

- in broodstock of tiger shrimp, *Penaeus monodon* and other crustaceans of Andaman waters. *Indian J. Marine Sci.*, 2011, **40**(3), 403–406.
9. Saravanan, K. *et al.*, Overview of aquatic animal diseases in Andaman and Nicobar Islands. *J. Immunol. Immunopathol.*, 2015, **17**(1), 17–24.
 10. Lightner, D. V., Bell, T. A. and Redman, R. M., A review of the known hosts, geographical range and current diagnostic procedures for the virus diseases of cultured penaeid shrimp. *Adv. Trop. Aquacul.*, 1989, 113–126.
 11. Walker, P. J., Cowley, J. A., Spann, K. M., Hodgson, R. A. J., Hall, M. R. and Withyachumnarnkul, B., Yellow head complex viruses: transmission cycles and topographical distribution in the Asia-Pacific region. In *The New Wave: Proceedings of the Special Session on Sustainable Shrimp Culture, Aquaculture 2001* (eds Browdy, C. L. and Jory, D. E.), The World Aquaculture Society, Baton Rouge, Louisiana, 2001, pp. 227–237.
 12. Nielsen, L., Sang-oum, W., Cheevadhanarak, S. and Flegel, T. W., Taura syndrome virus (TSV) in Thailand and its relationship to TSV in China and the Americas. *Dis. Aquat. Org.*, 2005, **63**, 101–106.
 13. Flegel, T. W., Current status of viral diseases in Asian shrimp aquaculture. *Isr. J. Aquacul. – Bamidgeh*, 2009, **61**, 229–239.
 14. Bonami, J. R. and Lightner, D. V., Unclassified viruses of Crustacea. In *Atlas of Invertebrate Viruses* (eds Adams, J. R. and Bonami, J. R.), CRC Press, Boca Raton, Florida, 1991, pp. 597–622.
 15. Vega-Heredia, S., Mendoza-Cano, F. and Sanchez-Paz, A., The infectious hypodermal and haematopoietic necrosis virus: a brief review of what we do and do not know. *Transbound. Emerg. Dis.*, 2012, **59**, 95–105.
 16. Bruce, L. D., Trumper, B. B. and Lightner, D. V., Methods for viral isolation and DNA extraction for a penaeid shrimp baculovirus. *J. Virol. Methods*, 1993, **34**, 245–254.
 17. OIE, Manual of diagnostic tests for aquatic animals, Office International des Epizooties, Paris, 2015.
 18. Tang, K. F. J., Navarro, S. A. and Lightner, D. V., A PCR assay for discriminating between infectious hypodermal and hematopoietic necrosis virus (IHHNV) and the virus-related sequences in the genome of *Penaeus monodon*. *Dis. Aquat. Org.*, 2007, **74**, 165–170.
 19. Tamura, K., Stecher, G., Peterson, D., Filipski, A. and Kumar, S., MEGA6: molecular evolutionary genetics analysis version 6.0. *Mol. Biol. Evol.*, 2013, **30**, 2725–2729.
 20. Kimura, M., A simple method for estimating evolutionary rate of base substitutions through comparative studies of nucleotide sequences. *J. Mol. Evol.*, 1980, **16**, 111–120.
 21. Bell, T. A. and Lightner, D. V., A handbook of normal penaeid shrimp histology. Special Publication no. 1, World Aquaculture Society, Baton Rouge, 1988.
 22. Rai, P., Pradeep, B., Safeena, M. P., Karunasagar, I. and Karunasagar, I., Simultaneous presence of infectious hypodermal and hematopoietic necrosis virus (IHHNV) and type A virus-related sequence in *Penaeus monodon* from India. *Aquaculture*, 2009, **295**, 168–174.
 23. Flegel, T. W., Major viral diseases of the black tiger prawn (*Penaeus monodon*) in Thailand. *World J. Microbiol. Biotechnol.*, 1997, **13**, 433–442.
 24. Primavera, J. H. and Quintino, E. T., Runt-deformity syndrome in cultured giant tiger prawn *Penaeus monodon*. *J. Crustacean Biol.*, 2000, **20**, 796–802.
 25. Tang, K. F. J., Poulos, B. T., Wang, J., Redman, R. M., Shih, H. H. and Lightner, D. V., Geographic variations among infectious hypodermal and hematopoietic necrosis virus (IHHNV) isolates and characteristics of their infection. *Dis. Aquat. Org.*, 2003, **53**, 91–99.
 26. Rai, P., Safeena, M. P., Krabsetsve, K., La Fauce, K., Owens, L. and Karunasagar, I., Genomics, molecular epidemiology and diag-

nostics of infectious hypodermal and hematopoietic necrosis virus. *Indian J. Virol.*, 2012, **23**, 203–214.

