



Figures 1–3. 1. Chromosomal organization of dividing nucleus. Note the nuclear membrane is still intact in *Terminalia* (nm, Nuclear membrane) ($\times 1400$); 2 and 3. Multinucleate fusiform initials of *Mangifera* and *Albizzia* respectively. Note the globular nuclei in the former and spindle-shaped nuclei in the latter (2. $\times 900$; 3. $\times 350$).

initials in *T. grandis* is confirmed and its presence in *Albizzia*, *Dalbergia*, *Mangifera*, *Morinda* and *Terminalia* is recorded for the first time.

The multinucleate condition is perpetuated even in the differentiation of vessel elements of *D. sissoo*⁹ while Patel³ observed the uninucleate xylem and phloem of *Solanum melongena*. This needs further studies. Occurrence of multinucleate condition in the ray cell initials is recorded for the first time here in *Albizzia*, *Dalbergia*, *Tectona*, *Terminalia*, *Mangifera* and *Morinda*.

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CHARACTERIZATION OF SHEATH FROM PHOTOHETEROTROPHICALLY GROWING STRAIN OF *CALOTHRIX MARCHICA* LEMM.

S. P. ADHIKARY

Department of Botany, Utkal University,
Bhubaneswar 751 004, India.

THE filamentous cyanobacterium *Calothrix marchica* grew photo- and chemoheterotrophically in the presence of sucrose and fructose in the medium. A distinct sheath layer was developed around the trichome of photoheterotrophically grown strain of *C. marchica*¹. It was thought that considerable proportions of the sugars were assimilated by *C. marchica* in certain specific biosynthetic process, (for example, in the synthesis of sheath layer around the trichome). The present work was undertaken to isolate and characterize the sheath from the heterotrophically growing strain of *Calothrix*.

Calothrix marchica Lemm. var. *intermedia* Rao strain Sahu 1978/3² was used as the experimental

material. The cyanobacterium was grown photoheterotrophically in the presence of 15 mM sucrose in Allen and Arnon's nitrogen-free medium³, irradiated continuously with white light from fluorescent tubes (2400 lux). The organism was harvested after 20 days of cultivation and washed once with 20 mM Tris-HCl buffer, pH 8 (used throughout sheath isolation). The sheath material was isolated from the cyanobacterium after homogenization with an ultrasonicator (4°C, 30 min). Discontinuous sucrose gradient (10 ml of 60% and 5 ml each 55, 50, 45 and 40%, w/w, sucrose in Tris/HCl buffer) was loaded with the cell homogenate and run in a swingout rotor (160,000 g, 4°C, 4 h). Sheaths were recovered from the 60% sucrose band and further purified by treatment with lysozyme (5 mg in 25 ml ammonium acetate buffer, pH 6.8, 37°C) followed by extraction with sodium dodecyl sulphate (2%, w/w in Tris/HCl buffer, 100°C, 15 min). The yield of the sheath material was 8.2% on a cell dry weight basis.

Hydrolysis of the carbohydrates and polypeptides and the determination of monomers have been described elsewhere⁴. The neutral sugars were separated, identified and quantified by gas liquid chromatography as alditol acetate derivatives (Varian aerograph model 1445-1, ECNSS-M column, 3% on Gas chrom Q, 100-200 mesh). Fatty acids were identified as methyl esters by gas liquid chromatography (EGSS-X column, 15% on Gas chrom P, 100-200 mesh). Uronic acids were quantitatively determined by colorimetric carbazole assay at 530 nm. Amino acids and amino sugars were separated (in 4N HCl, 110°C, 16 h) and quantitatively determined in an automatic amino acid analyser. Organic phosphorus content was determined following the method of Lowry *et al*⁵.

Chemical analysis of the isolated sheath of *C. marchica* showed that the neutral sugars were the major components of the sheath fraction; their content increased from 40.5% (gradient purified) to 84.5% (lysozyme + SDS). All fractions contained predominantly glucose and mannose. Absence of muramic acid, diaminopimelic acid and β -hydroxy fatty acids in the lysozyme + SDS extracted sheath fraction showed that it was free from the cell wall components. Glucosamine, uronic acids, fatty acids and phosphorus were detected only in trace in the purified sheath and their amount was 0.25% (w/w), 0.3% (w/w), 0.4% (w/w) and 0.2% (w/w) respectively (table 1). The sheath fraction from the sucrose gradient contained 10.8% (w/w) protein. The total amino acid content was not solubilized by treatment

Table 1 Chemical analysis of the sheath fractions of photoheterotrophically grown filaments of *Calothrix marchica* Lemm. var. *intermedia* Rao (Values represent per cent dry weight of the sheath material)

Constituent	From sucrose gradient	After lysozyme + SDS treatment
Sugars		
Ribose	3.3	—
3-O-methyl pentose	0.4	—
Mannose	5.4	1.3
Glucose	31.4	83.2
Total sugars	40.5	84.5
Total amino acids	10.8	4.2
Fatty acids		
C 14	0.1	0.04
C 16	0.4	0.2
C 16:1	0.1	0.03
C 18	0.1	0.08
C 18:1	0.1	0.05
β -C 16-OH	0.3	—
Total fatty acids	1.1	0.4
Glucosamine	1.7	0.25
Muramic acid	0.76	—
Diaminopimelic acid	1.14	—
Uronic acids	0.68	0.3
Phosphorus	3.4	0.2

— = absent.

with lysozyme and hot SDS, however, was reduced to 4.2% (w/w). A high proportion of glutamic acid and aspartic acid was observed in the purified sheath fraction (not shown in the table).

