- 7. Vijayalakshmi, S., Murali Mohan, P. and Sasira Babu, K., J. Ins. Physiol., 1977, 23, 195.
- 8. Tirri, R., Lagerspetz, K. Y. H. and Kohonen, J., Comp. Biochem. Physiol., 1973, 44B, 473.
- 9. Fiske, C. H. and Subba Row, Y., J. Biol. Chem., 1925, 66, 375.
- Vijayalakshmi, S., Pavan Kumar, I., Murali Mohan, P. and Sasira Babu, K., Comp. Physiol. Ecol., 1977, 2 (2), 58.
- 11. —, and Sasira Babu, K., Abstract 2nd All India Symposium on Comparative Animal Physiology, December 13 to 15, 1976, Nagpur (India).
- 12. Harker, J. E., Nature (Lond.), 1954, 173, 689.
- 13. —, The Physiology of Diurnal Rhythms (Eds.)
 T. A. Bennett-Clark, George Salt, V. B. Wigglesworth), Cambridge Monographs in Experimental Biology, 1964, No. 13.
- 14. Brady, J., Adv. Ins. Physiol., 1974, 10, 1.

MIXED-COMMUNAL ROOSTING OF INDIAN WHITE BACKED VULTURES IN POONA

WHILE observing the roosting behaviour of some common birds of Poona (Maharashtra) and around, from January 1976 to February 1977, I came across number of mixed-communal roosts of birds.

Usually the associates which I have observed at such roosts are Indian Mynas, Acridotheres tristis (Linn.); House and Jungle Crows, Corvus splendens Vieillot and C. macrorhynchos Wagler; Pariah Kites, Milvus migrans (Boddaert); Rose ringed Parakeets, Psittacula krameri (Scopoli) and House Sparrows, Passer domesticus (Linn.). These associations at mixed-communal are noticed throughout the year. The Cattle Egrets, Bubulcus ibis coromandus (Boddaert) and Indian Pond Herons, Ardeloa grayii grayii (Sykes) also assemble at such mixed roosts only during the non-breeding season while a migratory bird like Rosy Pastor, Sturnus roseus (Linn.) roosts together along with the above species for a part of the year. This has also been indicated by Gadgil and Ali¹ (1975) who have given a systematic account of communal roosting of Indian birds.

During the month of June 1976, it was noticed for the first time that the Indian Whitebacked or Bengal Vultures, Gyps bengalensis (Gmelin) formed a mixed-communal roost along with the Indian Mynas, House and Jungle Crows and Whitenecked Storks, Ciconia episcopus (Boddaert). This mixed congregation was observed on the Poona-Bombay Road near Dapodi, on a Banyan tree, Ficus bengalensis L. On counting the number of

birds at this mixed roost, it was found that there were 150 Indian Whitebacked Vultures, 300 Indian Mynas, about 150 House and Jungle Crows and a pair of Whitenecked Stork. These countings were made in the evening, when these birds return to the roost. Further observations showed that this mixed-communal roost was constant till september 1976, both in the number of species and in their total population. Subsequently, the roost totally disappeared till February 1977. In the meantime, the Vultures and Storks seem to have migrated from this locality, while Mynas and Crows shifted to another roost about half a kilometer away from the original roosting place.

From the above observations it would appear that Indian Whitebacked Vultures are seen on this mixed-communal roost only during the monsoon from June—September, which is a part of their non-breeding season.

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Zoological Survey of India, ANIL MAHABAL. Western Regional Station, Poona 411 016, April 29, 1977.

1. Gadgil Madhav and Salim Ali, J. Bombay Nat. Hist. Soc., 1975, 72 (3), 716.

NEW PARASITES RECORDED ON THE SORGHUM SHOOTFLY, ATHERIGONA SOCCATA (RONDANI)

Various cultural and chemical control methods have been investigated and recommended for the control of shootfly (A. soccata), a serious pest of sorghum. However, very little work has been done on identifying and utilizing the natural enemies of this pest. This aspect is now being systematically investigated under the All India Co-ordinated Sorghum Improvement Project at New Delhi.

One to two week old shootfly infested seedlings, having freshly formed dead hearts, were collected from the field and were kept in separate glass jais containing 5 cm layer of moist sand. The number of dead hearts with the shootfly larvae were counted in each jar to determine the percentage of parasitism. The jars were placed at 27° C ± 1° C, the relative humidity ranged between 60 and 70%.

The parasites and shootfly adults emerging in jars were periodically collected and sent to British Museum for identification. It is found that in addition to the two parasites already recorded, viz., Aprostocetus sp. and Callitula bipartitus

(Kundu et al. 1971), the shootfly larvae were also parasitised by Ganaspis sp. (Eucoilidae), Psilus sp. (Diapriidae), Hemiptarsenus sp. (Eulophidae) and Daudmopsis sp. (Eulophidae).

The observation on the percentage of parasitism taken during different periods showed that except for 2% parasitism by Ganaspis sp. in the month of April, 1975, these parasites were recorded only in the infested seedlings collected during the months of September and October. Even in these months the extent of parasitism was rather low and ranged from 1 to 4%. The two parasites recorded earlier, viz. Aprostocetus sp. and Callitula bipartitus were reared out in the month of August also. The percentage parasitism of Aprostocetus sp. was found to be higher than any other parasite and ranged from 4% (October) to 15% (September).