ACKNOWLEDGEMENTS. This work was carried out under the National Surveillance Programme for Aquatic Animal Diseases (NSPAAD), coordinated by the ICAR-National Bureau of Fish Genetic Resources (NBFGR), Lucknow. The authors thank the Indian Council of Agricultural Research (ICAR) and National Fisheries Development Board (NFDB), Govt. of India, for financial support to carry out this work. The authors are grateful to the Referral Laboratory at ICAR-CIBA, Chennai for validating the IHHNV positive samples.

Received 13 June 2016; revised accepted 12 May 2017

doi: 10.18520/cs/v113/i10/2027-2031

How NaCl, Na₂SO₄, MgCl₂ and CaCl₂ salts affect the germinability of *Pinus halepensis* Mill.

Bouzid Nedjimi*

Laboratory of Exploration and Valorization of Steppe Ecosystem, Faculty of Science of Nature and Life, University of Djelfa, Cité Ain Chih, P.O. Box 3117 Djelfa 17000, Algeria

In the Mediterranean forests, *Pinus halepensis* Mill. (Aleppo pine) plays an important role against desertification, reforestation of degraded lands and soil rehabilitation. Therefore, knowledge of its seed germinability requirements is necessary for its propagation in field conditions to colonize new territories habitually not conventional for other species. The study was carried out to assess the effects of different soluble salts (NaCl, Na₂SO₄, MgCl₂ and CaCl₂) on seed germination characteristics [germination percentage (GP) and rate of germination (RG)] of this conifer. Data show that all soluble salts decreased both parameters GP and RG. The highest GP was obtained in conditions without salinity. The maximum values of germination were obtained by low concentrations of MgCl₂. Comparatively, NaCl was generally the most toxic salt followed by CaCl₂ and Na₂SO₄. The present findings could be useful in the design of future projects for reforestation of degraded arid lands.

Keywords: Aleppo pine, rate of germination, reforestation, saline soils.

ECO-PHYSIOLOGICAL studies about regeneration of endemic conifers species grown in arid and semi-arid areas and the factors influencing them are important for the

*e-mail: bnedjimi@yahoo.fr

protection and propagation of tree species in projects of restoration of degraded lands facing erosion^{1,2}.

Algerian saline soils are formed from the accumulation of various chloride and sulphate salts dominated by NaCl (>50%). The main salt components of these soils are Na⁺, Ca²⁺ and Mg²⁺ cations, and SO₄²⁻ is the second major anion after Cl⁻ (ref. 3).

In many studies, sodium chloride (NaCl) is the principal soluble salt examined to evaluate salt tolerance during germination of pine species^{4,5}. However, rare information exists about other soluble salts such as sodium sulphate (Na₂SO₄), calcium chloride (CaCl₂) and magnesium chloride (MgCl₂) that were present at higher levels in the saline soils of arid regions³.

The amount of soluble salts in arid soils can be greater than the tolerable limits for seed germinability of most conventional species, and therefore the emergence and propagation of plant species in these areas can be limited⁶.

Mediterranean species use many strategies to cope with the harsh-ecological conditions in their local biotope, such as salinity and drought⁷. Saline soils in the arid regions of Algeria contain multiple types of soluble salts, which have various influences on germination and the first growth stage of species³.

Aleppo pine (*Pinus halepensis* Mill.) is one of the most abundant endemic conifer species growing throughout the Mediterranean basin. However, in the recent decades, the main threat to its conservation is the devastation of natural forests in addition to lack of knowledge among the local population on the importance of *P. halepensis* forest ecosystems. Therefore, the possibilities of rehabilitation and restoration of this valuable tree need to be explored. In this context, information about seed germination requirements is of relevance to reforestation or replanting of degraded forests, and to colonize new territories habitually not conventional for other species.

