A NOTE ON THE OCCURRENCE OF WHITE PLASTER DISEASE OF CULTIVATED MUSHROOMS (AGARICU'S BISPORUS) CAUSED BY SCOPULARIOPSIS FIMICOLA—A NEW FUNGUS FROM INDIA

S. C. BHARDWAJ*, P. K. SETH and D. S. GULERIA

Department of Mycology and Plant Pathology, Dr Y. S. Parmar University of Horticulture and Forestry, Solan 173 213, India.

*Present address: Scientist S-1, IARI Regional Station, Flowerdale, Shimla 171 002, India.

THE fungus Scopulariopsis fimicola (Cost. et. Matr.) Vuillemin is known to occur on beds of Agaricus bisporus (Lange) Imbach in other countries¹. The fungus is not reported from India². In the present investigation, it is recorded as a pathogen of A. bisporus. Some information is given on symptoms, morphology and control of the pathogen.

The pathogen occurred as a white mycelial mat (plaster) on the surface of compost and casing soil. The whitish growth changed to light pink after a week of formation of the spots. Sometimes plastered spots of more than 50 cm were observed. The presence of the pathogen inhibited the spawn run depending upon the plastered area. In some cases normal mycelium of A. bisporus developed; however, no fruiting bodies were produced. When plastered growth was found on the surface of casing soil, some trays had to be discarded as these failed to produce any crop.

The fungus is slow-growing in culture; 90 mm petri plates of potato dextrose agar (pH 6.5) were filled in 2 weeks at 25±1°C. Colonies initially membranous, folded, floccose, soon turning vinaceous buff at the centre with thin white margins; sporulation heavy and powdery pustules scattered all over the surface; the reverse surface of the colony dark avellaneous. Hyphae hyaline, $1.5-3.5 \mu m$ wide, conidiophores of variable length, annellophores borne laterally on aerial hyphae $10-28 \times 2-4 \mu m$, tapering to 1-2.5 μ m at the apex, sometimes look like pointed, bear spores. Annellospores ovate, occasionally subglobose, round or showing truncation, buff to avellaneous in mass, occur in chains or clusters, measure $4.8-9 \times 4-8 \mu m$. The fungus was identified as Scopulariopsis simicola (Cost. et. Matr.) Vuillemin (Bull. Soc. Mycol. Fr. 1911, 27, 143) as a new record in India and the culture has been deposited in the Indian Type Culture Collection under accession number 3548 (figure 1a, b).

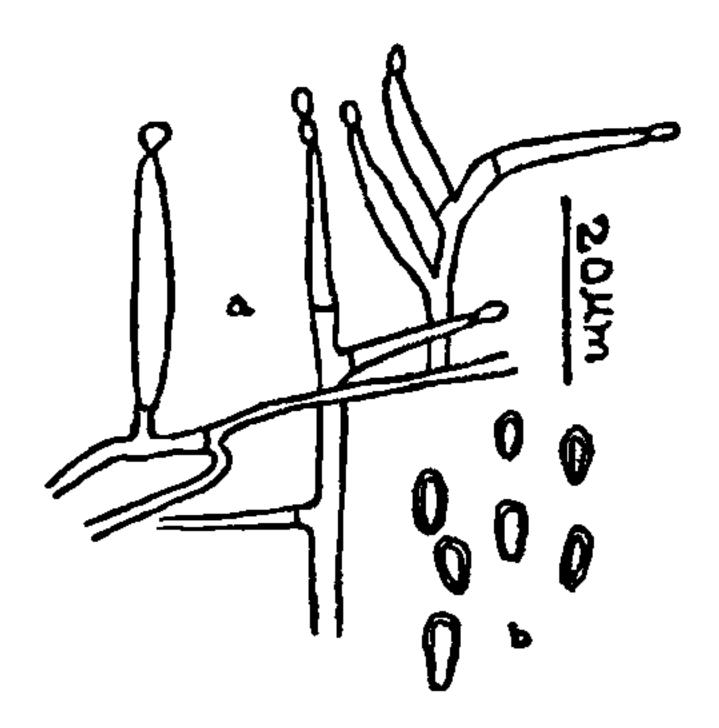


Figure 1a, b. Scopulariopsis fimicola (Cost. et. Matr.) Vuillemin. a. Developing conidia on annellophores, and b. Conidia.

Preliminary observations revealed that disease was more severe in under- or over-composted compost which retained the smell of ammonia and where pH was more than 7.5 at the time of spawning. Further studies on crops grown in small trays (30×25×15 cm, containing 6 kg of compost) according to the method of Munjal and Seth³ confirmed it. An average yield of 500 g was obtained in diseased condition and 800 g in healthy condition at pH 7. The corresponding yields for pH values 7.5, 8 and 6 were 470/815, 110/845 and 268/413 respectively (5 replicates and 3 crops).

