Table 1 Effect of Methoprene (ZR-515) on the prepupae of Phthorimaea operculella Zeller*

	Percentage of			
Conc. of Methoprene (%)	Prepupae that became normal pupae	Prepupae malformed or dead	Adult emergence	
0.25	70	30	30	
0.50	5 5	45	30	
0.75	30	70	20	
1.00	12	88	Nil	
1.50	05	95	Nil	
Control	95	05	70	
CD(P=0.01)	1.92	1.20	1.03	

^{*}Averages of results from four replicate experiments.

emergence in the control group was significantly higher than in the JHA-treated groups. These findings are supported by those of Outram⁵ who reported that adult emergence in *Prodenia litura* was affected at higher doses of a synthetic juvenile hormone (Roeller compound). Outram⁵ also reported that there was a low percentage of emergence of deformed adults when spruce budworm pupae were treated with high doses of a synthetic JHA. In the present study also, it was found that the adults that emerged from the Methoprene (ZR-515) treated pupae were malformed. Such malformations were also reported earlier⁴ in tobacco cutworm pupae treated with JHA (Aliozar).

The authors are thankful to Ms H. N. Rama for supplying the potato tuber moth culture. The Juvenile hormone analogue, Methoprene (ZR-515), received from M/s Zoecon Corporation, Palo Alto, California, USA, is gratefully acknowledged.

Table 2 Effect of Methoprene (ZR-515) on the pupae of Phthormaea operculella Zeller*

Coma of	Percentage of		
Conc. of Methoprene (%)	Normal adults	Adults with mal- formed characters	
0.25	30.0	Nil	
0.50	23.3	08 2	
0.75	15.0	12.5	
1.00	15.0	18.3	
1.50	05.0	32.0	
Control	70.0	Nil	
CD(P = 0.01)	2.04	2.30	

^{*}Averages of results from four replicate experiments.

22 April 1988

- 1. Lall, B. S., Indian J. Entomol., 1964, 184, (Silver Jubilee Number).
- 2. Devaraj Urs, K. C. and Ramamurthy, R., Curr. Sci., 1976, 45, 600.
- 3. Ambika, B. and Abraham, C. C., Agric. Res. J., Kerala, 1982, 20, 60.
- 4. Devaraj Urs, K. C. and Byakod, S. S., Curr. Sci., 1976, 45, 599.
- 5. Outram, I., J. Econ. Entomol., 1973, 66, 1033.

TOTAL DEVELOPMENTAL ARREST OF FOURTH INSTAR LARVAE OF CULEX QUINQUEFASCIATUS TREATED WITH PENFLURON

S. C. SAXENA and R. K. KAUSHIK Department of Zoology, University of Rajasthan, Jaipur 302 004, India.

Although several workers¹⁻³ have observed the adverse effects leading to mortality of Penfluron, this is the first report pertaining to complete developmental arrest of fourth instar larvae of Culex quinquefasciatus treated with Penfluron. The duration of the 4th instar is abnormally prolonged and the larvae fail to pupate and finally die as 4th instar larvae.

A laboratory colony of *C. quinquefasciatus* was maintained under controlled conditions (temperature 28°C and humidity 70–80%). Penfluron was dissolved in acetone (1% w/v) to obtain a standard solution. Other concentrations (0.5, 1 and 5 ppm) were prepared by diluting the standard solution with distilled water. Tween-40 was used as an emulsifier. In the control experiment, acetone and emulsifier alone were used. Larvae in the 1st day of the 4th instar were used.

It is evident from the data (table 1) that Penfluron significantly prolongs the duration of the 4th instar and completely suppresses pupation. The maximum survival recorded for larvae treated with 0.5 ppm Penfluron was 35 days. For larvae treated with 1 and 5 ppm of Penfluron, the maximum survival recorded was 26 and 7 days respectively. In the case of treatments with 0.5 and 1 ppm of the insecticide, about 90% of the larvae survived for 10 days without pupating. None of the larvae pupated. In the control group all the larvae pupated on the 4th day of the 4th instar and on the 4th day after pupation adults emerged from all the pupae.

Table 1 Effect of Penfluron on 4th mytar larvae of Culex quinquefasciatus

Conc. (ppm)	Repli- cates	No. of Jarvae treated	S				ays of	4th m	tar and	no, of su	rviving	arvae*				
6.5		77	1-2	3-8 9	= +	11-12	13-16	17	18-20	$\frac{12}{18} \frac{13-16}{16} \frac{17}{16} \frac{18-20}{14} \frac{21-23}{13} \frac{24}{10} \frac{25-26}{8} \frac{27}{10}$	24	25-26	27-28	<u>29</u>	30-32	2 33-35
	c 1	61	1-2	3-9 10	11 91	12-13	14-18	19-20	21-23	24-27	28-29	30-32	6 0			
	وسمه	23	1-2	$\frac{3-8}{22} = \frac{9}{2}$	-10 161	11-13	14-16	17-19	20-24	25-28	29-31	32	33	34-35	98 0	
1.0	-	15	1-3	1-9 10 13	11 91	$\frac{12-13}{9}$	7 7	15-16	17-18	19-20	$\frac{21-22}{2}$	$\frac{23-25}{1}$	<u>ද</u> ၁			
	7	2	1-3	17 -10	-13	17 17	15-16	17-18	19-21	$\frac{22-23}{2}$	24					
	į. E	15	1-3	13)-12 12	13-15	16	17-19	$\frac{20-22}{4}$	$\frac{23-25}{3}$	97 0					
5.0		20	- 15	C1 ±	3	4 IQ	N 14	1 1	7							
	7	20	16	2 12	w 11-	ব।ব	w 10	ס ורו	7 0							
	m	20	12	C1 11-	win	4 161	215	<u>~ 10</u>								
			-1	arval .	trh ins	tar)-1-4	upa							
Control		25	1.25	2 2	3 24	4 24	5 24	6 24	7 24	8 <u>24</u>						
					¥	II pupated			~ ¶	All adults emerged						

(Contd. . . . next page)

$\overline{}$
-
•
•
contd.