In Algeria, *P. halepensis* constitutes the dominant coniferous tree of the natural and artificial forests, where it covers 852,000 ha area⁸. Notwithstanding its ecological and economic importance, exhaustive studies about seed germinability and seedling establishment of *P. halepensis* are required for propagation in field conditions. Therefore the present study was conducted in order to determine the effects of different types of soluble salts on germination of this species. Information from this study offers new knowledge about germination requirements of *P. halepensis* in saline conditions that can be used to enhance the chance of successful propagation under these conditions.

Cones of *P. halepensis* were collected in August 2014 from the natural forest of Gotaïa in the province of Djelfa, Algeria (2°47'E long., 34°37'N lat.; 1198 m asl). Seeds were detached from the cones and surface-sterilized with 60% alcohol (ethanol) for 10 min, followed by a treatment with 10% NaClO solution for 1 min and then washed abundantly with distilled H₂O.

Seeds were germinated in different concentrations (0, 50, 100 and 150 mM) of four soluble salts (NaCl, Na₂SO₄, MgCl₂ and CaCl₂); these concentrations reflect the range of salt content in the Algerian salt steppe⁹. The experiment was carried out in an incubator with a 12 h photoperiod under 15–25°C dark light. Radicle elongation of 2 mm was used as the criterion for germinability. A completely randomized design was applied in the germination tests. For each treatment, 100 seeds (in four replicates each) were placed in 90 mm petri dishes with 5 ml of test solution.

Modified Timson's index was applied to calculate the rate of germination (RG) as follows: $RG = \sum g/t$, where g is the percentage of seeds germinated after two-days interval and t is the total period of germination¹⁰.

A two-way ANOVA was carried out to test the effects of salts (S), concentration (C) and their interactions ($S \times C$) on the germination percentage (GP) and RG using the SPSS 9.0 software package. To ensure homogeneity of variance, data were arcsine converted before statistical analysis. Newman-Keuls test was applied ($P < 0.001$) for a comparison between treatments.

A two-way ANOVA revealed significant effect of S and C ($P < 0.001$) on final GP of *P. halepensis* seeds, but their interactions ($S \times C$) did not significantly affect GP (Table 1). GP decreased with increasing salinity, the highest GP was found in non-saline (control) treatment (Figures 1 and 2). In field conditions this might occur because soluble salts reduce osmotic potential (ψ_0) of the soil and prevent seed hydration¹¹.

A strong inhibition of GP was observed in NaCl solution and maximum GP was recorded at all concentration of MgCl₂. The level of toxicity for different soluble salts in decreasing order was as follows: NaCl > CaCl₂ > Na₂SO₄ > MgCl₂. Strongest regressions were recorded between germination (GP) and salt concentration (S) with a coefficient of determination (R^2) ranging from 0.57 to 0.89 (Figure 3).

RG values calculated using a modified Timson's index, decreased in response to increasing salinity (Figure 4). A two-way ANOVA of RG showed significant ($P < 0.001$) effect of the two variants (S and C) but not of their interactions ($S \times C$) (Table 1). The inhibition action of salt stress on RG was highest for NaCl treatment than compared to other soluble salts. However, the harmful effect of salt stress was generally less with MgCl₂.

Table 1. A two-way ANOVA of the effects of salts (S), concentration (C), and their interaction ($S \times C$) on germination characteristics of *Pinus halepensis* seeds

Independent variable	S	C	$S \times C$
Germination %	1.52*	41.56**	0.56 ^{ns}
Rate of germination	0.72*	52.52**	0.28 ^{ns}

Numbers represent F -values.

ns, Not significant; * $P < 0.05$; ** $P < 0.001$.

