- Mycol. Soc., 1986, 87, 401,
- 9. Sujan Singh, Khan, S. N. and Misra, B. M., Indian For., 1983, 109, 185.
- Mohanan, C. and Sharma, J. K., Curr. Sci., 1987, 14, 732.
- 11. Sutton. B. C., The Coclomycetes. CMI, Kew, England, 1980, p. 696.
- 12. Anderson, D. M. W., Nitrogen Fixing Tree Res. Rep., 1984, 2, 24.
- 13. Kobayashi, T., Trans. Mycol. Soc. Jpn., 1978, 19, 373.
- 14. Sharma, J. K., Mohanan, C. and Maria Florence, E. J., Disease survey in nurseries and plantations of forest tree species grown in Kerala, KFRI Res. Report No. 36, 1986, p. 275.

AN INDUCED SHELL MUTANT IN GROUNDNUT (ARACHIS HYPOGAEA L.)

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Rest disease caused by Puccinia arachidis Speg., leaf miner (Aproaerema modicella Dev.) and drought are the major biotic and abiotic stresses limiting groundnut productivity in India. NCAC 17090, a Valencia Bunch genotype has been reported to be resistant to rust and drought and tolerant to leaf miner and is frequently used as a donor for infusing resistance to these stresses since it has

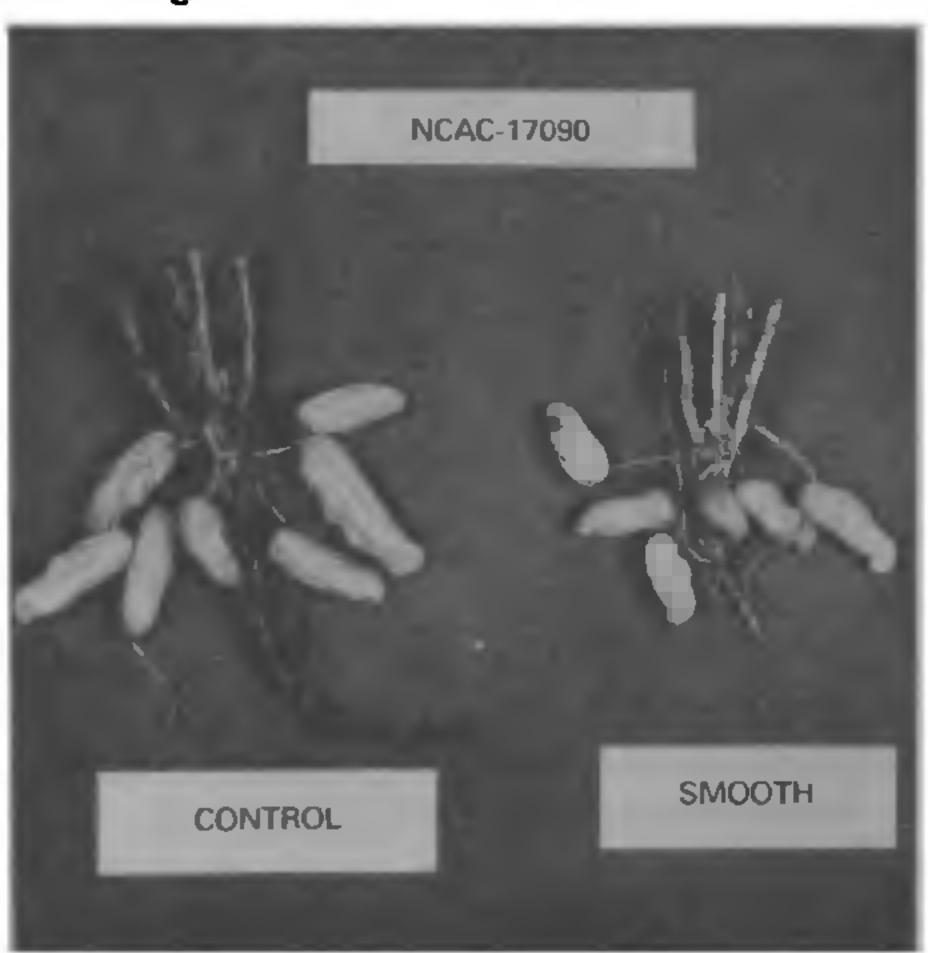


Figure 1.

Table 1 Shelling (%) and reaction to brotic stresses of the mutants

		Reaction to rust/leaf miner(1-9 scale)		
Mutants/control	Shelling %	Rust	Leaf miner	
Smooth shelled	72.0	2	4	
	72.5	3	4	
	71.5	2	4	
Control	68.0	2	4	

an acceptable testa colour (rose). However, its shell is thick, highly reticulated which makes it a poor sheller and unattractive. Hence, with a view to improving the shell characteristic of NCAC 17090 a study was taken up.

