characteristic of an aromatic proton adjacent to an oxygen substituent.

On the basis of these observations the product was identified as 7-hydroxy coumarin which was finally confirmed by direct comparison with an authentic sample (m.p., m.m.p., UV, IR, NMR).

In control experiment the compound (I) was added to medium without bacterial inoculum and incubated similarly. After 48 hr of incubation the medium was extracted with chloroform and evaporated as described above when a solid mass was obtained which was found to be the original compound by direct comparison.

Similar experiment with compound (II) furnished compound (III).

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RUTHENIUM(III)-CATALYZED EPOXIDATION OF OLEFINS BY N-METHYL MORPHOLINE N-OXIDE

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ALKYLDROPEROXIDES in combination with complexes of molybdenum, vanadium, tungsten and titanium have been used for the epoxidation of olefins\(^1\). The kinetics, mechanism and factors governing catalytic activity in such epoxidations have been investigated\(^2,3\). In the presence of catalytic amounts of (tetraphenyl-porphinato) manganese(III), py-cyano-N,N-dimethylaniline-N-oxide epoxidizes cyclohexene\(^4\). However, olefins are converted into the corresponding glycols by tertiary amine-N-oxides like trimethylene-N-oxide and N-methylmorpholine-N-oxide (NMO) in the presence of catalytic amounts of OsO\(_4\)\(^5,7\). Vijayasi et al\(^6\) reported that in the presence of Ru(III) as catalyst in DMF, NMO oxidizes secondary alcohols to the corresponding ketones. Studies on the use of RuCl\(_3\)-NMO combination in DMF as solvent to epoxidize olefinic substrates like cyclohexene, styrene and terminal alkenes are reported in this communication.

The kinetic investigations were carried out at 35±0.1°C. Oxygen-free nitrogen was bubbled into the reaction mixture to provide an inert atmosphere. The concentration of NMO remaining at any instant was determined titanometrically\(^9\) as reported earlier\(^8\). In the case of non-aromatic substrates the progress of the reaction, especially at low concentrations of the substrate was followed iodometrically. The reaction mixture was quenched with a known excess of bromine in acetic acid and the unreacted bromine was treated with excess of KI. The iodine liberated was titrated against standard sodium thiosulphate.

The stoichiometry of the reaction is in agreement with the equation Alkene + NMO \(\xrightarrow{\text{catalyst}}\) epoxide + N-methylmorpholine. There was no reaction between substrate and NMO in the absence of a catalyst. All the alkenes investigated so far are found to give rise to the corresponding epoxides. This was confirmed by gas chromatographic analysis using an FFAP column and comparison of the retention times with those of authentic samples. The reactions were carried out under pseudo first order conditions by maintaining [substrate] \(\gg\) [NMO]. The reaction was first order in NMO and catalyst. However, the order with respect to the substrate, as determined from the pseudo first order plots, varied depending on the concentration of substrate. Vijayasi et al\(^10\) established the formation of Ru(V) species by spectral and cyclic voltammetric studies. Accordingly the following mechanism could be proposed for the epoxidation of olefins.

\[
\text{Ru(III)} + \text{NMO} \xrightarrow{k_1} \frac{k_1}{k_{-1}} \text{Ru(V) oxo species,}
\]

\[
\text{Ru(V) oxo species} + S \xrightarrow{k_2} \text{product.}
\]

Assuming the Ru(V) oxo species to be in steady state concentrations one can deduce the following rate expression.

\[
\text{Rate} = -\frac{d[NMO]}{dt} = \frac{k_1 k_2 [\text{Ru(III)}][\text{NMO}][S]}{k_{-1} + k_2[S]},
\]

\[
= \frac{k_1 [\text{Ru(III)}][\text{NMO}][S]}{(k_{-1}/k_2) + [S]}. \tag{1}
\]
The variable orders in the substrate could be accounted for on the basis of relative magnitudes of $k_{-1}/k_2$ and $[S]$. Equation (1) can be rearranged to give (2).

$$\frac{1}{\text{rate}} = \frac{k_{-1}/k_2}{k_1[\text{Ru(III)}][S][\text{NMO}]} + \frac{1}{k_1[\text{Ru(III)}][\text{NMO}]}$$

Equation (2) has been verified by plotting $1/\text{rate}$ against $1/[S]$ using the data obtained for low concentrations of the substrate, at constant concentration of Ru(III) and NMO. At high substrate concentrations, $k_{-1}/k_2 < [S]$ and so the reaction is zero order in the substrate. A comparison (table 1) of the values of $k_1$ obtained from the linear plots with those obtained from the pseudo first order rate constants ($k_{\text{obs}}$) supports the proposed mechanism.

**Table 1 Evaluation of rate constants from double reciprocal plots at low concentrations of substrate**

<table>
<thead>
<tr>
<th>Substrate</th>
<th>$k_1$ in M$^{-1}$ min$^{-1}$</th>
<th>From pseudo first order plots</th>
<th>From double reciprocal plots</th>
<th>$(k_{-1}/k_2)$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cyclohexene</td>
<td>11.42±0.13</td>
<td>11.36</td>
<td>0.109</td>
<td></td>
</tr>
<tr>
<td>1-Octene</td>
<td>8.43±0.34</td>
<td>8.30</td>
<td>0.053</td>
<td></td>
</tr>
</tbody>
</table>

Since the olefinic substrates can form complexes with transition metal ions, spectral studies are in progress to investigate this possibility. Further work in epoxidizing a variety of olefinic substrates as well as amines, sulphides, etc is in progress.

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**SALINITY STRESS RESPONSE OF PLANTS AND CALLI IN WHEAT**

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Following reports1-3 of several fold intra-specific heritable differences in adaptation to saline environment, many investigators have advocated the use of tissue culture technique for identifying salt-tolerant genotypes for generating new genetic variability for this purpose4-6. This approach towards developing more salt-resistant crop varieties is supported by observations in several nonhalophytes revealing sizable inter-specific differences at the cellular level in respect of salt tolerance7. In addition, halophytes have developed more successful adaptive strategies, based on certain anatomical and morphological features, to live with excessive salt concentrations8,9. In this context, the present investigation on plants and calli of two wheat genotypes, var HD 4502 and C 306, was undertaken to study their comparative response to salinity stress.

Plants were grown in 20 kg capacity glazed porcelain pots with four repeats, each having five plants. Saline soil grade (ECe 7.0 dS m$^{-1}$) was prepared following the procedure described in the US Salinity Laboratory Handbook No. 60 using NaCl. Salinity level was maintained by flushing and saturating the soil with saline water of 7.0 dS m$^{-1}$ conductivity. Controls were raised in non-saline soil under comparable conditions. Performance of var