Biochemical effects of a newly synthesized cyclohexyloxy compound JHA on the lipids of fourth-instar larvae of *Culex pipiens* quinquefasciatus Say

Synthetic juvenile hormone analogues (JHAs) behave like endogenous juvenile hormone (JH) of insects and thus are insect-growth regulators. The newly synthesized compound 1-(3'mcthyl-6'isopropyl-cyclohexyl-oxy-3, 7-dimethyl-2 (E)),6-octadiene has been found to be a JHA'. Shortly before ecdysis, wax (consisting of long chain of hydrocarbons and the esters of fatty acids and alcohols) is secreted on the surface of the new cuticle and the layer adjacent to the cuticulin – the oriented monolayer². The importance of lipids in insects has been reviewed³. Obvious success of insects on the planet has been their ability to utilize lipids efficiently as substrate for reproduction, embryogenesis, metamorphosis and flight⁴. This study was undertaken to know the effect of synthetic test JHA on the lipids of treated fourth-instar larvae

of Culex along with control. Thin layer chromatography (TLC) and biochemical studies on lipids revealed that the JHA causes decrease in different lipids. These changes seem to be responsible for formation of intermoults and mortality of developing stages.

Fourth-instar larvae were selected because this is the transition stage between larvae and pupae of mosquitoes. It has a critical period when the ecdysiotropin secreted by neurosecretory cells is very low and this period is the most suitable one for synthetic JHA treatment for insect control⁵. Lipids are important constituents of the fat body cells in insects^{2,3}. These cells contribute lipids to the new cuticle during metamorphosis through oenocytes². Keeping in view the importance of lipids in metamorphosis in mosquitoes, the present study was undertaken to know

the mode of action of test compound in metamorphosis.

Fourth-instar larvae of pure colony of Culex pipiens quinquefasciatus were reared in 1 ppm, 2 ppm and 3 ppm aquatic solutions of JHA having molecular formula C₂₀H₃₆O along with controls. Total lipids, phospholipids and cholesterol were estimated following the method of Folch et al.⁶, and lipid fractions were separated using the TLC technique. Developed spots were estimated following the methods of Mangold⁷, Zlatkis et al.⁸, Lowry and Tinsley⁹, Sidney and Bernard¹⁰.

Biochemically estimated values of lipids (total lipids, phospholipids and cholesterol) are given in Table 1. A percentage decrease in total lipids, phospholipids and cholesterol was observed after 1 ppm, 2 ppm, 3 ppm doses compared to their con-

Table 1. Quantitative changes in lipids in the fourth-instar larvae of Culex treated with JHA

S. no.	Dose	Total lipids	Phospholipids	Cholesterol 3.231 ± 0.727	
[.	N (0 ppin)	51.68 ± 0.280	13.698 ± 1,166		
2.	C1 (1 ppm)	51.12 ± 0.740	12.016 ± 0.5999	2.764 ± 1.871	
3.	T1 (1 ppm)	50.88 ± 3.458***	10.376 ± 1.028	2.528 ± 1.650	
% Change		- 0.47%	- 13.65%	- 8.54%	
4.	C2 (2 ppm)	50.09 ± 1.089	11.529 ± 0.734	2.530 ± 0.148	
5.	T2 (2 ppm)	49.44 ± 3.315	08.710 ± 1.205***	$2.255 \pm 0.148*$	
% Change	• •	- 12.98%	- 24.45%	- 10.87%	
6.	C3 (3 ppm)	49.95 ± 2.645	11.346 ± 0.754	2.203 ± 1.102	
7.	T3 (3 ppm)	38.96 ± 0.841***	$04.501 \pm 0.491***$	1.194 ± 1.136**	
% Change	- • •	- 22%	- 60.33%	-45.0%*	

^{*}P < 0.05. All values (mg/g) are expressed as mean \pm SD of six replicates.

N = normal water, C = Acetone in water, T = JHA in acetonic water.

