W., Naunyn-Schmiedeberg's Arch. Pharmacol., 1985, 330, 193-202.
- 2. Molenaar, P. and Summers, R. J., J. Pharmacol. Exp. Ther., 1987, 241, 1041-1047.
- 3. Brodde, O-E., Karad, K., Zerkowski, H-R., Rohm, N. and Reidemaster, J. C., Circ. Res., 1983, 53, 752-758.
- 4. Hollenberg, M. D., Trends Pharmacol. Sci., 1985, 6, 299-302.
- 5. Lefkowitz, R. J., Caron, M. G. and Stiles, G. L., N. Engl. J. Med., 1984, 310, 1570-1579.
- Hughes, R. J., Mahan, L. C. and Insel, P. A., Circ. Res., 1988,
 63, 279-285.
- 7. Ferrante, J. and Triggle, D. J., *Pharmacol. Rev.*, 1990, **42**, 29-44.
- 8. Feldman, R. D., Can. J. Physiol. Pharmacol., 1987, 65, 1666-1672.
- 9. Michel, M. C., Brodde, O-E. and Insel, P. A., Hypertension, 1990, 16, 107-120.
- 10. Covin, J. M. and Berman, D. A., J. Pharmacol., 1959, 125, 137-148...
- Pasnani, J. S., Hemavati, K. G., Gulati, O. D. and Rajani, A. P.,
 J. Ethnopharmacol., 1988, 24, 287-302.
- 12. Selye, H., Hall, C. E. and Rowley, E. M., J. Can. Med. Assoc., 1943, 49, 88-92.
- 13. Ghosh, M. N., in Fundamentals of Experimental Pharmacology, Scientific Book Agency, Calcutta, 1984, 2nd edn, pp. 130-134.
- 14. Threatte, R. M., Barnery, C. C., Baker, S. P. and Fregly, M. J., Clin. Exp. Pharmacol. Physiol., 1983, 10, 101-114.
- 15. Helper, O. E., in Manual of Clinical Laboratory Methods (ed. Thomas, C. C.), Springfield, Illinois, 1950, pp. 336-339.
- 16. Ariens, E. J., in *Molecular Pharmacology*, Academic Press, New York, 1964, vol. I, pp. 153-156.
- 17. Snedecor, G. W. and Cochran, W. G., in Statistical Methods, Oxford and IBH Publishing Co., New Delhi, 1967, 6th edn, pp. 59-61.
- 18. Brodde, O. E., Pharmacol. Rev., 1991, 43, 203-242.
- 19. Hiatt, W. R., Wolfel, E. E., Stoll, S., Nies, A. S. and Zerbe, G. O., Clin. Pharmacol. Ther., 1985, 37, 2-6.
- 20. Klausner, M. A., Ventura, D. F., Coelho, J., Mullane, J. F. and Irwin, C., J. Clin. Pharmacol., 1988, 28, 495-504.
- 21. Bhugra, P. and Gulati, O. D., *Indian Pharmacol.*, 1996, 28, 77-83.
- 22. Alexandre, J. H. and Chevillard, C., Br. J. Pharmacol., 1980, 69, 35-40.
- 23. Street, J. A. and Walsh, A., Eur. J. Pharmacol., 1985, 102, 315-324.
- 24. Drapper, A. J., Kingsburg, M. P., Redfern, P. H. and Todd, M. H., J. Auton. Pharmacol., 1992, 12, 89-96.

Received 15 April 1997; revised accepted 10 February 1998.

Octopamine titer in the circulating fluid of tropical tasar silkworm,

Antheraea mylitta Drury

(Lepidoptera: Saturniidae) and its response to injected estrogen during critical phase of diapause termination

Anathbandhu Chaudhuri*, N. Krishnan*, Pradip Kumar Sarkar, Ashok Kumar Sinha**, S. S. Sinha** and Arun Kumar Ray

*Central Sericultural Research and Training Institute, Berhampore 742 101, India

Bose Institute, P/12 CIT Scheme, VII-M, Kankurgachi, Calcutta 700 054, India

**Central Tasar Research and Training Institute, Ranchi 835 303, India

The presence and role of the biogenic amine, octopamine has been demonstrated in Antheraea mylitta during pupal diapause. For elucidation of estrogeninduced responsiveness of insects, three consecutive injections of estradiol-17- β (E₂) at doses of 1, 5, 10 and 50 µg/pupa on days 130, 135, and 140 of pupal age were injected to both male and female pupae of A. mylitta during diapause. E₂ treatment caused a significant enhancement in plasma octopamine concentration of haemolymph on day 150 (except in males with 50 µg dose) and reduction on day 165 in both the male and female A. mylitta. On the contrary, plasma protein concentration was found to be higher only on day 150 when treated with E₂ between 1 and 50 µg doses. Octopamine titer in haemolymph plasma always remained higher in male than its female counterpart while in case of plasma protein titer it was found to be just reverse in control animals. E2, @1-50 µg/pupa caused a significant reduction in pupal duration inducing early moth eclosion. Egg production increased at lower doses and decreased at higher doses of this hormone. E₂ at the dose by 0.5 µg/pupa remained ineffective in all the cases except in elevating the female plasma protein titer and egg production. Hence, diapausing pupae of tropical tasar silkworm, A. mylitta is physiologically responsive to vertebrate estrogen, E₂.

PUPAL diapause is a common phenomenon in wild tropical tasar silkworm Antheraea mylitta Drury, which continues up to 200-210 days depending on the ambient environmental conditions. In Lepidoptera, pupal diapause occurs because the pupal brain stops secreting the peptidic prothoracicotropic hormone (PTTH) in response to diapause programming signals (mainly short day photoperiod) received in the larval stage. According to Denlinger², since PTTH is necessary for maintaining

¹For correspondence.