Dose	Duration (hr)	Total % of aberrations	Mitotic index	DNA content (mg/g)	
				Liver	Spleen
10 mg/kg	24	46	2.95	0.75	1.29
20 mg/kg					
(10  mg/24  hr)	48	50·7	2.29	0 · 75	1.37
30 mg/kg	72	55.7	3.33	0 · 79	1 · 37

TABLE I Alserations in the nuclear contents of rat with esmone

increase in the quantity of DNA in the liver and spleen (Table 1), showing that estrone is a cell prolific agent.

(10 mg/24 hr)

Although the chromosomal aberrations were found in many cells, their type and extent varied from cell to cell even in the same individual. The gaps and breaks were randomly distributed and no particular chromosome was affected. These aberrations were of the chromatid type showing that most probably the damage to the chromosomes occurred, while they were in the  $G_2$  phase.

The induction of small chromosomal deletions or translocations results in significant changes in neoplastic initiation. These rarely arise as a result of direct damage to DNA, but are mainly due to indirect disturbances in chron-osome synthesis. The analysis of chromosomes can, therefore, be one of the means for screening cancer.

September 10, 1980.

1. Lacassagne, A., Les Cancers Produits par des Chimiques Endogenes, Hermann, Substances Paris, 1950.

## ON A RARE TREMATODE, TRANSVERSOTREMA CHAUHANI N.SP., FROM A FRESHWATER FISH, NANDUS NANDUS (HAM.)

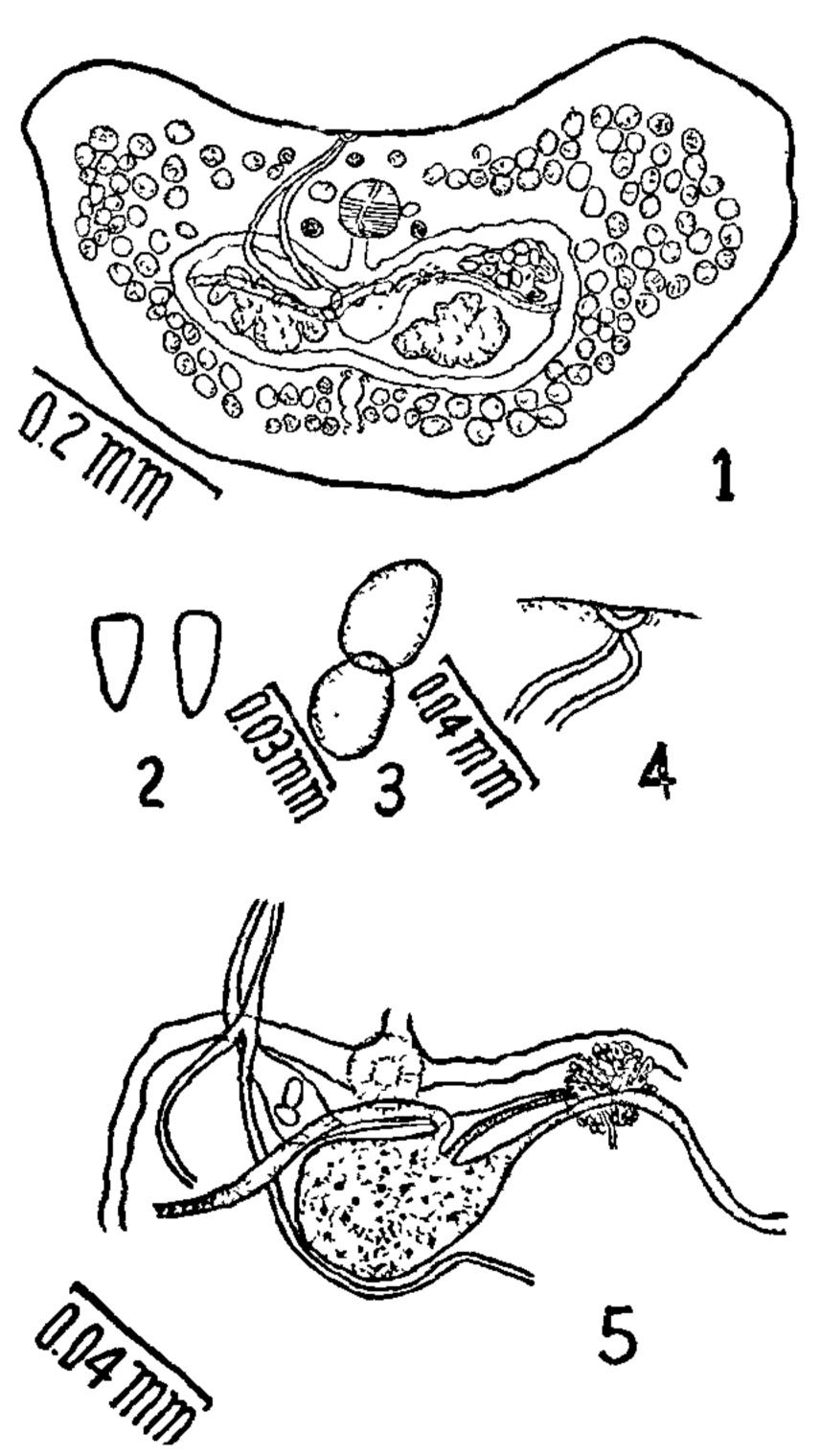
NIRUPAMA AGRAWAL AND HRIDAYA SHANKER SINGH Zoology Department, Lucknow University, Lucknow