Table 2. Quantitative changes in TLC fractions of phospholipids in fourth-instar larvae with JHA

	Phospholipids								
	Tri- glyccrides	Phosphatidyl serinc Phosphatidyl inositol	Lyso- phosphatidyl choline	Sphingo- myelin	Phosphatidyl choline	Phosphatidyl ethanolamine	Free fatty acids	Cholesterol	Cholesterol ester
C	2.235	3.3	1.9	2.123	2.3	3.3	3.5	3.23	1.117
1 ppm	1.647	1.9	1.4	1.169	1.82	3.0	3.26	2.456	1.03
2 ppm	1.294	1.5	1.1	1.63	0.981	2.2	2 82	2.212	0.867
3 ppm	0.941	1.1	0.983	L199	0.732	2.0	2.72	1.166	0.829

All values are in mg/g.

^{**}P < 0.01

^{***}P < 0.001.

trols. Increased dose caused increased decrease in lipids. Quantitative estimation of TLC spots of individual lipid classes is given in Table 2. These values showed a decrease in phospholipids (phosphatidyl serine, phosphatidyl inositol, lysophosphatidyl choline, sphingomyelin, phosphatidyl, ethanolamine), cholesterol and its ester, triglycerides and free fatty acids after 1 ppm, 2 ppm and 3 ppm compared to their control.

The fat body is the important site for the storage and biosynthesis of lipids^{2,3} and contributes lipids to the different layers of new cuticle of insects during metamorphosis through oenocytes². It seems that JHA interferes in biosynthesis of lipids by the fat body and thus the formation of new cuticle is impaired as reported by Mittal et al.¹. Decrease of cholesterol ester and cholesterol seems to be responsible for decreased ecdysone biosynthesis which causes incomplete moulting as observed after treatment with

test compound². As the phosphatides are present in different tissues in insects, free fatty acids are required for the growth of larvae, and triglycerides are major reserve food lipids in developing stages of insects^{2,3}; their decrease seems to be responsible for mortality of developing stages, incomplete metamorphosis and formation of intermoults observed after treatment¹.

- 1. Mittal, P. K., Kad, G. L. and Navpreet, J. Environ. Biol., 1997, communicated.
- 2. Chapman, R. F., The Insects: Structure and Functions, Hodder and Stoughton Ltd, Bristol, England, 1971.
- 3. Beenakkers, A. M. Th., Van Der Horst, D. J. and Van Marrevijk, W. J. A., *Prog. Lipid Res.*, 1985, 24, 19-67.
- 4. Gilbert, L. I. and Chino, H. J., Lipid Res., 1974, 15, 439.
- 5. Slama, K., Annu. Rev. Biochem., 1971, 40, 1079-1102.
- Folch, J., Lees, M. and Sloane-Stanely,
 G. H., J. Biol. Chem., 1957, 226, 497-509.

- 7. Mangold, J. Am. Oil C 38, 708.
- 8. Zlatkis, A., Zak, B. and Boyle, A. J., J. Lab. Chem. Med., 1953, 41, 486.
- 9. Lowry, R. R. and Tinsley, J. J., J. Am. Oil Chem. Soc., 1976, 53, 470.
- 10. Sidney, P. G. and Bernard, L., Clin. Chem., 1973, 19, 1077.

ACKNOWLEDGEMENTS. We thank the Chairman, Department of Zoology, Panjab University. Chandigarh for providing laboratory facilities and Dr G. L. Kad of the Department of Chemistry, Panjab University, Chandigarh for the gift of test compound.

P. K. MITTAL NAVPREET KAUR

Department of Zoology,
Panjab University,
Chandigarh 160014, India

FORM IV

Particulars of Current Science—as per Form IV under the Rule 8 of the Registration of Newspapers (Central) 1956.

- 1. Place of Publication: Bangalore
- Publishers' Name, Nationality and Address:
 P. Balaram and S. Ramaseshan,
 Indian,
 Current Science Association, Bangalore 560 080
- 2. Periodicity of Publication: Fortnightly
- 5. Editors' Name, Nationality and Address: P. Balaram and S. Ramaseshan, Indian,

Current Science Association, Bangalore 560 080

- Printers' Name and Address:
 P. Balaram and S. Ramaseshan
 Current Science Association, Bangalore 560 080
- 6. Name and Address of the owner: Current Science Association, Bangalore 560 080

We, P. Balaram and S. Ramaseshan, hereby declare that the particulars given above are true to the best of our knowledge.

Bangalore
1 March 1998

(Sd/-)
P. Balaram and S. Ramaseshan
Publishers, *Current Science*