

Fig. 3. Liver 14 days after exposure to 900 R of gamma rays showing a mononucleated giant hepatocyte (arrow) × 400.

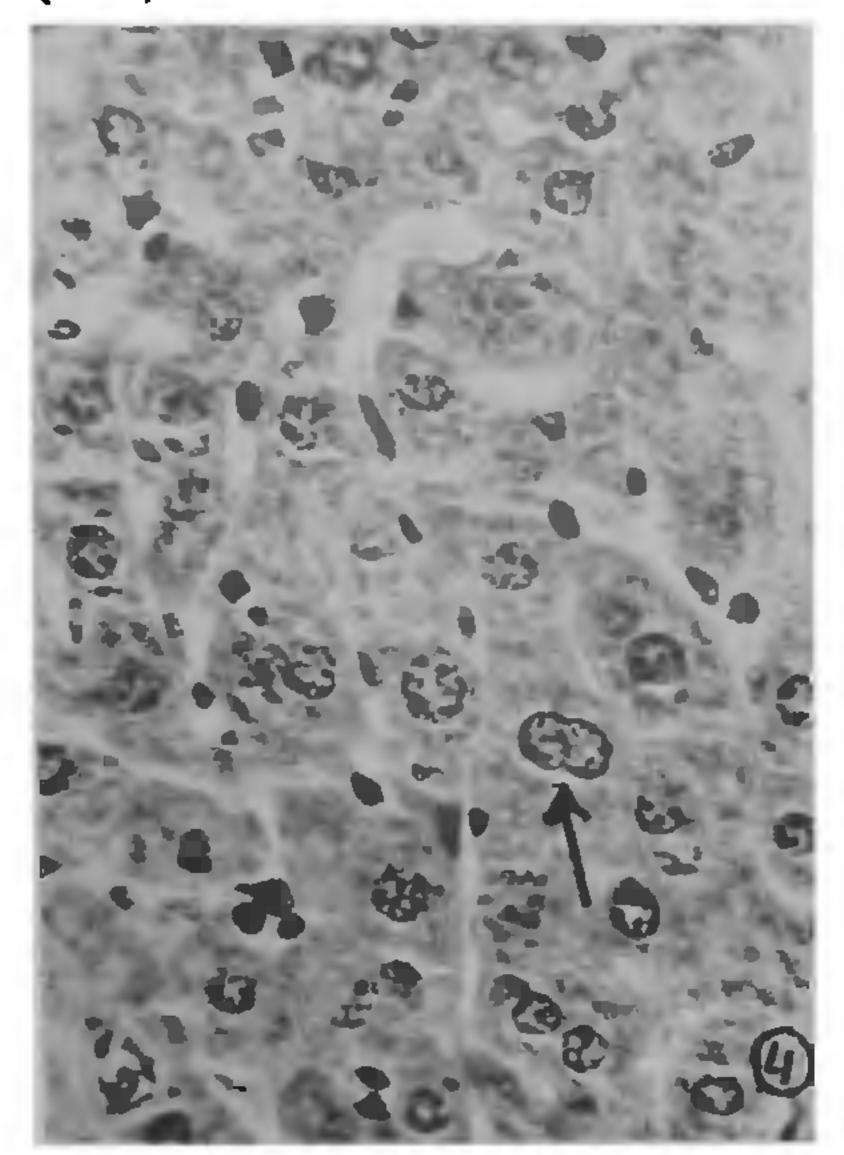


Fig. 4. Liver 14 days after exposure to 900 R of gamma rays showing the fusion of nuclei in a binucleated cell (arrow) × 400.

phenomenon⁴ and it seems to be a step before degeneration and cell death⁸.

One of us (MLG) is thankful to UGC for providing Teacher-Fellowship under the Faculty Improvement Programme.

August 16, 1980.

- Montgomery, P. O. B., Karney, D., Reynolds, R. C. and McClendon, D., Am. J. Pathol., 1964, 44, 727.
- 2. Altman, K. I., Gerber, G. B. and Okada, S., In Radiation Biochemistry, 1970, 1, 239.
- 3. Bhatia, A. L., Curr. Sci., 1975, 44, 470.
- 4. Rao, A. R. and Srivastava, P. N., Experientia, 1967, 23, 381.
- 5. Dev, P. K., A Thesis Approved by University of Rajasthan, Jaipur, 1971.
- 6. Gupta, M. L., Unpublished.