Chemical analysis of the isolated sheath fraction of photoheterotrophically grown *Calothrix* filaments showed that it was predominantly composed of sugars (84.5%, w/w). Formation of sugar-rich sheath layers in the heterotrophic culture of the cyanobacterium, which in various other cyanobacteria is also composed mostly of neutral sugars^{4,6,7}, may be the consequence of a specific biosynthetic process which is involved in the utilization of exogenous carbon compounds. By utilizing radioactive exogenous carbon sources, various steps of the biosynthetic process can be further characterized.

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FUSARIUM SPECIES ASSOCIATED WITH GRAIN MOULD AND STALK ROT OF SORGHUM AND THEIR EFFECT ON SEED GERMINATION AND GROWTH OF SEEDLINGS

D. M. MAHALINGA*, K. H. ANAHOSUR and R. K. HEGDE

Department of Plant Pathology, College of Agriculture, Dharwad 580 005, India.

*Present address: Agricultural Research Station, CADA, UKP, Bheemarayangudi 585 287, India.

SORGHUM [*Sorghum bicolor* (Linn.) Moench] is an important food crop. Among several diseases infecting sorghum, grain moulds or head moulds and stalk rot caused by species of *Fusarium* are of great economic importance. In recent years they are serious in all tropical sorghum-producing areas.

Curvularia sp., *F. moniliforme*, *F. oxysporum* and *C. lunata*, *Helminthosporium tetramera*, *F. moniliforme* and *F. semitectum* Berk and Rav. were isolated^{1,2} from mouldy grains of sorghum. The association of *F. roseum*, *F. solani* and *F. oxy-*

sporum with root zones of gramenaceous plants and *F. moniliforme* with stalk rot of sorghum was reported earlier^{3,4}.

In the present investigation an attempt was made to isolate and identify the species of *Fusarium* associated with sorghum grain, peduncle, stalk and root and their effect on seed germination and growth of seedlings. The different species of *Fusarium* were isolated following the blotter technique⁵. The cultures were subjected to reisolation by single spore isolation method and maintained on PDA slants at $27 \pm 1^\circ\text{C}$. The individual species of *Fusarium* were identified based on colony characters, growth rate, pigmentation, and full range of spore forms⁶.

The species of *Fusarium* were identified as *F. moniliforme* shed, *F. semitectum* Berk and Rav. and *F. oxysporum* Schlect. Among these *F. semitectum* was isolated from grains, *F. moniliforme* from grains, stalk and peduncle, and *F. oxysporum* from roots of sorghum.

The pathogenicity of these species of *Fusarium* to sorghum earheads, peduncle and stalk was proved by cross-inoculation studies by using sorghum genotypes CSH-1 and CSH-5. The study indicated that all the three species of *Fusarium* were potentially pathogenic on sorghum genotypes CSH-1 and CSH-5 and produced distinct symptoms on grains, peduncle and stalk. It is clear that species of *Fusarium* involved in sorghum grain mould are also pathogenic to stalk and peduncle and the species of *Fusarium* involved in peduncle rot, stalk rot and root rot are pathogenic to grain.

The effect of each species of *Fusarium* on seed germination in soil and on moist blotter was tested separately using CSH-1 seeds by smearing the sorghum seeds with equal quantity of eight-day-old pure culture of each species of *Fusarium*. The per cent germination ranged from 59.5 to 69.8 and 80.1 to 93.2 in soil and on moist blotter respectively

Table 1 Effect of species of *Fusarium* on seed germination and growth of seedlings of CSH-1 sorghum (all values in %)

Fungi	In soil			On moist blotter in petri plate		
	Germination on 7th day	Pre-emergence death on 7th day	Post-emergence death of existing seedlings on 16th day	Germination on 7th day	Pre-emergence death on 7th day	Post-emergence death of existing seedlings on 16th day
<i>F. moniliforme</i>	61.6	38.4	100.0	80.1	19.9	86.7
<i>F. semitectum</i>	59.5	40.5	83.3	91.5	8.5	73.8
<i>F. oxysporum</i>	69.8	30.2	76.6	93.2	6.8	69.2
Control	95.5	0.5	0.0	99.8	0.2	0.0