Perusal of literature showed that these parasites have been recorded from a number of dipterous hosts from different parts of the world. Kerrich (1962)² reported Hemiptarsenus semialbiclavus (Gir.) parasitizing Agromyzid leaf miners of vegetable crops in Africa, while Psilus sp. has been recorded as pupal parasite of Pholeomyia comans (Diptera: Milichiidae) by Mosev and Neff (1972)3. Diaulinopsis sp. was rared from Agromyza pusilla Mg. (Diptera: Agromyzidae) from North America, U.S.A. (Thompson 1955)⁴. Eucoila (Ganaspis) haywardi. Blanch, has been used for the control of fruit flies, Anasterpha spp. (Diptera: Tephritidae) in Argentina (Turica 1968)⁵. Tho sorghum shootfly (A. soccata) has not so far been recorded as host of these parasites and thus constitutes a new host record. These four parasites also constitute first record from India.

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Division of Entomology, PREM KISHORE.
Indian Agricultural Research M. G. JOTWANI.
Institute, T. R. SUKHANI.
New Delhi 110 012, K. P. SRIVASTAVA.
April 27, 1977.

1. Kundu, G. G., Prem Kishore and Jotwani, M. G., Investigations on Insect Pests of Sorghum and Millets, Final Technical Report, Division of Entomology, IARI, New Delhi, 1971, p. 145.

2. Kerrich, G. J., Bull. ent. Res., London, 1969, 59 (2), 195,

3. Mosey, J. C. and Neff, S. E, Z. angew. Ent., 1972, 69 (4), 343.

Thompson, W. R., Parasitic Catalogue Sec.,
 Part 3—Host of Hymenoptera (Calliceratid to Evaniid), 1955.

5. Turica, A., Lucha Biologica como medio de control de Las moscas de Los frutos (Biological control as a means for the control of fruit flies), India, 1968, No. 241, p. 29 [RAE, 57 A, 1820].

CYANOPHAGE AC-1 INFECTING THE BLUE GREEN ALGA ANACYSTIS NIDULANS

AS-1 TYPE cyanophage which infects the unicellular algae, Anacystis nidulans and Synechococcus cedrorum was first characterized by Safferman et al¹. It is the largest BGA phage so far examined with a head diameter of 90 nm. and tail-head ratio of 3 to 1, based on a tail length of 243.5 nm. It has a rigid tail with a contractile sheath and a base plate with tail pins. In the present report, a new phage type infecting Anacystis nidulans 14011 and Chroococcus minor ARM was isolated from a waste stabilization pond inside the campus of the Indian Agricultural Research Institute, New Delhi².

The phage formed clear plaques of 4-6 mm after 10 days of incubation. Several blue green algal species of Nostoc, Anabaena, Tolypothrix, Aulosira and Spirulina, the green alga Chlorella vulgaris and the bacteria, Azotobacter chroococcuta, Rhizobium spp. and Rhodopseudomonas capsulatus were also tested for susceptibility to this phage, but none of them was found susceptible.

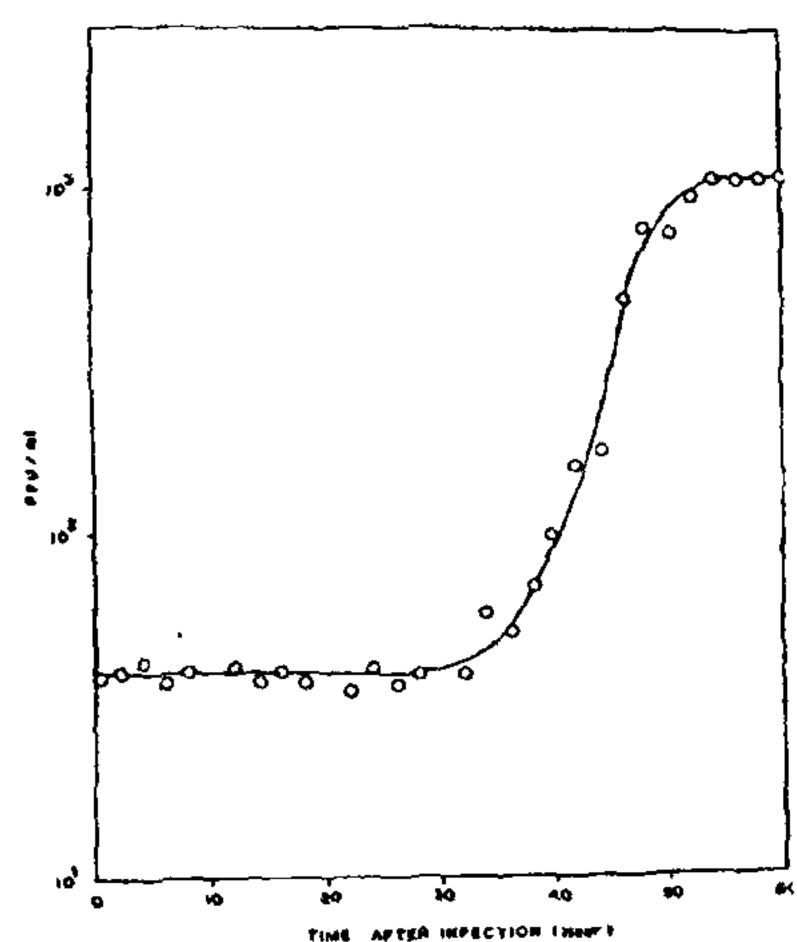


Fig. 1. One step growth curve of AC-1 cyanophage.