

ON THE SPIROCHAETE BACTERIA INFESTING THE CRYSTALLINE STYLE OF BIVALVIA (PHYLUM: MOLLUSCA)

P. SHAHUL HAMEED and A. L. PAULPANDIAN

Centre of Advanced Study in Marine Biology,
Parangipettai 608 502, India.

THE spirochaete bacteria, belonging to the genus *Cristispira* are large ($150\ \mu\text{m} \times 3\ \mu\text{m}$), motile, without flagella and with spiral axis¹ and reported to infest the crystalline style of several bivalve species²⁻⁷ since their discovery by Certes². Although there are about 20,000 described bivalve species⁸, only 62 species have been reported to harbour *Cristispira* in the crystalline style and none reported from India. The present communication records the occurrence of *Cristispira* in the style of estuarine bivalves belonging to the species, *Crassostrea madrasensis*, *Meretrix meretrix*, *M. casta* and *Katelysia opima*.

During the present investigation, specimens of 12 bivalve species were collected from the marine and estuarine habitats of Parangipettai ($11^{\circ}29'$ N lat. and $79^{\circ}46'$ E long.). The crystalline styles were removed by dissection from the bivalves and washed in sterile distilled water. The style was examined for the presence of *Cristispira* under low power of microscope in the dark field illumination. For measuring morphological characters, the style was softened in the sterile filtered sea water for 10 min and a bit of style was squashed and observed under high power. The bacteria were also stained in Giemsa, following the method of Berkeley⁶.

It is evident from table 1 that only the estuarine oyster, *C. madrasensis* and clams *M. meretrix*, *M. casta* and *K. opima* carry a dense population of *Cristispira* in their styles and none of the marine

bivalves harboured the bacteria. The style of every *C. madrasensis* examined was colonised by *Cristispira* and in the clams the percentage of infection ranged between 87-90%.

The morphological characters of *Cristispira* recorded in the four species varied within a narrow range (table 1). The striking common features are the pointed ends, cell length ($46-57\ \mu\text{m}$), cell width ($1.6-2.1\ \mu\text{m}$) and the number of turns (4-5) in the helix (figure 1). In general, they resemble the characters of *Cristispira anodontae* as described in the Bergey's manual⁵. Since *in vitro* axenic cultivation of *Cristispira* still remains a challenge in this field the taxonomic relationship between the 20 morpho-species described so far could not be established¹.

The bacteria were most abundant in the style of *C. madrasensis*. The soft nature of its style perhaps may provide suitable environmental conditions thus accounting for greater abundance of the bacteria recorded among the bivalves. In all the four bivalve species, the style has a central semi-fluid core surrounded by firmer crystalline layers (figure 2). Along the style matrix the bacteria move back and forth by undulation, the direction of movement being non-specific. The spirochaetes were distributed in large numbers in the central core than in the crystalline layers in which they orient themselves in the space between layers (figure 2). A similar differential distribution of the bacteria within the style was reported by Noguchi³.

However, at the extreme gastral end of the style projecting into the stomach lumen, they were absent in all the four bivalve species. Berkeley⁶ made similar observation in the style of *Saxidomus* and opined that *Cristispira* is anaerobic and the toxic oxidising condition prevailing at the gastral end may cause the bacteria to retreat from the region. But *in vitro*

Table 1 Range of morphological characters (μm) of *Cristispira* in *C. madrasensis*, *K. opima*, *M. meretrix* and *M. casta*

Host Species	Number examined	Cell length	Cell width	Spiral wave-length	Spiral width	Number of turns
<i>C. madrasensis</i>	90	34.1-55.8 (46.2)	Constant (1.6)	6.2-15.5 (8.9)	3.1-9.3 (5.8)	2.5-6.0 (4.3)
<i>K. opima</i>	120	27.9-55.8 (56.8)	1.0-3.1 (2.0)	6.2-10.9 (8.5)	4.2-6.2 (5.2)	3.0-5.5 (3.8)
<i>M. meretrix</i>	120	31.0-59.0 (46.8)	1.6-3.1 (2.1)	9.3-17.1 (12.2)	6.2-12.4 (7.3)	3.0-5.0 (4.0)
<i>M. casta</i>	120	31.0-68.2 (46.2)	1.6-3.1 (1.9)	9.3-15.5 (11.0)	1.6-4.7 (3.1)	3.0-6.5 (5.4)