Maintenance of a temperature of 60°C throughout the compost during peak heat phase of pasteurization eliminated the pathogen completely, as indicated by absence of growth in repeated platings. Good hygiene combined with careful removal of stray patches with subsequent treatment with formalin (4%) further insured the crops against the disease.

The authors are grateful to Dr J. N. Kapoor, Sr Mycologist, Division of Mycology and Plant Pathology, IARI, New Delhi, India, for his kind help in confirming the identity of the fungus.

13 August 1986; Revised 17 October 1988

- 1. Fletcher, J. T. and Atkinson, K., Mushrooms—A guide to the recognition and control of diseases, weed moulds, competitors and pests. Ministry of Food and Agric. div. and Advisory Service, Lawnswood, Leeds, England, 1977, p 12.
- 2. Bilgrami, K. S., Jamaludin and Rizwi, M. A.,

Fungi of India, Part 1, Today and Tomorrow's Printers and Publishers, New Delhi, 1979, p 467.

3. Munjal, R. L. and Seth, P. K., Mushroom cultivation in India, Special priced bulletin of Indian Mushroom Growers' Association 1980, p. 40.

POLY-β-HYDROXYBUTYRIC ACID CONTENT OF RHIZOBIUM STRAINS IN RELATION TO GROWTH

A. K. PODDER, S. I. KHAN and A. A. CHOWDHURY

Department of Microbiology, University of Dhaka, Dhaka, Bangladesh.

POLY-\(\beta\)-HYDROXYBUTYRIC acid (PBH) is a storage product in pure cultures of rhizobia¹ and rhizobial cells in root nodules and accumulation of PBH in root nodules is related to photosynthetic products made available by nodulated legumes². During stress, PBH serves as a reservoir of energy for biological nitrogen fixation. However, it is not known whether PBH content of rhizobial strains is related to the ability of strains to grow fast or slow, a criterion being increasingly recognized in speciation and effectiveness of rhizobia. With this objective in mind, root nodules of seven economically important leguminous crops (Lens culinaris, Lathyrus sativus, Pisum sativum, Sesbania aculeata, Glycine max, Vigna radiata and Vigna mungo) were collected from different places of Bangladesh and the native rhizobia isolated by following well-known procedures³. The generation time of several strains from nodules of each legume was calculated on the basis

Table 1 Generation time and PBH content of rhizobia from root nodules of some crop legumes from Bangladesh

Host	Strain Nos.	Range of generation time (min)	Range of PBH content (g/100 g wet weight of cells)
L. culinaris	DS11-DS16	180208	5.5-8.5
L. sativus	DS17-DS21	170-193	5.8 - 10.8
P. sativum	DS22-DS26	210-243	5.6-11.3
S. aculeata	DS27 DS33	88-94	5.3 13.5
G. max	DS39- DS45	313-340	1.1-3.7
V. radiata	DS46-DS51	382 401	6.0 11.2
V. mungo	DS52 DS57	451-478	4.8 9.2

of growth curves on yeast extract mannitol broth based on turbidimetry³. The PBH content of different strains was determined gravimetrically following the method of Low and Slepecky⁴ and expressed on percentage basis. The results show that both fast and slow growing rhizobial strains had PBH in varying amounts, the highest range in strains from fast-growing strains of S. aculeata and the lowest range in slow-growing strains of G. max; an intermediate situation, however, was noticed in another very slow-growing group of strains of V. mungo which had medium range of PBH (table 1). These results indicate that growth rate of rhizobia (fast or slow) has a bearing on their PBH content.

1 September 1988

- 1. Schlegel, H. G., Flora, 1962, 152, 236.
- Wong, P. P. and Evans, H. J. Plant Physiol., 1971, 47, 750.
- 3. Vincent, J. M., A manual for the practical study of the root nodule bacteria, Blackwell Scientific Publications, Oxford, 1970, UK.
- 4. Low, J. M. and Slepecky, R. A., J. Bacteriol., 1961, 82, 33.

GENETIC DIVERSITY AMONG PLANTAGOS VI. SATELLITE ASSOCIATIONS IN *PLANTAGO* OVATA FORSK.

P. K. SHARMA*, A. LANGER and A. K. KOUL Department of Biosciences, University of Jammu, Jammu 180 001, India.

*Present address: Regional Research Laboratory, Sanat Nagar 190 005, India.

THE phenomenon of 'Satellite association' (i.e. the tendency of the Sat. chromosomes to so orient that their satellites appear fused) is a common feature of mitotic metaphase spreads of human beings. A linear relationship exists between the transcriptional activity of nucleolar chromosomes and the frequency of satellite associations. This holds true for many animal taxa as well¹⁻⁴. Among plants, the phenomenon is comparatively infrequent. Recently, Sato et al⁵ recorded satellite associations in Nothoscordum fragrams, a bulbous angiosperm, and published it as