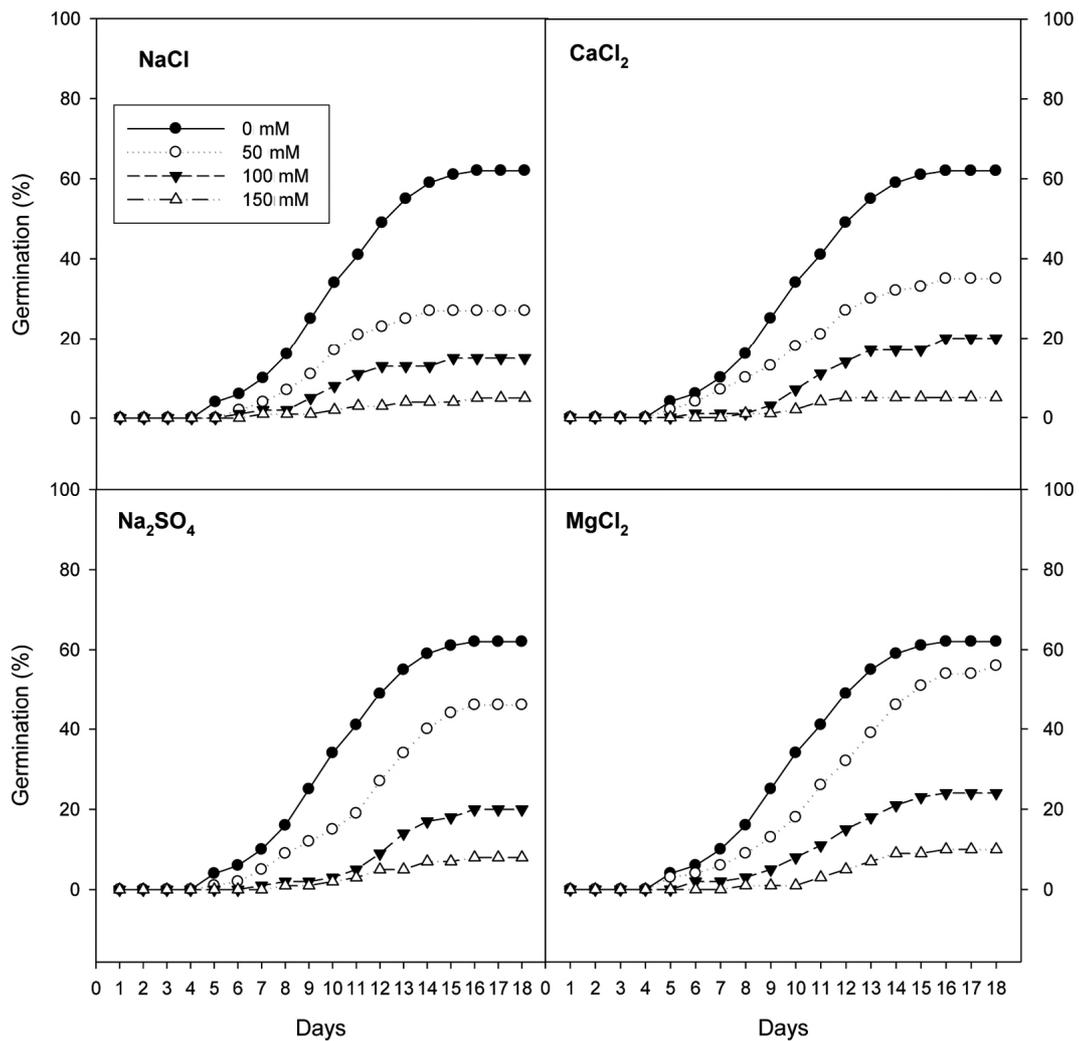


Figure 1. Mean germination percentage as a function of time of *P. halepensis* seeds treated with different soluble salts.

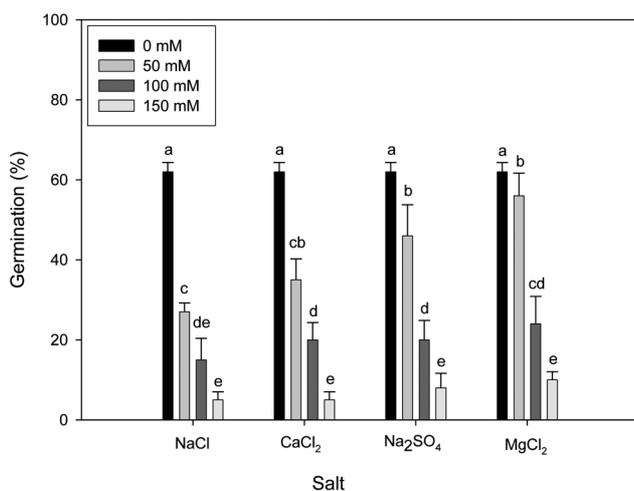


Figure 2. Final germination percentages of *P. halepensis* seeds treated with different soluble salts. Bars represent mean \pm SE ($n = 4$). Different letters indicate significant difference between treatments (Newman–Keuls test, $P < 0.001$).