Transversottema chauhani n.sp.

Host: Nandus nandus (Ham.); Host examined: 220; Host infected: 1; specimens collected: 2; Locality: Lucknow.

anterior margin, spinose, spines triangular and equal, Fig. 5. Portion of body enlarged showing ootype

increase in the dose. This was in relation to an black, round, at the level of pharynx, 0 01 mm. Oral sucker absent. Pharynx subterminal, spherical, 0.04- $0.05 \times 0.05$ -0.06 mm. Oesophagus short, 0.03-



Figs. 1-5. Fig. 1. Transversoterma chauhani n.sp. Fig. 2. Body spines enlarged. Fig. 3. Eggs. Fig. 4. Body flattened, transversely elongated, concave Portion of body enlarged showing genital aperture.  $0.30-0.44 \times 0.64-0.71$  mm. Eye spots, one pair, complex, vitelline reservoir and portion of uterus,

0.04 mm. Intestinal caeca cyclocoelid, margins crenated, in median body region. Ventral sucker behind the pharynx, 0.028-0.030 mm in diameter. Ventral sucker half the size of pharynx.

Testes lobed, post-acetabular, intercaecal. Left testis  $0.05-0.07 \times 0.04-0.06$  mm. Right testis 0.05- $0.06 \times 0.06 - 0.07$  mm. Cirrus sac absent. Vesicula seminalis S-shaped, near left testis. Ejaculatory duct narrow, opening in genital atrium, at anterior body region. Ovary small, oval, diagonal to right testis,  $0.050-0.054 \times 0.05-0.06$  mm. Laurer's canal not visible. Oviduct short, opening at ootype. Vitelline follicles extracaecal, confluent anteriorly. Two long yolk ducts, one from either body side, opening into vitelline reservoir. Uterus passing towards right, over intestinal caeca, parallel to ejaculatory duct, opening into genital atrium through short metraterm. Eggs, two,  $0.030-0.033 \times 0.025-0.029$  mm. Excretory bladder short, tubular, opening outside through excretory pore. The genus Transversotrema, with T. hassi as the type, was erected by Witenberg<sup>5</sup> for worms, collected from an unidentified piscine host from Red sea. Subsequently, T. patialensis Crusz, Ratnayke and Sathananthan<sup>1</sup>; T. larui Velasquez<sup>4</sup>; T. licinum Manter<sup>2</sup> and T. soparkari Pande and Shukla<sup>3</sup> were described under the genus.