Figures in parantheses indicate mean values

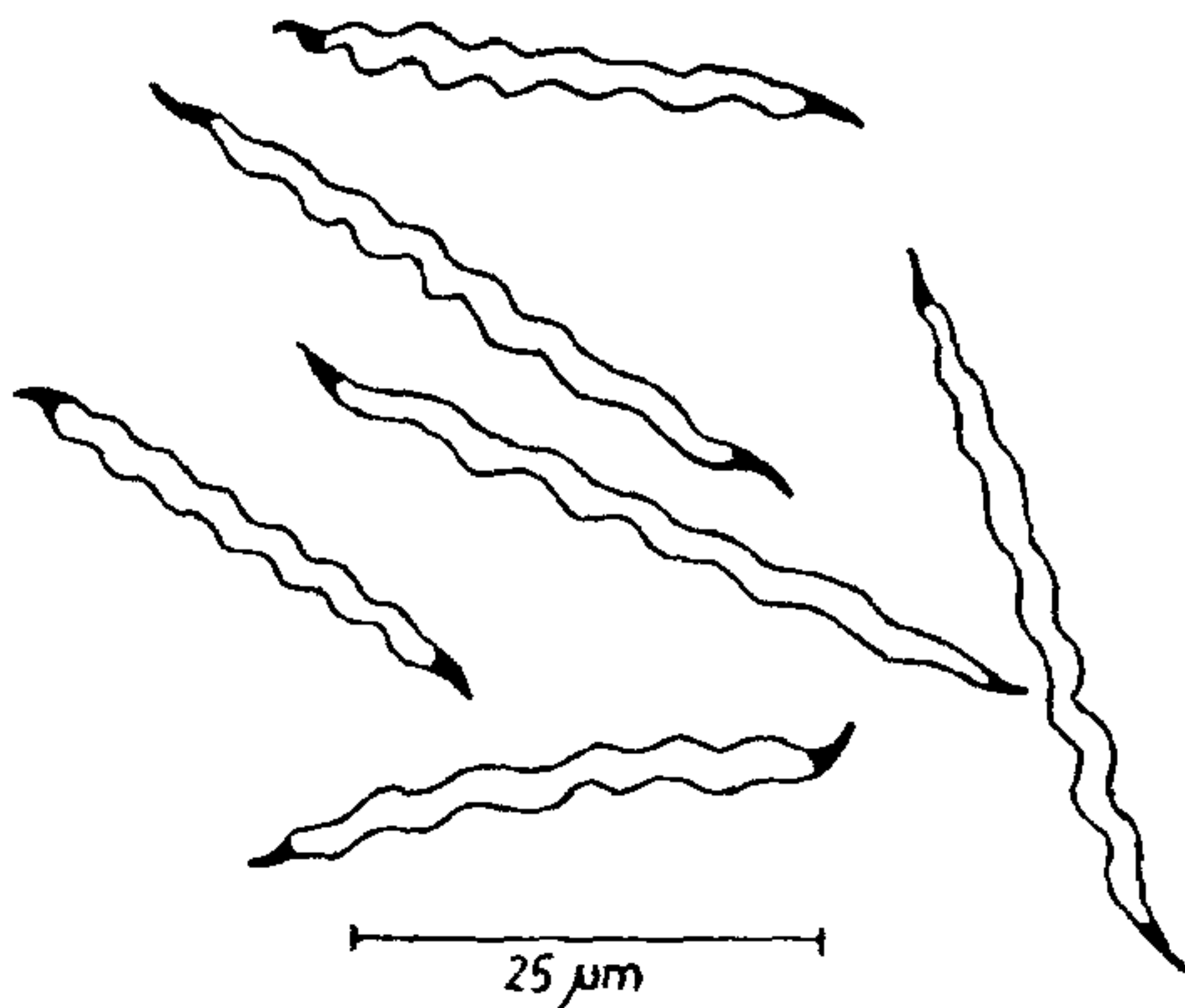


Figure 1. Structure of *Cristispira*.