In order to survive under harsh conditions (high soil salinity and water-deficit), salt-tolerant plants have adapted many strategies, such as delaying germinability to prevent seedling mortality due to lowest water potentials (ψ_w)¹². The ability of seeds to preserve their longevity under saline conditions for prolonged periods constitutes an imperative characteristic of salt-tolerant plants, permitting them to colonize the saline soils with low osmotic potential (ψ_0)¹³.

P. halepensis seeds showed a highest GP under non-saline conditions for all salinities. This result has been reported by several other studies, which indicates that optimal GP of different Mediterranean coniferous trees occurs under non-saline conditions¹⁴.

The probable reason for limited germination in the control treatments (reaching maximum of 62%) may be the dormancy of seeds. The enforced dormancy response in arid conditions is a selective strategy of plants growing in harsh habitats (drought and salinity). In these situations,

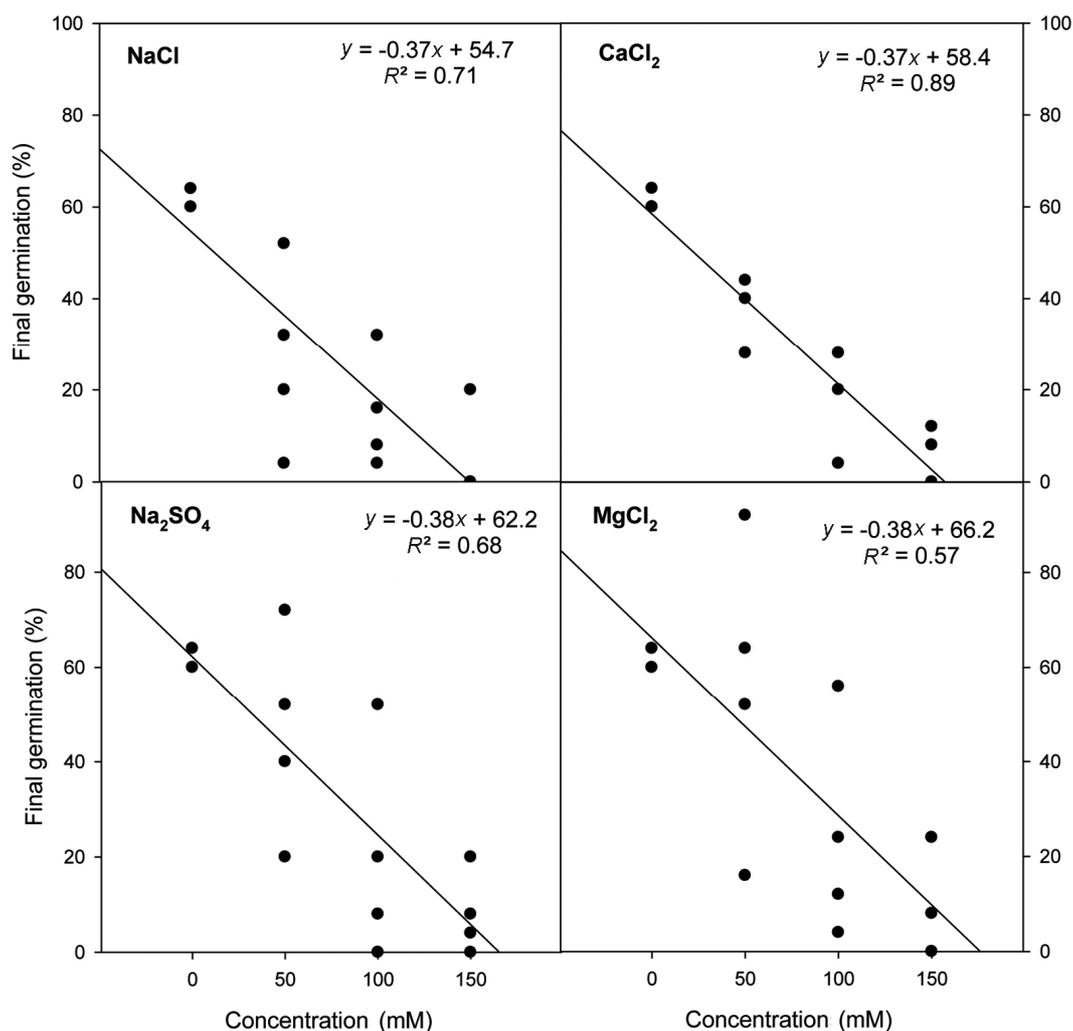


Figure 3. Regression plots of final germination percentage of *P. halepensis* seeds treated with different soluble salts.