~
9
Ç.
-
•
Ø
(Table
-
, U

rvae*			
,			
f 4th instar and no. of surviving la	25	All adults emerged	8 23 All adults emerged
star and	<u>~ 2</u>	•	7 131
4th in	25		23
Days of	55		2 2
Da	4 25	Ail pupated	4 24 All pupated
	3		24
	25		2 5
	1 25		1 25
No. of larvae treated	25		25
Repli- cates	7		~
Conc. (ppm)			

*Figures in upper row

The results show that Penfluron disrupts the growth and development of 4th instar larvae of C. quinquefasciatus. The significant observations are prolongation of larval period and complete suppression of pupation. All the concentrations tested blocked development; with higher concentrations the effect was quicker.

Prolongation of larval period and suppression of pupation may be due to imbalance caused by the insecticide in growth-stimulation and growth-inhibition hormone levels as suggested by Novak⁴. The juvenilizing activity of chitin inhibitor compounds against mosquito larvae has been reported earlier⁵⁻⁸. Gujar and Mehrotra⁹ recorded the juvenilizing effect of plant extracts on the last larval instar of Spodoptera litura and related the juvenilizing action to disruption in the insect's endocrine system. It is quite possible that complete arrest of development induced by Penfluron in C. quinquefasciatus is a juvenilizing effect of a disrupted neuroendocrine system. Whether such an effect is insect-specific is to be investigated.

The authors are grateful to Dr A. B. Borkovec, Agricultural Research Centre, Beltsville, for the gift of Penfluron.

4 April 1987; Revised 14 March 1988

- 1. Saxena, S. C. and Mathur, G., J. Adv. Zool., 1980b, 2, 1.
- 2. Saxena, S. C. and Kumar, V., *Entomology*, 1982, 7, 141.
- 3. Saxena, S. C. and Kaushik, R., Curr. Sci., 1986, 18, 902.
- 4. Novak, V. J. A., In: *Insect hormones*, 1966, p. 18.
- 5. Jakob, W. L. and Schoof, H. F., Mosquito News, 1971, 31, 737.
- 6. Jakob, W. L. and Schoof, H. F., Mosquito News, 1972, 32, 6.
- 7. Jakob, W. L., Mosquito News, 1972, 32, 592.
- 8. Jakob, W. L., J. Med. Entomol., 1973, 10, 452.
- 9. Gujar, G. T. and Mehrotra, K. N., Indian J. Exp. Biol., 1983, 21, 292.

EFFECT OF VARIOUS INSECTICIDES ON HONEY BEE, APIS FLOREA FABRICIUS IN 'BER' (ZIZYPHUS MAURITIANA LAMK)

B. H. PATEL, V. R. UPADHYAY, C. M. MURALIDHARAN and G. S. JUDAL Department of Entomology, College of Agriculture, Gujarat Agricultural University, Sardar Krushinagar 385 506, India.

'BER' (Zizyphus mauritiana Lamk) is a cross-pollinated fruit crop, where pollinators play a vital role in fruit setting. It is reported that 85% of the cross pollinated flowers depend on insect pollinators¹. In India, the activities of honey bees on different crops were reported earlier^{2,3}. Ber is an important fruit crop in the semi-arid zones of North Gujarat, where Apis florea is an important pollinator. However, scanty information is available on the adverse effects of various insecticides (to A. florea) which are commonly used in 'ber' for insect pest management. The present study was carried out to determine the effect of insecticidal spraying on the population of A. florea.

The popular ber variety 'Umran' was used for this study and the experiment was set in a split-plot design with nine insecticidal treatments and one untreated control. The details of insectides used are given in table 1. Spraying was done with a knapsack sprayer using 5 1 mixture per tree. The first spraying was immediately after flowering and subsequent sprayings were done at a three week interval. Initial observations on the honey bees were recorded 24 h before each spraying and there after observations were taken at 24, 48, 72 h and one week after spraying. For recording the honey bee population, four branches (50 cm each) each from south, north, east and west direction were tagged. The observations on honey bee population were recorded by four persons at a time standing in four different directions, observing the tagged branches at 5 min interval. The results subjected to statistical analysis are presented in table 1.

The data on the population of the noney bee recorded before and after spraying (table 1) revealed a significant difference in the honey bee population due to insecticide, spray, period, insecticide × period and spray period. The control recorded the highest bee population (3.17/0.5 m branch) which was statistically at par with 0.07% endosulfan (3.05/0.5 m branch), 0.03% thiometon (2.82/0.5 m branch) and 0.03% demeton-o-methyl (2.81/0.5 m branch) whereas the rest of the insecticidal treatment remained at par and showed lower