14CO, INCORPORATION STUDIES IN PEANUT (ARACHIS HYPOGAEA L.) UNDER PHOSPHORUS DEFICIENCY

G. Gururaja Rao*, S. K. Mahaboob Basha and G. Rajeswara Rao

Department of Botany, Sri Venkateswara University Tirupati 517 502, India

* Present address: Plant Physiology Division, Rubber Research Institute of India, Kottayam 686 009, Kerala, India.

Plants usually form characteristic symptoms in response to the lack of essential elements. Phosphorus with nitrogen and potassium is classified as a macronutrient. A good supply of phosphorus has been associated with increased root growth. Deficiency of phosphorus causes stunted plant growth and delayed maturity¹. Phosphorus is indeed the ubiquitous and essential element in the energy transfer processes such as photosynthesis which is so vital to life and growth². Peanut, an important oil yielding crop showed decreased growth and altered metabolism under phosphorus deficiency³. In the present study an attempt is made to find out the effect of phosphorus deficiency on ¹⁴CO₂ uptake in peanut.

Uniform seeds of peanut (Arachis hypogaea L. Var-TMV-2) were surface sterilized in 0.1% HgCl₂ solution, washed repeatedly in distilled water and sown in porcelain pots containing acid washed sand. The cotyledonary leaves were excised gently on 10th day after sowing and thinning was done to 3 plants per pot. The pots were divided into two sets and the

TARIE I

Radioactivity (Mean of three replications) ¹¹CC₂ incorporation in peamet leaves under phosphorus deficiency.

	7.000 T			Alco	Alcohol saluble fraction	ble fraci	ion				**	Alcohol ir solul	ir solubl	ble fraction	ū		
Age in	recuing time		Š	Centrol			P-Def ciert	iert			သိ	Control			P-Deficient	'nt	
udys	(Hilling)	TI	AAF	OAF	SuF	TI	AAF OAF	}	SuF	II	AAF	OAF	SuF	12	AAF	OAF	SuF
20*	10	490-3	490-3 147-1	137.3	2.35-9	373.2	235.9 373.2 155.0 140.2	146.2	78.0	386.2	186.7	64.9	64.9 134.6	2.937	62.0	62.0 103.6	40.6
	20	636.4	190.9	241.8	203-7	438.1	170.9	170.9 162.2	105-0	596.8	236.8	134.8	225.2	491.9	206.0	149.7	136.2
30**	10	530.8	153.9	132.7	244.2	4.24.9	123.9	173.8	127.2	492.7	103.8	126.2	262-7	456.4	176.9	177.5	102.0
	20	719.0	172.4 158.0	158.0	388.6	465.2	134.9	162.8	167.5	673-6	163.9	0.66	410.7	516.6	117.3	182.6	216.7

Total incorporation; - counts per minute; TI - 20 days after treatment; c.p.m.--Sugar fraction. SuF-• — 10 days after treatment; Organic acid fraction; Sul

first set received the nutrient solution of Hoagland and Arnon⁴ which served as control. Phosphorus deficiency was created by replacing KH₂PO₄ with KHSO₄ in the second set. All the plants received the respective nutrient solutions once in 3 days.

14CO, feeding technique as described by Berry et al. 5 was adapted with 10 and 20 min as feeding time. The incoporation studies were carried out in 20 day (10 days after treatment) and 30 day (20 days after treatment) old plants. The first compound leaf from the base was cut under water and used for feeding. After the selected periods of feeding the leaves were killed by plunging in boiling 80% ethanol, extracted and centrifuged. The supernatant was taken out and the residue was re-extracted with ethanol and centrifuged. The two alcohol soluble fractions were pooled and the alcohol insoluble residue was hydrolysed with 6N HCl. After hydrolysis, the acid was evaporated and the resulting residue was taken in 80% ethanol constituting the alcohol insoluble fraction. Both alcohol soluble and insoluble fractions were evaporated to a known volume and an aliquot was examined for radio activity using end window GM counter. Preparation of water soluble fraction and fractionation of alcohol soluble and insoluble fractions into amino acids, organic acids and sugar fractions were done according to Atkins and Canvin⁶. Total ¹⁴CO₂ incorporated into each of these fractions was determined as mentioned earlier. Triplicate. samples were maintained for each experiment.