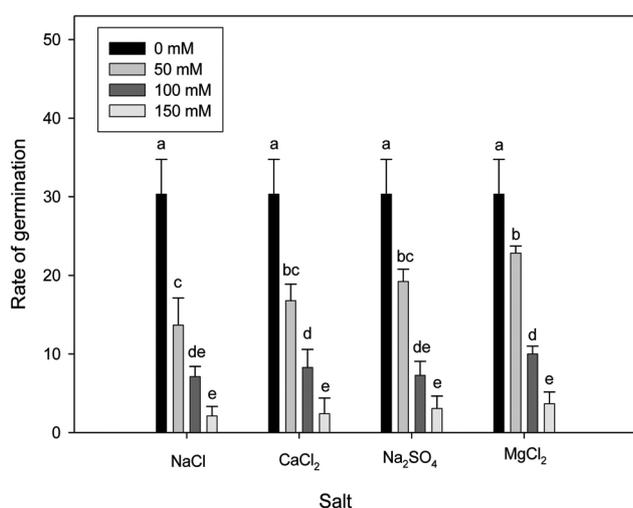


Figure 4. Rate of germination of *P. halepensis* seeds treated with different soluble salts. Bars represent mean \pm SE ($n = 4$). Different letters indicate significant difference between treatments (Newman-Keuls test, $P < 0.001$).

seed germination would be limited to periods when soil humidity and salinity levels are within the species tolerance limits¹⁵.

In the Mediterranean climate, rapid establishment of species after spring precipitation presents a successful key to plant colonization in saline environments. These species can propagate quickly and develop a deep root system which permits them to absorb water (H₂O) during drought from the profound soil horizons¹⁶.

GP of *P. halepensis* seeds decreased when salinity increased in the medium. Similar results were found in other *Pinus* species as well^{4,5,17}. This comportment may be due to high osmotic potential (ψ_0) that affects seed imbibition and delayed germination process.

In saline soils of arid regions, successful plant establishment depends on salt tolerance during seed emergence¹⁸. A critical phenomenon in these regions is high salt concentration at the superficial horizons of soils due to high evapo-transpiration causing germination failure by both osmotic and toxic effects¹⁹. However, seed

germinability under hyper-saline conditions arises after leaching by abundant precipitations²⁰.

In field conditions, germination of *P. halepensis* seeds occurs during spring season when soil salinity is reduced by winter rainfall. Germination after wet season confers an optimal benefit to species growing in saline environments. The rain water dissolves soluble salts from the superficial soil layers and provides water for seed imbibition. In this situation, species propagate rapidly and develop high root density which permits them to absorb water and nutrients from deep horizons during harsh seasons (especially drought periods).

Thus it can be concluded that *P. halepensis* seeds germinate under a wide range of soluble salts and have the ability to tolerate salt stress. GP is less influenced by MgCl₂ followed by CaCl₂ and Na₂SO₄, whereas NaCl prevents germination more than the other soluble salts. Introduction of this coniferous tree has been proposed as a promising strategy to promote restoration in North African saline soils. Further studies are essential to explore these aspects under field conditions.