The kernels of NCAC 17090 were treated with 25, 50 and 75 krad of gamma rays. For each dose 60 seeds were treated and sown along with untreated control in three replications during summer 1987. The M₂ generation was raised on M₁ plant basis. For each M₁, 20 seeds were sown in a progeny line and the remaining seeds, if any, were bulked dosewise and raised as bulk in *Kharif* 1987.

Among the progenies, one tamily (NCAC 17090–25 KR-14) in 25 krad treatment segregated for shell characters, i.e. 15 with normal types and three variants. The variants in the progeny exhibited smooth shell and small pods (figure 1) while retaining other features of untreated control. These three mutants were advanced to M₃ generation and they were found to breed true. In order to study the genetics of these mutants studies are in progress to effect crosses.

The breeding behaviour of the mutants in respect of shell character and reaction to rust³ and leaf miner⁴ was recorded according to the methods reported earlier. The shelling percentage of the plant progenies was higher when compared to that of the parents and they were equally resistant to rust and leaf miner as their parents (table 1).

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- 1. Subrahmaniyam, P., Gibbons, R. W. and Nigam, S. N., Peanus Sci., 1980, 7, 10.
- Annual Report, ICRISAT, Patancheru, 1985,
 p. 215.
- 3. Subrahmanyam, P., McDonald, D., Gibbons, R. W., Nigam, S. N. and Nevill, D. J., Peanut Sci., 1982, 9, 6.

4. Anon, Proceedings of XXVIII annual kharif oilseeds workshop, Andhra Pradesh Agricultural University, Rajendranagar, 28 April to 1 May, 1986, p. 2.

ON THE OCCURRENCE OF STYLET-BEARING NEMATODES ASSOCIATED WITH MANGROVES OF GANGETIC ESTUARY, WEST BENGAL, INDIA

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Mangroves are very specialized forest ecosystem found at the land-see interface of the tropical and subtropical regions of the world bordering the sheltered sea coast and estuaries. The Gangetic estuary, located at the confluence of the river Ganges and Bay of Bengal, is unique in harbouring world's most luxurient mangrove swamp, the deltaic Sundarbans. This very vibrating ecosystem supports numerous benthic life forms in the form of micro, meio and macro benthos. Nematodes are found to be the predominant meiofaunal component amongst the litter soil fauna in this complex environment.

Although a good number of plant parasitic nematodes of the order Tylenchide and Dorylaimide have been reported from brackish water¹, sand dunes² and mangrove swamps³ from American, Australian and European subcontinents respectively, records of stylet-bearing plant parasitic nematodes from Sundarbans mangrove swamps of the Indian territory are limited with the exception of a solitary report4. The present paper reports on the occurrence of some stylet-bearing nematodes encountered during the study of nematode fauna of mangrove swamp and adjoining sandy beaches of the Gangetic estuary associated with roots of various mangrove vegetations. The current monitoring may help to elucidate on the high salt tolerance of the nematode specimens of this dynamic environment.

Soil samples were collected from estuarine habitats at the following locations:

- 1. Habitat a: Intertidal mudflat at Harinbari of Sagar Island (21°37 to 21°52 N and 88°03 to 88°11 E), habitat sheltered, important mangrove vegetations are Acanthus ilicifolius L, Excoecaria agallochia L and Bruguiera gymnorhyza (L) Lam, salinity 28% S.
- 2. Habitat b: Intertidal sandflat at Gangasagar of Sagar Island, habitat exposed, dominant mang-

Table 1	Stylet-bearing	nematodes	associated	with	mangrove	flora of	Gangetic	estuary,	West	Bengal
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Orders	Families	Species	Habitats
Dorylaimida	Leptonchidae	Proleptonchus paucipapillatus (Meyl 1965; Goseco et al 1974)	C
·		Doryllum aestuarii (Timm, 1967)	b
		Dorylaimoides sp.	a
	Dorylaimidae	Laimydorus parabastiani (Paetzold 1958; Siddiqui 1969)	b
		Timmus sp.	3
	Aporcilainiedae	Thonus sp.	а
	Nygolaimidae	Nygolaimus sp.	b
		Nygolaimoides sp.	a
	Longidoridae	Paralongidorus sp.	а
	Belondridae	Paraoxydirus sp.	a
	Nygellidae	Nygellus sp.	b
Tylenchida	Anguinidae	Indoditylenchus sundarbanensis (Sinha et al 1955)	c
	Tylenchidae	Tylenchus sp.	C
	Criconematidae	Hemicriconemoides sundarbanensis (Ganguly & Khan, 1983)	a
		Nothocriconema sp. nov.	a
	Hopelaimidae	Helichotylenchus sp.	ь
	Pratylenchidae	Hirschmannialla gracilis (De Man, 1880; Lue & Goodey, 1963)	þ