The results of the present study indicated a decreased ¹⁴CO₂ incorporation into alcohol soluble and insoluble fractions under P-deficient conditions. Incorporation rates were high 20 min after feeding both in the 20 day and 30 day old plants (Table I). A decrease of 23.88 and 31.16% incorporation over controls after 10 min and 20 min feeding time in 20 day old plants; and 19.95 and 35.30% decrease into alcohol soluble fraction in 30 day old plants was observed under P-deficiency. ¹⁴C incorporation into alcohol insoluble fraction showed less decrease except in 20 day old plants after 10 min feeding; 53.39, 17.58% in 20 day old plants and 7.37 and 23.31% decrease in 30 day old plants was observed at 10 and 20 min feeding time respectively, under phosphorus deficiency.

Fractionation of alcohol soluble and insoluble fractions indicated higher rates of incorporation into amino acids and organic acids under phosphorus; deficiency. However amino acids of alcohol insoluble fraction showed lowered incorporation in 20 day old. P-deficient plants initially, i.e., 10 min after feeding. But ¹⁴C incorporation into sugar fraction was found to decrease significantly both in 20 and 30 day old P-deficient plants (Table I).

Phosphorus is one of the essential elements that plays a pivotol role in plant metabolism. Deficiency of P in nutrient medium results in stunted growth of peanut plants and lowers the uptake of potassium³.

The decreased potassium in turn reduces stomatal opening, which ultimately results in poor uptake of ¹⁴CO₂. The lowered K⁺ under P-deficiency as observed in our previous studies2 may even be responsible for the reduced photosynthesis and increased respiration, thereby lowering the amounts of carbohydrates. Our previous studies^a also indicated low amounts of sugars in peanut leaves under P-deficiency. Thus the lowered incorporation into alcohol insoluble fraction indicates lowered synthesis of starch under P-deficiency. High rates of incorporation into amino acids under P-deficiency is explained in terms of their lowered utilization in protein synthesis. High rates of incorporation into organic acids and poor rates of incorporation into sugars indicate that the conversion of organic acids to carbohydrates may be slowed down under P-deficiency,

Phosphorus compounds have been shown to be essential for photosynthesis, the interconversion of carbohydrates and related compounds, fat metabolism and a host of other life processes. The reduced rates of ¹⁴CO₂ uptake in peanut leaves as observed in the present study indicates that the poor growth of plants might be due to lesser availability of carbohydrates for the vital growth processes.

GGR is grateful to CSIR (New Delhi) for providing financial assistance.

September 2, 1980.

- 1. Salisbury, F. B. and Ross, C., Plant Physiology, Prentice-Hall of India Pvt. Ltd., New Delhi, 1977.
- 2. Tisdale, S. L. and Nelson, W. L., Sail Fertility and Fertilizers, Macmillan Pub. Co., Inc., New York, 1975.
- 3. Mahaboob Basha, S. K. and Rajeswara Rao, G., "Physiological changes in peanut (Arachis hypogaea L.) under phosphorus deficiency; Indian J. Pl. Physiol., 1981 (in press).
- 4. Hoagland, D. R. and Arnon, D. I., "Water and sand culture methods for growing plants without soil; Cir. 347, Calif. Agric. Expt. Stn., 1950.
- 5. Berry, J. A., Downton, W. J. S. and Tregunna, E. B., Can. J. Bot., 1970, 48, 777.
- Atkins, C. A. and Canvin, D. T., Ibid., 1971, 49, 1225.

PRESS MUD AS ADDITIVE TO INCREASE BIOGAS PRODUCTION FROM CATTLE WASTE

RAJENDRA SINGH, M. K. JAIN AND P. TAURO Department of Microbiology Haryana Agricultural University Hissar 125 004, India

of P in nutrient medium results in stunted growth of Anaeronic digestion depends on several different peanut plants and lowers the uptake of potassium. microbial activities functioning concurrently and at