1. Quezel, P. and Médail, F., *Écologie et biogéographie des forêts du bassin Méditerranéen*, Elsevier SAS, Paris, 2003.
2. Guit, B., Nedjimi, B., Guibal, F. and Chakali, G., Dendroécologie du pin d'Alep (*Pinus halepensis* Mill.) en fonction des paramètres stationnels dans le massif forestier de Senalba (Djelfa – Algérie). *Rev. Ecol.-Terre Vie*, 2015, **70**, 32–43.
3. Halitim, A. (ed.), *Arid Soils in Algeria*, Universities Publications Office, Algiers, Algeria, 1988.
4. Yücel, E., Ecotoxicological effects of different concentrations of alkaline metal salts and an acid on the seed germination of *Pinus nigra* sp. *pallasiana*. *Pak. J. Bot.*, 2008, **40**, 1331–1340.
5. Sidari, M., Mallamaci, C. and Muscolo, A., Drought, salinity and heat differently affect seed germination of *Pinus pinea*. *J. For. Res.*, 2008, **13**, 326–330.
6. Ungar, I. A., Seed germination and seed bank ecology in halophytes. In *Seed Development and Germination* (eds Kigel, J. and Galili, G.), Marcel Dekker, New York, USA, 1995, pp. 599–628.
7. Nedjimi, B., Salinity tolerance: growth, mineral nutrients, and roles of organic osmolytes, case of *Lygeum spartum* L., a review. In *Osmolytes and Plants Acclimation to Changing Environment: Emerging Omics Technologies* (eds Iqbal, N. et al.), Springer, Dordrecht, The Netherlands, 2016, pp. 27–35.
8. Kadik, B. (ed.), *Contribution to Study Aleppo pine (Pinus halepensis Mill) in Algeria: Ecology, Dendrometry, and Morphology*, OPU, Algiers, Algeria, 1987.
9. Pouget, M., Salinity on the quaternary glacia in Algerian calcareous soils. *Bull. Soc. Hist. Nat. Afr. Nord*, 1973, **64**, 15–24.
10. Khan, M. A. and Zia, S., Alleviation of salinity effects by sodium hypochlorite on seed germination of *Limonium stocksii*. *Pak. J. Bot.*, 2007, **39**, 503–511.
11. Nedjimi, B., Mohammedi, N. and Belkheiri, S., Germination responses of medic tree (*Medicago arborea*) seeds to salinity and temperature. *Agric. Res.*, 2014, **3**, 308–312.
12. Shahbazi, A., Nosrati, K. and Egan, T. P., Germination and early seedling growth of two salt-tolerant Atriplex species that prevent erosion in Iranian deserts. In *Sabkha Ecosystems: Volume IV: Cash Crop Halophyte and Biodiversity Conservation* (eds Khan, M. A. et al.), Springer, Dordrecht, The Netherlands, 2014, pp. 273–282.
13. Gul, B., Ansari, R., Flowers, T. J. and Khan, M. A., Germination strategies of halophyte seeds under salinity. *Environ. Exp. Bot.*, 2013, **92**, 4–18.
14. Nasri, S. and Benmahiou, B., Effet de la contrainte saline sur la germination et la croissance de quelques provenances algériennes d'arganier (*Argania spinosa* L.). *Alg. J. Arid Environ.*, 2015, **5**, 98–112.
15. Baskin, C. C. and Baskin, J. M., *Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination*, Academic Press, San Diego, USA, 1998.
16. Le Houérou, H. N., Utilization of fodder trees and shrubs in the arid and semiarid zones of west Asia and North Africa. *Arid Soil Res. Rehab.*, 2000, **14**, 101–135.
17. Croser, C., Renault, S., Franklin, J. and Zwiazek, J., The effect of salinity on the emergence and seedling growth of *Picea mariana*, *Picea glauca* and *Pinus banksiana*. *Environ. Pollut.*, 2001, **115**, 9–16.
18. Khan, M. A. and Gulzar, S., Light, salinity, and temperature effects on the seed germination of perennial grasses. *Am. J. Bot.*, 2003, **90**, 131–134.
19. Khadhri, A., Neffati, M. and Smiti, S., Germination responses of *Cymbopogon schoenanthus* to salinity. *Acta Physiol. Plant.*, 2011, **33**, 279–282.
20. Khan, M. A. and Ungar, I. A., Influence of salinity and temperature on the germination of *Haloxylon recurvum* Bunge ex. Boiss. *Ann. Bot.*, 1996, **78**, 547–551.

ACKNOWLEDGEMENTS. This study was funded by the Ministry of Higher Education and Scientific Research of Algeria, agreement CNEPRU # D04N01UN170120140017. I thank the anonymous reviewer for constructive comments and Z. M. Souissi for technical assistance in the laboratory.

Received 12 January 2017; revised accepted 6 June 2017

doi: 10.18520/cs/v113/i10/2031-2035