The present form differs from T. hassi in the presence of eye spots (absent in T. hassi) and diagonal ovary which is above right testis in T. hassi. In T. licinum, T. soparkari and T. parialensis, the vitelline follicles are not confluent in anterior region and are intercaecal also but in the present form they are confluent anteriorly and are extracaecal. In T. lauri, the body is oval and the pharynx and acetabulum are of same diameter but in the present form, it is semilunar and the pharynx is nearly double the size of acetabulum.

Thanks are due to Prof. B. K. Tandan for laboratory facilities, to Dr. K. C. Pandey for helpful suggestions and to SCS T for financial assistance.

October 23, 1980.

- 1. Crusz, H., Ratnayke, W. E. and Sathananthan, A. H., Cey. J. Sci. Biol. Sci., 1964, 5, 8.
- 2. Manter, H. W., J. Parasit., 1970, 56, 486.
- 3. Pande, B. P. and Shukla, R. P., Curr. Sci., 1972, 41, 682.
- 4. Velasquez, C. C., J. Parasit., 1958, 44, 449.
- 5. Witenberg, G., Ibid., 1944, 30, 179.
- 6. Yamaguti, S., Systema Helndudum, Int. Pub., New York, 1958, Vol. 1, 1.

## ROLE OF THYMUS AND BONE MARROW CELLS IN IMMUNITY TO NEMATOSPIROIDES DUBIUS IN MICE\*

M. Vyas, P. N. Lakshmi, P. K. Sanghvi and G. N. Johri

Helminthology-Immunology Lab.
School of Studies in Zoology
Vikram University, Ujjain (M.P.), India

Singly sensitized thymus and bone marrow cells from donors infected with 100 Nematospiroides dubius larvae were injected into three groups of recipients A  $(40 \times 10^4)$  thymus cells, B  $(40 \times 10^4)$  bone marrow cells and C  $(20 \times 10^4)$  each of thymus and bone marrow cells mixture) and then challenged 14 days after cell transfer. It was found that recipients with mixture of thymus and bone marrow cells (group C) produced significant immunity in comparison with those with either cell population, alone.

## Introduction

Transfer of adoptive immunity in recipients through different cell populations from infected donors concerning parasitic infections have been reviewed by Larsh<sup>1</sup>. Recently several workers<sup>2-6</sup> have successfully transferred immunity through a variety of cell populations in several infections.

In the case of Nematospiroides dubius, Cypess and Sanghvi, Vyas and Johri<sup>8-10</sup> transferred adoptive immunity in mice through spleen, peritoneal exudate, mesenteric lymph node and thymus cells respectively. Since thymus cells from sensitized donors were successfully employed to transfer delayed hypersensitivity in mice<sup>11,12</sup>, it was, therefore, thought worthwhile to investigate whether thymus and bone marrow cells singly or together would transfer immunity in the N. dubius model.

## Materials and Methods

Infective larvae of N. dubius were cultured according to the method of Van Zandt<sup>13</sup>. Thirty female Swiss albino mice of approximately 20-23 g wt. and 6-8 week old were inoculated per os with 100 N. dubius larvae; after 14 days thymus and bone marrow cells were collected and suspended in Ringer's solution and approximately  $40 \times 16^4$  cells were injected intraperitoncally into 3 separate syngeneic groups within 4 hours after their collection. Group A received  $40 \times 10^4$  thymus cells, group B  $40 \times 16^4$  bone marrow cells and group C mixture of thymus  $(20 \times 10^4)$  and bone marrow  $(20 \times 10^4)$  cells. Groups D, E and F with unsensitized cells collected from a batch

<sup>\*</sup> Presented at the Third National Congress of Parasitology, Haryana Agricultural University, Hissar, April 1980.