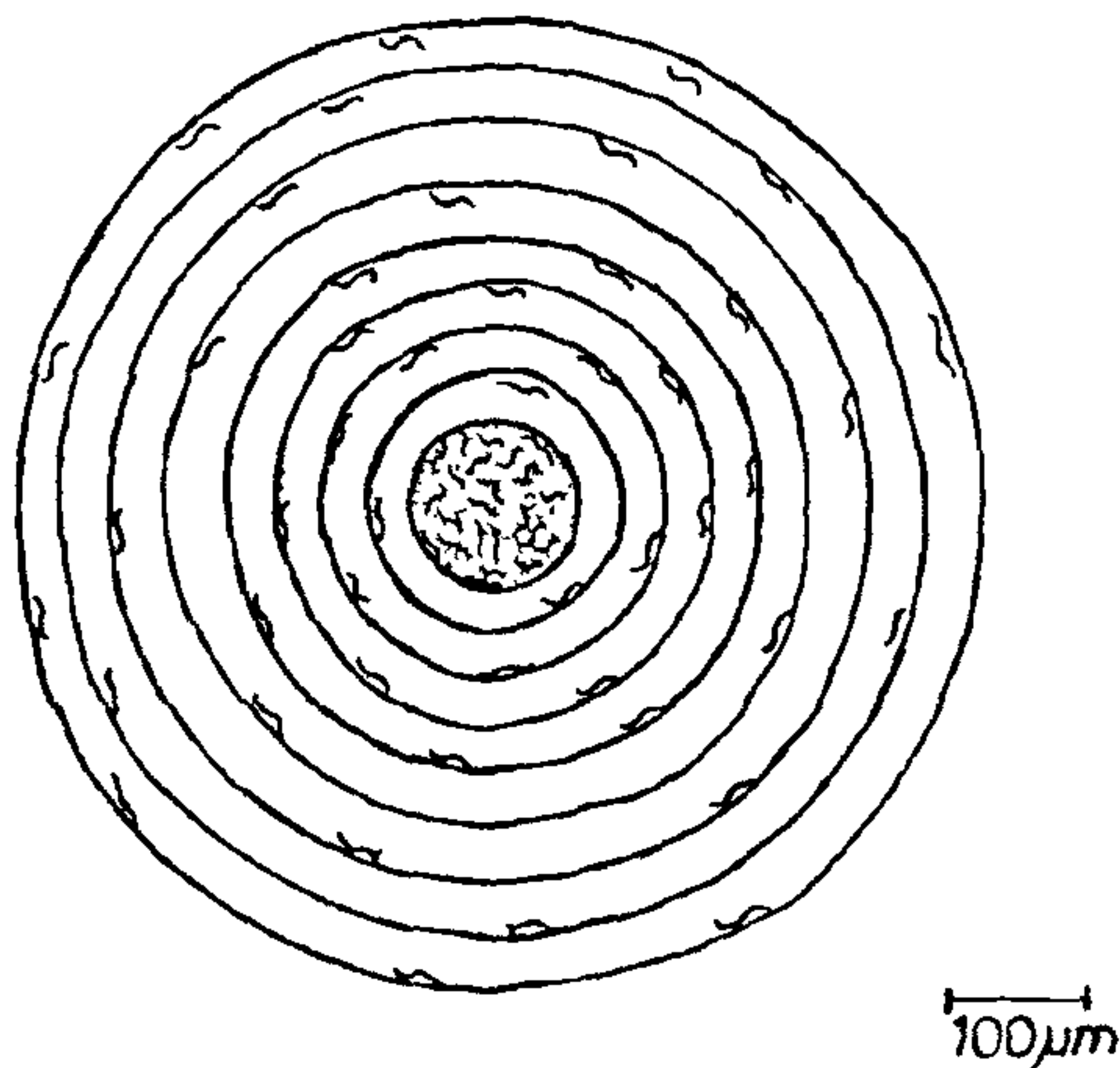


Figure 2. Transverse section of crystalline style of the oyster *Crassostrea madrasensis* showing the distribution of *Cristispira*.

observation of the style under low power of microscope, showed that the central core of the style flows out continuously through the gastral end and therefore, the absence of *Cristispira* may be attributed to the negative response (negative taxis) of the bacteria to this anteriorly directed flow.

Crystalline style is a secreted mucoprotein rod containing several carbohydrases and thus aids the extracellular digestion in stomach⁹. Morton¹⁰ sug-

gested that cellulase activity of crystalline style may be directly linked with *Cristispira* population. The quantitative study of style cellulase activity revealed that a true cellulolytic activity was absent in all the four bivalve species that carry the bacteria (Personal observation). Further, Horiuchi and Lane¹¹ and Payne *et al.*¹², also proved that the cellulolytic property of style is molluscan but not bacterial. The bivalves inhabit the shallow estuarine water wherein organic matter undergoing degradation is abundant promoting the transmission of bacteria.

Dimitroff⁴ considered them as parasites, Berkeley⁶ as facultative anaerobes. Since the infection of *Cristispira* does not seem to affect the well-being of the host, it could be regarded as a commensal.

The authors are thankful to the Director for facilities. The fellowship awarded to PSH by UGC, New Delhi is gratefully acknowledged.

30 January 1984

1. Kuhn, D. A., *The prokaryotes*, Springer-Verlag, Berlin, 1981, 555.
2. Certes, A., *Bull. Soc. Zool.*, France, 1882, 7, 374.
3. Noguchi, H., *J. Exp. Med.*, 1921 34, 295.
4. Dimitroff, V. T., *J. Bacteriol.*, 1926, 12, 135.
5. Bergey, D. H., *Manual of determinative bacteriology*, Williams and Williams Co., Baltimore, 1957, 1094.
6. Berkeley, C., *Can. J. Zool.*, 1959, 37, 53.
7. Bernard, F. R., *The Veliger*, 1970, 13, 33.
8. Barnes, R. D., *Invertebrate zoology*, Saunders College, Philadelphia, 1980, 1089.
9. Purchon, R. D., *The biology of mollusca*, Pergamon Press, Oxford, 1968, 560.
10. Morton, J. E., *Biol. Rev.*, 1960, 35, 92.
11. Horiuchi, S. and Lane, C. E., *Comp. Biochem. Physiol.*, 1966, 17, 1189.
12. Payne, D. W., Thorpe, N. A. and Donaldson, E. M., *Proc. Malac. Soc. London*, 1972, 40, 147.